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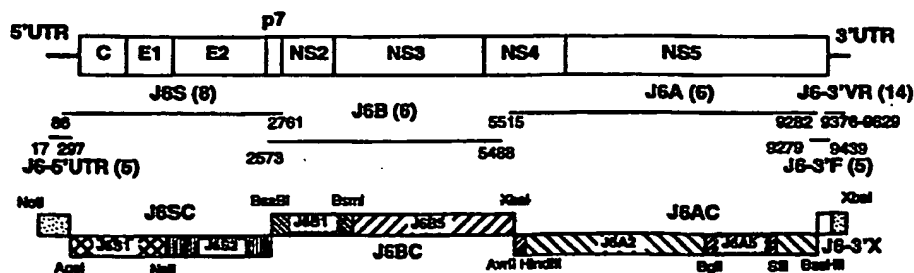
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(54) Title: CLONED GENOME OF INFECTIOUS HEPATITIS C VIRUS OF GENOTYPE 2a AND USES THEREOF



(57) Abstract: The present invention discloses nucleic acid sequence which encodes infectious hepatitis C virus of strain HC-J6_{CH}, genotype 2a, and the use of the sequence, and polypeptides encoded by all or part of the sequence, in the development of vaccines and diagnostics for HCV and in the development of screening assays for the identification of antiviral agents for HCV.

WO 00/75338 A2

-1-

Title Of Invention

Cloned Genome Of Infectious
Hepatitis C Virus of Genotype 2a And Uses Thereof

Field Of Invention

The present invention relates to molecular approaches to the production of nucleic acid sequence which comprises the genome of infectious hepatitis C virus. In particular, the invention provides a nucleic acid sequence which comprises the genome of an infectious hepatitis C virus of genotype 2a. The invention therefore relates to the use of the nucleic acid sequence and polypeptides encoded by all or part of the sequence in the development of vaccines and diagnostic assays for HCV and in the development of screening assays for the identification of antiviral agents for HCV.

Background Of Invention

Hepatitis C virus (HCV) has a positive-sense single-strand RNA genome and is a member of the genus *Hepacivirus* within the *Flaviviridae* family of viruses (Rice, 1996). As for all positive-stranded RNA viruses, the genome of HCV functions as mRNA from which all viral proteins necessary for propagation are translated.

The viral genome of HCV is approximately 9600 nucleotides (nts) in length and consists of a highly conserved 5' untranslated region (UTR), a single long open reading frame (ORF) of approximately 9,000 nts and a complex 3' UTR. The 5' UTR contains an internal ribosomal entry site (Tsukiyama-Kohara et al., 1992;



- 2 -

° Honda et al., 1996). The 3' UTR consists of a short variable region, a polypyrimidine tract of variable length and, at the 3' end, a highly conserved region of approximately 100 nucleotides (Kolykhalov et al., 1996; 5 Tanaka et al., 1995; Tanaka et al., 1996; Yamada et al., 1996). The last 46 nucleotides of this conserved region were predicted to form a stable stem-loop structure thought to be critical for viral replication (Blight and 10 Rice, 1997; Ito and Lai, 1997; Tsuchihara et al., 1997). The ORF encodes a large polypeptide precursor that is cleaved into at least 10 proteins by host and viral proteinases (Rice, 1996). The predicted envelope proteins contain several conserved N-linked 15 glycosylation sites and cysteine residues (Okamoto et al., 1992a). The NS3 gene encodes a serine protease and an RNA helicase and the NS5B gene encodes an RNA-dependent RNA polymerase.

20 A remarkable characteristic of HCV is its genetic heterogeneity, which is manifested throughout the genome (Bukh et al., 1995). The most heterogeneous regions of the genome are found in the envelope genes, in particular the hypervariable region 1 (HVR1) at the 25 N-terminus of E2 (Hijikata et al., 1991; Weiner et al., 1991). HCV circulates as a quasispecies of closely related genomes in an infected individual. Globally, six major HCV genotypes (genotypes 1-6) and multiple 30 subtypes (a, b, c, etc.) have been identified (Bukh et al., 1993; Simmonds et al., 1993).

The nucleotide and deduced amino acid sequences among isolates within a quasispecies generally differ by < 2%, whereas those between isolates of 35 different genotypes vary by as much as 35%. Genotypes

- 3 -

1, 2 and 3 are found worldwide and constitute more than 90% of the HCV infections in North and South America, Europe, Russia, China, Japan and Australia (Forns and Bukh, 1998). Throughout these regions genotype 1 accounts for the majority of HCV infections but genotypes 2 and 3 each account for 5-15%.

At present, more than 80% of individuals infected with HCV become chronically infected and these chronically infected individuals have a relatively high risk of developing chronic hepatitis, liver cirrhosis and hepatocellular carcinoma (Hoofnagle, 1997). The only effective therapy for chronic hepatitis C, interferon (IFN), alone or in combination with ribavirin, induces a sustained response in less than 50% of treated patients (Davis et al., 1998; McHutchinson et al., 1998). Consequently, HCV is currently the most common cause of end stage liver failure and the reason for about 30% of liver transplants performed in the U.S. (Hoofnagle, 1997). In addition, a number of recent studies suggested that the severity of liver disease and the outcome of therapy may be genotype-dependent (reviewed in Bukh et al., 1997). In particular, these studies suggested that infection with HCV genotype 1b was associated with more severe liver disease (Brechot, 1997) and a poorer response to IFN therapy (Fried and Hoofnagle, 1995). As a result of the inability to develop a universally effective therapy against HCV infection, it is estimated that there are still more than 25,000 new infections yearly in the U.S. (Alter 1997) Moreover, since there is no vaccine for HCV, HCV remains a serious public health problem.

- 4 -

Despite the intense interest in the development of vaccines and therapies for HCV, progress has been hindered by the absence of a useful cell culture system and the lack of any small animal model for laboratory study. For example, while replication of HCV in several cell lines has been reported, such observations have turned out not to be highly reproducible. In addition, the chimpanzee is the only animal model, other than man, for this disease. Consequently, HCV has been studied only by using clinical materials obtained from patients or experimentally infected chimpanzees, an animal model whose availability is very limited.

However, several researchers have recently reported the construction of infectious cDNA clones of HCV, the identification of which would permit a more effective search for susceptible cell lines and facilitate molecular analysis of the viral genes and their function. For example, Yoo et al., and Dash et al., (1997) (1995) reported that RNA transcripts from cDNA clones of HCV-1 (genotype 1a) and HCV-N (genotype 1b), respectively, resulted in viral replication after transfection into human hepatoma cell lines. Unfortunately, the viability of these clones was not tested in vivo and concerns were raised about the infectivity of these cDNA clones in vitro (Fausto, 1997). In addition, both clones did not contain the terminal 98 conserved nucleotides at the very 3' end of the UTR.

Kolykhalov et al., (1997) and Yanagi et al. (1997, 1998) reported the derivation from HCV strains H77 (genotype 1a) and HC-J4 (genotype 1b) of cDNA clones

- 5 -

° of HCV that are infectious for chimpanzees. However, while these infectious clones will aid in studying HCV replication and pathogenesis and will provide an important tool for development of in vitro replication and propagation systems, it is important to have
5 infectious clones of more than one genotype, given the extensive genetic heterogeneity of HCV and the potential impact of such heterogeneity on the development of effective therapies and vaccines for HCV.

10 In addition, synthetic chimeric viruses can be used to map the functional regions of viruses with different phenotypes. In flaviviruses and pestiviruses, infectious chimeric viruses have been successfully
15 engineered to express different functional units of related viruses (Bray and Lai, 1991; Pletnev et al., 1992, 1998; Vassilev et al., 1997) and in some cases it has been possible to make chimeras between non-related or distantly related viruses. For instance, the IRES
20 element of poliovirus or bovine viral diarrhoea virus has been replaced with IRES sequences from HCV (Frolov et al., 1998; Lu and Wimmer, 1996; Zhao et al., 1999). Recently, the construction of an infectious chimera of
25 two closely related HCV subtypes has been reported. The chimera contained the complete ORF of a genotype 1b strain but had the 5' and 3' termini of a genotype 1a strain (Yanagi et al., 1998).

30 It is important to determine whether chimeras constructed from more divergent HCV strains are infectious because such chimeras could be used to define the functions of viral units and to dissect the immune
35 response.

- 6 -

Summary Of The Invention

The present invention relates to nucleic acid sequence which comprises the genome of infectious hepatitis C virus and in particular, nucleic acid sequence which comprises the genome of infectious hepatitis C virus of genotype 2a. It is therefore an object of the invention to provide nucleic acid sequence which encodes infectious hepatitis C virus. Such nucleic acid sequence is referred to throughout the application as "infectious nucleic acid sequence".

For the purposes of this application, nucleic acid sequence refers to RNA, DNA, cDNA or any variant thereof capable of directing host organism synthesis of a hepatitis C virus polypeptide. It is understood that nucleic acid sequence encompasses nucleic acid sequences, which due to degeneracy, encode the same polypeptide sequence as the nucleic acid sequences described herein.

The invention also relates to the use of the infectious nucleic acid sequences to produce chimeric genomes consisting of portions of the open reading frames of nucleic acid sequences of other genotypes (including, but not limited to, genotypes 1, 2, 3, 4, 5 and 6) and subtypes (including, but not limited to, subtypes 1a, 1b, 2a, 2b, 2c, 3a, 4a-4f, 5a and 6a) of HCV. For example, infectious nucleic acid sequence of the 2a strain HC-J6, described herein can be used to produce chimeras with sequences from the genomes of other strains of HCV from different genotypes or subtypes. Nucleic acid sequences which comprise sequences from two or more HCV genotypes or subtypes are designated "chimeric nucleic acid sequences".

- 7 -

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The invention further relates to mutations of the infectious nucleic acid sequence of the invention where mutation includes, but is not limited to, point mutations, deletions and insertions. In one embodiment, a gene or fragment thereof can be deleted to determine the effect of the deleted gene or genes on the properties of the encoded virus such as its virulence and its ability to replicate. In an alternative embodiment, a mutation may be introduced into the infectious nucleic acid sequences to examine the effect of the mutation on the properties of the virus.

The invention also relates to the introduction of mutations or deletions into the infectious nucleic acid sequence in order to produce an attenuated hepatitis C virus suitable for vaccine development.

The invention further relates to the use of the infectious nucleic acid sequence to produce attenuated viruses via passage in vitro or in vivo of the viruses produced by transfection of a host cell with the infectious nucleic acid sequence.

The present invention also relates to the use of the nucleic acid sequence of the invention or fragments thereof in the production of polypeptides where "nucleic acid sequence of the invention" refers to infectious nucleic acid sequence, mutations of infectious nucleic acid sequence, chimeric nucleic acid sequence and sequences which comprise the genome of attenuated viruses produced from the infectious nucleic acid sequence of the invention. In one embodiment, said polypeptide or polypeptides are fully or partially purified from hepatitis C virus produced by cells transfected with nucleic acid sequence of the invention.

- 8 -

° In another embodiment, the polypeptide or polypeptides are produced recombinantly from a fragment of the nucleic acid sequences of the invention. In yet another embodiment, the polypeptides are chemically synthesized.

5 The polypeptides of the invention, especially structural polypeptides, can serve as immunogens in the development of vaccines or as antigens in the development of diagnostic assays for detecting the presence of HCV in biological samples.

10 The invention therefore also relates to vaccines for use in immunizing mammals especially humans against hepatitis C. In one embodiment, the vaccine comprises one or more polypeptides made from the nucleic acid sequence of the invention or fragment thereof. In 15 a second embodiment, the vaccine comprises a hepatitis C virus produced by transfection of host cells with the nucleic acid sequences of the invention.

20 The present invention therefore relates to methods for preventing hepatitis C in a mammal. In one embodiment the method comprises administering to a mammal a polypeptide or polypeptides encoded by the nucleic acid sequence of the invention in an amount 25 effective to induce protective immunity to hepatitis C. In another embodiment, the method of prevention comprises administering to a mammal a hepatitis C virus of the invention in an amount effective to induce protective immunity against hepatitis C.

30 In yet another embodiment, the method of protection comprises administering to a mammal the nucleic acid sequence of the invention or a fragment thereof in an amount effective to induce protective 35 immunity against hepatitis C.

- 9 -

° The invention also relates to hepatitis C viruses produced by host cells transfected with the nucleic acid sequence of the present invention.

The invention therefore also provides
5 pharmaceutical compositions comprising the nucleic acid sequence of the invention and/or the encoded hepatitis C viruses. The invention further provides pharmaceutical compositions comprising polypeptides encoded by the
10 nucleic acid sequence of the invention or fragments thereof. The pharmaceutical compositions of the invention may be used prophylactically or therapeutically.

The invention also relates to antibodies to
15 the hepatitis C virus of the invention or their encoded polypeptides and to pharmaceutical compositions comprising these antibodies.

The invention also relates to the use of the
20 nucleic acid sequences of the invention to identify cell lines capable of supporting the replication of HCV in vitro.

The invention further relates to the use of
25 the nucleic acid sequences of the invention or their encoded viral enzymes (e.g. NS3 serine protease, NS3 helicase, NS5B RNA polymerase) to develop screening assays to identify antiviral agents for HCV.

Brief Description Of Figures

30 Figure 1 shows the amplification and cloning of hepatitis C virus genotype 2a (strain HC-J6_{CH}). The nucleotide positions correspond to the sequence of PJ6CF, a full length cDNA clone of hepatitis C virus,
35 genotype 2a, strain HC-J6_{CH}. Products from polymerase

- 10 -

chain reaction are also shown. The names of the clones obtained from these products are indicated (number of clones sequenced are shown in parenthesis). The composition of the full-length cDNA clone is shown at the bottom. The restriction enzymes used for cloning are indicated. An *Xba*I site in HC-J6_{CH} was eliminated by a silent substitution at position 5494.

Figure 2 shows tree analysis of clones amplified from an infectious acute phase plasma pool generated in a chimpanzee inoculated with human plasma containing strain HC-J6 (Okamoto et al., 1991) as well as a tree of the predicted polyprotein sequence of HC-J6_{CH} and the infectious HC-J6_{CH} cDNA clone (pJ6CF). The nucleotide positions with deletions or insertions were stripped in the analysis of the clones. Multiple sequence alignments and tree analyses were performed with GeneWorks (Oxford Molecular Group) (Bukh et al., 1995). Genotype designations are indicated. Other sequences included in the analysis are HC-J8 (Okamoto et al., 1992), genotype 1a infectious clone BEBE1 (Nakao et al., 1996), H77C (Yanagi et al., 1997); genotype 1b infectious clone J4L6S (Yanagi et al., 1998). The scale in each tree indicates the calculated genetic distance.

Figure 3 shows the alignment of the hypervariable region 1 sequences from 8 J6S clones of strain HC-J6_{CH}. HC-J6_{CH} represents the consensus amino acid sequence of the infectious plasma pool from an experimentally infected chimpanzee. HC-J6 is the published amino acid sequence of the original inoculum (Okamoto et al., 1991).

Figure 4 shows the construction of four intertypic chimeric cDNA clones. White boxes are

-11-

sequences derived from genotype 2a clone pJ6CF, and
black boxes are sequences derived from genotype 1a clone
pCV-H77C (Yanagi et al., 1997). An *NdeI* site (mutation
at position 9158 of pCV-H77C) was eliminated and an
artificial *NdeI* site (mutation at position 2765 of
pCV-H77C) was created by site-directed mutagenesis;
silent mutations are underlined.

Figures 5A and 5B show the alignment of the
nucleotide sequences of the 5' (Fig. 5A) and 3' UTRs
(Fig. 5B) and the amino acid sequences of E2/p7/NS2
junctions (Fig. 5B) in the intertypic 1a, 2a chimeric
cDNA clones. In the 5' UTR alignment, the first 39 nts
of core believed to be important for the IRES function
were included (Lemon and Honda, 1997). Top line: the
sequence of the infectious genotype 1a clone pCV-H77C
(Yanagi et al., 1997). Bottom line: the sequence of the
infectious genotype 2a clone pJ6CF. Dot: identity with
the sequence of H77C. Capital letter: different from the
sequence of H77C. Dash: deletion. Bold face: initiation
or stop codon of the ORF. Underlined: *AgeI* cleavage
site. Arrow: putative sites in the HCV polyprotein
cleaved by host signal peptidases. Numbering
corresponds to the sequence of pCV-H77C.

Figures 6A-6F show the nucleotide sequence of
the infectious hepatitis C virus clone of genotype 1a
strain H77C and Figures 6G-6H show the amino acid
sequence encoded by the clone.

Figures 7A-7F show the nucleotide sequence of
the infectious hepatitis C virus clone of genotype 1b
strain HC-J4 and Figures 7G-H show the amino acid
sequence encoded by the clone.

- 12 -

DESCRIPTION OF THE INVENTION

The present invention relates to nucleic acid sequence which comprises the genome of an infectious hepatitis C virus. More specifically, the invention relates to nucleic acid sequence which encodes infectious hepatitis C virus of strain HC-J6_{CH}, genotype 2a. The infectious nucleic acid sequence of the invention is shown in SEQ ID NO:1 and is contained in a plasmid construct deposited with the American Type Culture Collection (ATCC) on May 28, 1999 and having ATCC accession number PTA-153.

The invention also relates to "chimeric nucleic acid sequences" where the chimeric nucleic acid sequences consist of open-reading frame sequences and/or 5' and/or 3' untranslated sequences taken from nucleic acid sequences of hepatitis C viruses of different genotypes or subtypes.

In one embodiment, the chimeric nucleic acid sequence consists of sequence from the genome of infectious HCV of genotype 2a which encodes structural polypeptides and sequence from the genome of a HCV of a different genotype or subtype which encodes nonstructural polypeptides.

Alternatively, the nonstructural region of infectious HCV of genotype 2a and structural region of a HCV of a different genotype or subtype may be combined. This will result in a chimeric nucleic acid sequence consisting of sequence from the genome of infectious HCV of genotype 2a which encodes nonstructural polypeptides and sequence from the genome of a HCV of a another genotype or subtype which encodes structural polypeptides.

- 13 -

°
Preferably, the nucleic acid sequence from the genome of the infectious HCV clone of genotype 1a (deposited with the ATCC on June 2, 1999 ; Figures 6A-6F), or the nucleic acid sequence from the genome of the infectious HCV clone of genotype 1b (ATCC accession number 209596; Figures 7A-7F) is used to construct the chimeric nucleic acid sequence with the HCV of genotype 2a of the invention.

10 It is believed that the construction of such chimeric nucleic acid sequences will be of importance in studying the growth and virulence properties of hepatitis C virus and in the production of candidate hepatitis C virus vaccines suitable to confer protection against multiple genotypes of HCV. For example, one might produce a "multivalent" vaccine by putting epitopes from several genotypes or subtypes into one clone. Alternatively one might replace just a single gene from an infectious sequence with the corresponding gene from the genomic sequence of a strain from another genotype or subtype or create a chimeric gene which contains portions of a gene from two genotypes or subtypes. Examples of genes which could be replaced or which could be made chimeric, include, but are not limited to, the E1, E2 and NS4 genes.

20 The invention further relates to mutations of the infectious nucleic acid sequences where "mutations" include, but are not limited to, point mutations, deletions and insertions. Of course, one of ordinary skill in the art would recognize that the size of the insertions would be limited by the ability of the resultant nucleic acid sequence to be properly packaged within the virion. Such mutations could be produced by

- 14 -

- ° techniques known to those of skill in the art such as site-directed mutagenesis, fusion PCR, and restriction digestion followed by religation.

In one embodiment, mutagenesis might be undertaken to determine sequences that are important for viral properties such as replication or virulence. For example, one may introduce a mutation into the infectious nucleic acid sequence which eliminates the cleavage site between the NS4A and NS4B polypeptides to examine the effects on viral replication and processing of the polypeptide.

Alternatively, one may delete all or part of a gene or of the 5' or 3' nontranslated region contained in an infectious nucleic acid sequence and then transfect a host cell (animal or cell culture) with the mutated sequence and measure viral replication in the host by methods known in the art such as RT-PCR. Preferred genes include, but are not limited to, the P7, NS4B and NS5A genes. Of course, those of ordinary skill in the art will understand that deletion of part of a gene, preferably the central portion of the gene, may be preferable to deletion of the entire gene in order to conserve the cleavage site boundaries which exist between proteins in the HCV polyprotein and which are necessary for proper processing of the polyprotein.

In the alternative, if the transfection is into a host animal such as a chimpanzee, one can monitor the virulence phenotype of the virus produced by transfection of the mutated infectious nucleic acid sequence by methods known in the art such as measurement of liver enzyme levels (alanine aminotransferase (ALT) or isocitrate dehydrogenase (ICD)) or by histopathology

- 15 -

° of liver biopsies. Thus, mutations of the infectious nucleic acid sequences may be useful in the production of attenuated HCV strains suitable for vaccine use.

The invention also relates to the use of the
5 infectious nucleic acid sequence of the present invention to produce attenuated viral strains via passage in vitro or in vivo of the virus produced by transfection with the infectious nucleic acid sequence.

10 The present invention therefore relates to the use of the nucleic acid sequence of the invention to identify cell lines capable of supporting the replication of HCV.

In particular, it is contemplated that the
15 mutations of the infectious nucleic acid sequence of the invention and the production of chimeric sequences as discussed above may be useful in identifying sequences critical for cell culture adaptation of HCV and hence, may be useful in identifying cell lines capable of
20 supporting HCV replication.

Transfection of tissue culture cells with the nucleic acid sequences of the invention may be done by methods of transfection known in the art such as
25 electroporation, precipitation with DEAE-Dextran or calcium phosphate or liposomes.

In one such embodiment, the method comprises the growing of animal cells, especially human cells, in vitro and transfecting the cells with the nucleic acid
30 of the invention, then determining if the cells show indicia of HCV infection. Such indicia include the detection of viral antigens in the cell, for example, by immunofluorescence procedures well known in the art; the
35 detection of viral polypeptides by Western blotting

- 16 -

° using antibodies specific therefor; and the detection of newly transcribed viral RNA within the cells via methods such as RT-PCR. The presence of live, infectious virus particles following such tests may also be shown by
5 injection of cell culture medium or cell lysates into healthy, susceptible animals, with subsequent exhibition of the signs and symptoms of HCV infection.

Suitable cells or cell lines for culturing HCV include, but are not limited to, lymphocyte and
10 hepatocyte cell lines known in the art.

Alternatively, primary hepatocytes can be cultured, and then infected with HCV; or, the hepatocyte cultures could be derived from the livers of infected
15 chimpanzees. In addition, various immortalization methods known to those of ordinary skill in the art can be used to obtain cell lines derived from hepatocyte cultures. For example, primary hepatocyte cultures may be fused to a variety of cells to maintain stability.
20

The present invention further relates to the in vitro and in vivo production of hepatitis C viruses from the nucleic acid sequences of the invention.

In one embodiment, the sequences of the
25 invention can be inserted into an expression vector that functions in eukaryotic cells. Eukaryotic expression vectors suitable for producing high efficiency gene transfer in vivo are well known to those of ordinary skill in the art and include, but are not limited to,
30 plasmids, vaccinia viruses, retroviruses, adenoviruses and adeno-associated viruses.

In another embodiment, the sequences contained in the recombinant expression vector can be transcribed
35 in vitro by methods known to those of ordinary skill in

- 17 -

° the art in order to produce RNA transcripts which encode the hepatitis C viruses of the invention. The hepatitis C viruses of the invention may then be produced by transfecting cells by methods known to those of ordinary skill in the art with either the in vitro transcription mixture containing the RNA transcripts or with the recombinant expression vectors containing the nucleic acid sequences described herein.

10 The hepatitis C viruses produced from the sequences of the invention may be purified or partially purified from the transfected cells by methods known to those of ordinary skill in the art. In a preferred embodiment, the viruses are partially purified prior to their use as immunogens in the pharmaceutical compositions and vaccines of the present invention.

20 The present invention therefore relates to the use of the hepatitis C viruses produced from the nucleic acid sequences of the invention as immunogens in live or killed (e.g., formalin inactivated) vaccines to prevent hepatitis C in a mammal.

25 In an alternative embodiment, the immunogen of the present invention may be an infectious nucleic acid sequence, a chimeric nucleic acid sequence, or a mutated infectious nucleic acid sequence which encodes a hepatitis C virus. Where the sequence is a cDNA sequence, the cDNAs and their RNA transcripts may be used to transfect a mammal by direct injection into the liver tissue of the mammal as described in the Examples.

30 Alternatively, direct gene transfer may be accomplished via administration of a eukaryotic expression vector containing a nucleic acid sequence of the invention.

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- 18 -

° In yet another embodiment, the immunogen may be a polypeptide encoded by the nucleic acid sequences of the invention. The present invention therefore also relates to polypeptides produced from the nucleic acid sequences of the invention or fragments thereof. In one 5 embodiment, polypeptides of the present invention can be recombinantly produced by synthesis from the nucleic acid sequences of the invention or isolated fragments thereof, and purified, or partially purified, from 10 transfected cells using methods already known in the art. In an alternative embodiment, the polypeptides may be purified or partially purified from viral particles produced via transfection of a host cell with the 15 nucleic acid sequences of the invention. Such polypeptides might, for example, include either capsid or envelope polypeptides prepared from the sequences of the present invention.

20 When used as immunogens, the nucleic acid sequences of the invention, or the polypeptides or viruses produced therefrom, are preferably partially purified prior to use as immunogens in pharmaceutical compositions and vaccines of the present invention. 25 When used as a vaccine, the sequences and the polypeptide and virus products thereof, can be administered alone or in a suitable diluent, including, but not limited to, water, saline, or some type of 30 buffered medium. The vaccine according to the present invention may be administered to an animal, especially a mammal, and most especially a human, by a variety of routes, including, but not limited to, intradermally, intramuscularly, subcutaneously, or in any combination 35 thereof.

- 19 -

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Suitable amounts of material to administer for prophylactic and therapeutic purposes will vary depending on the route selected and the immunogen (nucleic acid, virus, polypeptide) administered. One
5 skilled in the art will appreciate that the amounts to be administered for any particular treatment protocol can be readily determined without undue experimentation. The vaccines of the present invention may be
10 administered once or periodically until a suitable titer of anti-HCV antibodies appear in the blood. For an immunogen consisting of a nucleic acid sequence, a suitable amount of nucleic acid sequence to be used for prophylactic purposes might be expected to fall in the
15 range of from about 100 μ g to about 5 mg and most preferably in the range of from about 500 μ g to about 2mg. For a polypeptide, a suitable amount to use for prophylactic purposes is preferably 100 ng to 100 μ g and
20 for a virus 10^2 to 10^6 infectious doses. Such administration will, of course, occur prior to any sign of HCV infection.

A vaccine of the present invention may be employed in such forms as capsules, liquid solutions,
25 suspensions or elixirs for oral administration, or sterile liquid forms such as solutions or suspensions. An inert carrier is preferably used, such as saline or phosphate-buffered saline, or any such carrier in which
30 the HCV of the present invention can be suitably suspended. The vaccines may be in the form of single dose preparations or in multi-dose flasks which can be utilized for mass-vaccination programs of both animals
35 and humans. For purposes of using the vaccines of the present invention reference is made to Remington's

- 20 -

° Pharmaceutical Sciences, Mack Publishing Co., Easton, Pa., Osol (Ed.) (1980); and New Trends and Developments in Vaccines, Voller et al. (Eds.), University Park Press, Baltimore, Md. (1978), both of which provide much
5 useful information for preparing and using vaccines. Of course, the polypeptides of the present invention, when used as vaccines, can include, as part of the composition or emulsion, a suitable adjuvant, such as
10 alum (or aluminum hydroxide) when humans are to be vaccinated, to further stimulate production of antibodies by immune cells. When nucleic acids, viruses or polypeptides are used for vaccination purposes, other specific adjuvants such as CpG motifs (Krieg, A.K. et
15 al. (1995) and (1996)), may prove useful.

When the nucleic acids, viruses and polypeptides of the present invention are used as vaccines or inocula, they will normally exist as
20 physically discrete units suitable as a unitary dosage for animals, especially mammals, and most especially humans, wherein each unit will contain a predetermined quantity of active material calculated to produce the desired immunogenic effect in association with the
25 required diluent. The dose of said vaccine or inoculum according to the present invention is administered at least once. In order to increase the antibody level, a second or booster dose may be administered at some time after the initial dose. The need for, and timing of,
30 such booster dose will, of course, be determined within the sound judgment of the administrator of such vaccine or inoculum and according to sound principles well known in the art. For example, such booster dose could
35 reasonably be expected to be advantageous at some time

- 21 -

- ° between about 2 weeks to about 6 months following the initial vaccination. Subsequent doses may be administered as indicated.

The nucleic acid sequences, viruses and polypeptides of the present invention can also be administered for purposes of therapy, where a mammal, especially a primate, and most especially a human, is already infected, as shown by well known diagnostic measures. When the nucleic acid sequences, viruses or polypeptides of the present invention are used for such therapeutic purposes, much of the same criteria will apply as when it is used as a vaccine, except that inoculation will occur post-infection. Thus, when the nucleic acid sequences, viruses or polypeptides of the present invention are used as therapeutic agents in the treatment of infection, the therapeutic agent comprises a pharmaceutical composition containing a sufficient amount of said nucleic acid sequences, viruses or polypeptides so as to elicit a therapeutically effective response in the organism to be treated. Of course, the amount of pharmaceutical composition to be administered will, as for vaccines, vary depending on the immunogen contained therein (nucleic acid, polypeptide, virus) and on the route of administration.

The therapeutic agent according to the present invention can thus be administered by subcutaneous, intramuscular or intradermal routes. One skilled in the art will certainly appreciate that the amounts to be administered for any particular treatment protocol can be readily determined without undue experimentation. Of course, the actual amounts will vary depending on the route of administration as well as the sex, age, and

- 22 -

- ° clinical status of the subject which, in the case of human patients, is to be determined with the sound judgment of the clinician.

The therapeutic agent of the present invention can be employed in such forms as capsules, liquid solutions, suspensions or elixirs, or sterile liquid forms such as solutions or suspensions. An inert carrier is preferably used, such as saline, phosphate-buffered saline, or any such carrier in which the HCV of the present invention can be suitably suspended. The therapeutic agents may be in the form of single dose preparations or in the multi-dose flasks which can be utilized for mass-treatment programs of both animals and humans. Of course, when the nucleic acid sequences, viruses or polypeptides of the present invention are used as therapeutic agents they may be administered as a single dose or as a series of doses, depending on the situation as determined by the person conducting the treatment.

The nucleic acids, polypeptides and viruses of the present invention can also be utilized in the production of antibodies against HCV. The term "antibody" is herein used to refer to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules. Examples of antibody molecules are intact immunoglobulin molecules, substantially intact immunoglobulin molecules and portions of an immunoglobulin molecule, including those portions known in the art as Fab, F(ab')₂ and F(v) as well as chimeric antibody molecules.

Thus, the polypeptides, viruses and nucleic acid sequences of the present invention can be used in

- 23 -

° the generation of antibodies that immunoreact (i.e., specific binding between an antigenic determinant-containing molecule and a molecule containing an antibody combining site such as a whole antibody molecule or an active portion thereof) with antigenic determinants on the surface of hepatitis C virus particles.

10 The present invention therefore also relates to antibodies produced following immunization with the nucleic acid sequences, viruses or polypeptides of the present invention. These antibodies are typically produced by immunizing a mammal with an immunogen or vaccine to induce antibody molecules having

15 immunospecificity for polypeptides or viruses produced in response to infection with the nucleic acid sequences of the present invention. When used in generating such antibodies, the nucleic acid sequences, viruses, or

20 polypeptides of the present invention may be linked to some type of carrier molecule. The resulting antibody molecules are then collected from said mammal. Antibodies produced according to the present invention have the unique advantage of being generated in response

25 to authentic, functional polypeptides produced according to the actual cloned HCV genome.

The antibody molecules of the present invention may be polyclonal or monoclonal. Monoclonal antibodies are readily produced by methods well known in

30 the art. Portions of immunoglobulin molecules, such as Fabs, as well as chimeric antibodies, may also be produced by methods well known to those of ordinary skill in the art of generating such antibodies.

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The antibodies according to the present invention may also be contained in blood, plasma, serum, hybridoma supernatants, and the like. Alternatively, the antibody of the present invention is isolated to the extent desired by well known techniques such as, for example, using DEAE Sephadex. The antibodies produced according to the present invention may be further purified so as to obtain specific classes or subclasses of antibody such as IgM, IgG, IgA, and the like. Antibodies of the IgG class are preferred for purposes of passive protection.

The antibodies of the present invention are useful in the prevention and treatment of diseases caused by hepatitis C virus in animals, especially mammals, and most especially humans.

In providing the antibodies of the present invention to a recipient mammal, preferably a human, the dosage of administered antibodies will vary depending on such factors as the mammal's age, weight, height, sex, general medical condition, previous medical history, and the like.

In general, it will be advantageous to provide the recipient mammal with a dosage of antibodies in the range of from about 1 mg/kg body weight to about 10 mg/kg body weight of the mammal, although a lower or higher dose may be administered if found desirable. Such antibodies will normally be administered by intravenous or intramuscular route as an inoculum. The antibodies of the present invention are intended to be provided to the recipient subject in an amount sufficient to prevent, lessen or attenuate the severity, extent or duration of any existing infection.

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° The antibodies prepared by use of the nucleic acid sequences, viruses or polypeptides of the present invention are also highly useful for diagnostic purposes. For example, the antibodies can be used as in
5 vitro diagnostic agents to test for the presence of HCV in biological samples taken from animals, especially humans. Such assays include, but are not limited to, radioimmunoassays, EIA, fluorescence, Western blot analysis and ELISAs. In one such embodiment, the
10 biological sample is contacted with antibodies of the present invention and a labeled second antibody is used to detect the presence of HCV to which the antibodies are bound.

15 Such assays may be, for example, direct where the labeled first antibody is immunoreactive with the antigen, such as, for example, a polypeptide on the surface of the virus; indirect where a labeled second
20 antibody is reactive with the first antibody; a competitive protocol such as would involve the addition of a labeled antigen; or sandwich where both labeled and unlabeled antibody are used, as well as other protocols well known and described in the art.

25 In one embodiment, an immunoassay method would utilize an antibody specific for HCV envelope determinants and would further comprise the steps of contacting a biological sample with the HCV-specific
30 antibody and then detecting the presence of HCV material in the test sample using one of the types of assay protocols as described above. Polypeptides and antibodies produced according to the present invention may also be supplied in the form of a kit, either
35 present in vials as purified material, or present in

- 26 -

° compositions and suspended in suitable diluents as previously described.

In a preferred embodiment, such a diagnostic test kit for detection of HCV antigens in a test sample
5 comprises in combination a series of containers, each container a reagent needed for such assay. Thus, one such container would contain a specific amount of HCV-specific antibody as already described, a second
10 container would contain a diluent for suspension of the sample to be tested, a third container would contain a positive control and an additional container would contain a negative control. An additional container could contain a blank.

15 For all prophylactic, therapeutic and diagnostic uses, the antibodies of the invention and other reagents, plus appropriate devices and accessories, may be provided in the form of a kit so as to facilitate ready availability and ease of use.

20 The present invention also relates to the use of nucleic acid sequences and polypeptides of the present invention to screen potential antiviral agents for antiviral activity against HCV. Such screening
25 methods are known by those of skill in the art. Generally, the antiviral agents are tested at a variety of concentrations, for their effect on preventing viral replication in cell culture systems which support viral
30 replication, and then for an inhibition of infectivity or of viral pathogenicity (and a low level of toxicity) in an animal model system.

In one embodiment, animal cells (especially human cells) transfected with the nucleic acid sequences
35 of the invention are cultured in vitro and the cells are

- 27 -

° treated with a candidate antiviral agent (a chemical, peptide etc.) by adding the candidate agent to the medium. The treated cells are then exposed, possibly under transfecting or fusing conditions known in the art, to the nucleic acid sequences of the present invention. A sufficient period of time would then be allowed to pass for infection to occur, following which the presence or absence of viral replication would be determined versus untreated control cells by methods known to those of ordinary skill in the art. Such methods include, but are not limited to, the detection of viral antigens in the cell, for example, by immunofluorescence procedures well known in the art; the detection of viral polypeptides by Western blotting using antibodies specific therefor; the detection of newly transcribed viral RNA within the cells by RT-PCR; and the detection of the presence of live, infectious virus particles by injection of cell culture medium or cell lysates into healthy, susceptible animals, with subsequent exhibition of the signs and symptoms of HCV infection. A comparison of results obtained for control cells (treated only with nucleic acid sequence) with those obtained for treated cells (nucleic acid sequence and antiviral agent) would indicate, the degree, if any, of antiviral activity of the candidate antiviral agent. Of course, one of ordinary skill in the art would readily understand that such cells can be treated with the candidate antiviral agent either before or after exposure to the nucleic acid sequence of the present invention so as to determine what stage, or stages, of viral infection and replication said agent is effective against.

- 28 -

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In an alternative embodiment, viral enzyme such as NS3 protease, NS2-NS3 protease, NS3 helicase or NS5B RNA polymerase may be produced from a nucleic acid sequence of the invention and used to screen for inhibitors which may act as antiviral agents. The structural and nonstructural regions of the HCV genome, including nucleotide and amino acid locations, have been determined, for example, as depicted in Houghton, M. (1996), Fig. 1; and Major, M.E. et al. (1997), Table 2.

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Such above-mentioned protease inhibitors may take the form of chemical compounds or peptides which mimic the known cleavage sites of the protease and may be screened using methods known to those of skill in the art (Houghton, M. (1996) and Major, M.E. et al. (1997)). For example, a substrate may be employed which mimics the protease's natural substrate, but which provides a detectable signal (e.g. by fluorimetric or colorimetric methods) when cleaved. This substrate is then incubated with the protease and the candidate protease inhibitor under conditions of suitable pH, temperature etc. to detect protease activity. The proteolytic activities of the protease in the presence or absence of the candidate inhibitor are then determined.

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In yet another embodiment, a candidate antiviral agent (such as a protease inhibitor) may be directly assayed in vivo for antiviral activity by administering the candidate antiviral agent to a chimpanzee transfected with a nucleic acid sequence of the invention or infected with a virus of the invention and then measuring viral replication in vivo via methods such as RT-PCR. Of course, the chimpanzee may be treated with the candidate agent either before or after

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- 29 -

transfection with the infectious nucleic acid sequence or infected with a virus of the invention so as to determine what stage, or stages, of viral infection and replication the agent is effective against.

The invention also provides that the nucleic acid sequences, viruses and polypeptides of the invention may be supplied in the form of a kit, alone or in the form of a pharmaceutical composition.

All scientific publication and/or patents cited herein are specifically incorporated by reference. The following examples illustrate various aspects of the invention but are in no way intended to limit the scope thereof.

15

EXAMPLES

Materials and Methods

Source of HCV

An infectious plasma pool of HCV genotype 2a (HC-J6_{CH}) prepared from acute phase plasma of a chimpanzee experimentally inoculated with plasma from a Japanese patient infected with strain HC-J6 (Okamoto et al., 1991) was used for cloning. An infectious cDNA clone of HCV strain H77, genotype 1a was also used (pCV-H77C; Yanagi et al., 1997).

25

Amplification, cloning and sequence analysis

Viral RNA was extracted from 100 µl aliquots of the HC-J6_{CH} plasma pool with the TRIzol system (GIBCO/BRL) (Yanagi et al., 1997). Primers used in cDNA synthesis and PCR amplification were based on the genomic sequence of strain HC-J6 (Okamoto et al., 1991) and from the conserved region (3'X) of the 3' UTR of HCV genotype 2a (Tanaka et al., 1996) (Table 1). The RNA

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was denatured at 65°C for 2 min, and cDNA was synthesized at 42°C for 1 hour with Superscript II reverse transcriptase (GIBCO/BRL) and specific reverse primers in 20 µl reaction volumes. The cDNA mixtures were treated with RNase H and RNase T1 (GIBCO/BRL) at 37°C for 20 min.

TABLE 1

Oligonucleotides used for amplification and cloning of strain HC-J6_{CH}, genotype 2a

Designation	Sequence (5' → 3') ^a
2427S-H77	ACTGGACACGGAGGTGGCCGCGTC
2426S-H77	TTGTTCTTGTCGGGTTAATGGCGC
2645R-H77	GGGTGTACTACACACATGAGTAAG
2832R-H77	AAGCGCCCTAACTGATGATG
H2751SII	CGTCTCTAGAC ACCTCAGCGGGCATATGCACTGGACACGGA
H2786R	GTCCAGTGCATATGCCCGCTGAGG
H2870R	CATGCACCAGCTGATATAGCGCTTGTAATATG
H7851S	TCCGTAGAGGAAGCTTGACGCTGACGCC
H9140S (M)	CAGAGGAGGCAGGGTGCTATATGTGGCAAGTAC
H9173R (M)	GTACTTGCCACATATAGCAGCCCTGCCTCCTCTG
H9471R	CGTCTCTAGAC CAGGAAATGGCTTAAGAGCGCGAGTGTTTACC
J6-H2556S	TTATGGATGCTCATCTTGTGGGCCAGGCCGAAGCAGCTTTGGAGAACCTCGTAATACTCAATGC
356RF-J6H	AGGATTTGTGCTCATGGTGACGGTCTACGAG
1S-J6F ^b	TTTTTTTTGGCGCGCTAATACGACTCACTATAG ACCCGCCCTAATAGG
333S-J6	CCGTGCACCATGAGCACAAATCCTAAACCTC
753R-J6	GGATGTACCCCATGAGGTGGCAAAG
2543S-J6F	GTTTGGCGCTGCTTATGGATGCTCATCTTG
2787R-J6 (26)	GCGTCATAAGCATATGCCTGTTGGGG
3329R-J6	CCCTCAGCACTGGAGTACATCTG
5487-J6F	CGTCAATGCATA CCCTAGGGCGGCTCTCATTGAAGAGGG
5518R-J6F	CGTCCCTCTTCAATGAGAGCCGCTCTAGA
9251S-J6F	GCGGTGAAGACCAAGCTCAAACCTCACTC
9305R-J6F	AATCTAGA AGGCGGCTTCCGGCAATGGAGTGAGTTTGAGC
9310R-J6F	CGTCTCTAGAGG ATAATCCAGGAGGCGCGCTTCCGGC
9399S-J6F	TACTTTTTGTAGGGGTAGGCCTTTTCC
9464-J6F	CGTCTCTAGAGT GTAGCTAATGTGTGCCGCTCTA
9470 (24)-J6	CTATGGAGTGTAGCTAATGTGTGC
J6-3' XR	CGTCTCTAGAC ATGATCTGCAGAGAGACCAGTTACGGCACTCTCTGFCAGTCATGCGGC
	TCACGGACCTTTACAGCTAGCCGTGACTAGGGCTAAGATGGAGCCACC

^a HCV-specific sequences are shown in plain text, non HCV-specific sequences are shown in bold face, and cleavage sites used for cDNA cloning are underlined.

^b The core sequence of the T7 promotor is shown in italics.

The strategy used to amplify and clone the full-length HC-J6_{CH} sequence is shown in Fig. 1.

Nucleotide positions correspond to those of the 2a

- 31 -

° infectious clone (pJ6CF) that is described herein. The 5' end of HC-J6_{CH} (nts. 17-297, excluding primer sequences) was amplified from 2 µl of cDNA synthesized with primer a-2 (Yanagi et al., 1996). PCR was performed with *AmpliTaq Gold* DNA polymerase (Perkin-Elmer) as described previously (Yanagi et al., 1996) using primers 1S-J6F and a-2. After purification, the amplified products were cloned into pGEM-T Easy vector (Promega) using standard procedures and 5 clones (pJ6-5'UTR) were sequenced.

The 3' end of HC-J6_{CH} was amplified in 3 overlapping pieces. RT-PCR of a short fragment of NS5B (nts. 9279-9439) was performed with primers 9251S-J6F and 9464R-J6F as described above. The PCR products were cloned into pGEM-T Easy vector and sequence analysis was performed from 5 pJ6-3'F clones. A second region spanning from NS5B to the conserved region of the 3' UTR (nts. 9376-9629) was amplified in RT-nested PCR (external primers H9261F and H3'X58R, internal primers H9282F and H3'X45R) (Yanagi et al., 1997). The amplified products were cloned into pGEM-9zf(-) by using *HindIII* and *XbaI* sites and 14 pJ6-3'VR clones were sequenced. The third fragment, which included the 3' terminal sequence was amplified with primers 9399S-J6F and J6-3'XR from one of the pJ6-3'VR clones, and cloned into one of the pJ6-3'F clones by using *StuI* and *XbaI* sites (pJ6-3'X).

The ORF of HCV HC-J6_{CH} was amplified by long RT-PCR in 3 overlapping pieces. The amplification was performed on 2 µl of the cDNA mixtures with the Advantage cDNA polymerase mix (Clontech) (Yanagi et al., 1997). The J6S fragment (nts. 86-2761) was amplified

- 32 -

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with primers a-1 (Yanagi et al., 1996) and J6-2787R from
cDNA synthesized with primer J6-3329R. A single PCR
round was performed in a Robocycler thermal cycler
(Stratagene), and consisted of denaturation at 99°C for
5 35 sec, annealing at 67°C for 30 sec and elongation at
68°C for 4 min 30 sec during the first 5 cycles, 5 min
during the next 10 cycles, 5 min 30 sec during the
following 10 cycles and 6 min during the last 10 cycles.
10 The J6B fragment (nts. 2573-5488) was amplified with
primers 2543S-J6F and 5518R-J6F from cDNA synthesized
with primer 5518R-J6F. Finally, the J6A fragment (nts.
5515-9282) was amplified with primers 5487S-J6F and
9310R-J6F from cDNA synthesized with primer
15 9470R(24)-J6F. PCR amplifications of fragments J6B and
J6A consisted of denaturation at 99°C for 35 sec,
annealing at 67°C for 30 sec and elongation at 68°C for 6
min during the first 5 cycles, 7 min during the next 10
20 cycles, 8 min during the following 10 cycles and 9 min
during the last 10 cycles.

After purification of the long PCR products
with QIAquick PCR purification kit (QIAGEN), A-tailing
reactions were performed with *AmpliTaq* DNA polymerase
25 (Perkin Elmer) at 72 °C for 1 hour. The gel-purified
A-tailed PCR products were cloned into pCR2.1 vector
(Invitrogen) or pGEM-T Easy vector (Promega). DH5-alpha
competent cells (GIBCO BRL) were transformed and
30 selected on LB agar plates containing 100 µg/ml
ampicillin (SIGMA) and amplified in LB liquid cultures
at 30°C for 18 - 20 hrs (Yanagi et al., 1997). Midiprep
was performed using Wizard *Plus* Midipreps DNA
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-33-

Purification System (Promega). Multiple clones of the J6S, J6A and the J6B fragments were sequenced.

The consensus sequence of strain HC-J6_{CH} (nts. 17-9629) was determined by direct sequencing of PCR products (nts. 297-3004 and nts. 4893-5762) and by sequence analysis of the TA clones (nts. 17-5488 and nts. 5515-9629) (Fig. 1). Both strands of DNA were sequenced in all cases. Analyses of genomic sequences, including multiple sequence alignments and tree analyses, were performed with GeneWorks (Oxford Molecular Group) (Bukh et al., 1995).

Construction of chimeric cDNA clones of genotypes 1a & 2a

Four full-length intertypic chimeric cDNA clones were constructed (Figs. 4, 5A, 5B). In each clone the C, E1 and E2 genes encoded the consensus amino acid sequence of HC-J6_{CH}. The p7 protein was encoded either by the HC-J6_{CH} or pCV-H77C consensus sequence, and the NS proteins were all encoded by pCV-H77C genes. To engineer these cDNA clones, an *NdeI* site from pCV-H77C was first eliminated by a silent substitution (C to T) at position 9158. In brief, two fragments were amplified from pCV-H77C with primers H7851S and H9173R(M) and with primers H9140S(M) and H9417R (Table 3), gel-purified and used for fusion PCR with primers H7851S and H9417R. The fusion PCR products were cloned into pCV-H77C by using *HindIII* and *AflIII* sites. A new artificial *NdeI* site was introduced by a silent substitution (C to T) at position 2765. PCR products, which were amplified from pCV-H77C with primer H2751SII containing artificial *ClaI* and *NdeI* sites and primer H2870R, were cloned into the modified pCV-H77C by using

- 34 -

- *Cla*I and *Eco*47III sites. The final construct (pH77CV) was used as a cassette vector to construct the intertypic chimeric HCV cDNA clones.

The four chimeric cDNA clones were constructed as follows. pH77CV-J6S (nucleotide sequence shown in SEQ ID No:3 and amino acid sequence shown in SEQ ID No:4): The *Age*I/*Bsm*I fragment of clone J6S2 and the *Bsm*I/*Nde*I fragment of clone J6S1, were cloned into pH77CV by using *Age*I and *Nde*I sites; pH77 (p7)CV-J6S (nucleotide sequence shown in SEQ ID No:5 and amino acid sequence shown in SEQ ID No:6): A fragment of pH77CV-J6S was replaced with a fragment amplified from pCV-H77C with primers J6-H2556S and H2786R by using *Bsa*BI and *Nde*I sites; J6S (nucleotide sequence shown in SEQ ID No:7 and amino acid sequence shown in SEQ ID No:8): A fragment amplified from pH77pCV-H77C with primers a-1 and 356RF-J6H77 and another fragment amplified from pH77CV-J6S with primers 333S-J6 and 753R-J6 were gel-purified and a fusion-PCR was performed with primers a-1 and 753R-J6. The *Age*I/*Cla*I fragment of the subcloned fusion PCR products and the *Cla*I/*Nde*I fragment of pH77CV-J6S were cloned into pH77CV-J6S by using *Age*I and *Nde*I sites; pH77(p7)-J6S (nucleotide sequence shown in SEQ ID No:9 and amino acid sequence shown in SEQ ID No:10): The *Age*I/*Cla*I fragment of J6S and the *Cla*I/*Nde*I fragment of (p7)CV-J6S were cloned into pH77(p7)CV-J6S by using *Age*I and *Nde*I sites.

Each intertypic chimeric cDNA clone was retransformed to select a single clone, and large-scale preparation of plasmid DNA was performed with a QIAGEN plasmid Maxi kit as described previously (Yanagi et al., 1997). Each of the four cDNA clones was completely

- 35 -

° sequenced before inoculation. Each clone was genetically stable since the digestion pattern was as expected following retransformation and the complete sequence was the expected one.

5 Construction of full-length cDNA clone HC-J6_{CH}

An overview of the full-length HC-J6_{CH} clone is presented in Fig. 1. In the final construct pJ6CF, which encodes the consensus polyprotein of HC-J6_{CH}, an
10 *Xba*I site was eliminated by a silent substitution (A to G) at position 5494. Digested fragments containing the consensus sequence were purified from the appropriate subclones and ligated using the sites indicated. The full-length cDNA clone (pJ6CF) was retransformed to
15 select a single clone, and large-scale preparation of plasmid DNA followed by the complete sequence analysis was performed. Clone pJ6CF was genetically stable.

20 Intrahepatic transfection of chimpanzee with transcribed RNA

In duplicate 100 µl reactions, RNA was transcribed *in vitro* with T7 RNA polymerase (Promega) from 10 µg of template plasmid linearized with *Xba*I
25 (Promega) as described previously (Yanagi et al., 1997). The integrity of the RNA was checked by electrophoresis through agarose gel stained with ethidium bromide (Yanagi et al., 1997). Each transcription mixture was diluted with 400 µl of ice-cold phosphate-buffered
30 saline without calcium or magnesium and then immediately frozen on dry ice and stored at -80°C. Within 24 hours, both transcription mixtures were injected into the same chimpanzee by percutaneous intrahepatic injection guided
35 by ultrasound (Yanagi et al., 1998, 1999). If the

- 36 -

° chimpanzee did not become infected, the same transfection was repeated once. After two negative results, the next clone was inoculated into the same chimpanzee following the same protocol. Injections were
5 performed at weeks 0 and 2 with pH77CV-J6S, at weeks 5 and 8 with pH77(p7)CV-J6S, at weeks 14 and 16 with pH77-J6S, at weeks 19 and 23 with pH77(p7)-J6S, at week 28 with pJ6CF, and finally at week 34 with pCV-H77C. The chimpanzee was maintained under conditions that met
10 or exceeded all requirements for its use in an approved facility.

Serum samples were collected weekly from the chimpanzee and monitored for liver enzyme levels by
15 standard procedures, anti-HCV antibodies by the second-generation ELISA (Abbott) and HCV RNA by a sensitive RT-nested PCR assay with *AmpliTaq Gold* DNA polymerase using primers from the 5' UTR (Yanagi et al., 1996). Samples were scored as negative for HCV RNA if
20 two independent tests on 100 µl of serum were negative. The genome equivalent (GE) titer of HCV in positive samples was determined by RT-nested PCR on 10-fold serial dilutions of the extracted RNA (Bukh et al.,
25 1998). The consensus sequence of the complete ORF from the chimpanzee infected with RNA transcripts of pJ6CF was determined by direct sequencing of overlapping PCR products obtained by long RT-nested PCR as previously
30 described (Yanagi et al., 1997) with HC-J6 specific primers. After the intrahepatic transfection with RNA transcripts of pCV-H77C, we performed H77(genotype 1a)-specific RT-nested PCR with primers 2427S-H77 and
2832R-H77 for the 1st round and with primers 2462S-H77
35 and 2645R-H77 for the 2nd round (Table 3). The

- 37 -

° sensitivity of this assay was equivalent to that of the assay using 5' UTR primers when testing serum containing only H77, genotype 1a. The genome titer of genotype 1a was determined by using this specific RT- nested PCR on
5 10-fold serial dilutions of the extracted RNA.

EXAMPLE 1

Sequence analysis of HCV strain HC-J6_{CH}

10 As minor deviations from the consensus amino acid sequence were found previously to render full-length HCV cDNA clones noninfectious (Yanagi et al., 1997, 1998), the consensus sequence of the cloning source of genotype 2a (strain HC-J6_{CH}) was determined
15 prior to constructing any full-length clones. In brief, a plasma pool containing strain HC-J6_{CH} was prepared from acute phase plasmapheresis units collected from a chimpanzee experimentally infected with HC-J6 (Okamoto et al., 1991). The HCV genome titer of this pool was
20 10^{5.4} genome equivalents (GE)/ml (Quantiplex HCV RNA bDNA 2.0, Chiron) and the infectivity titer was 10⁴ chimpanzee infectious doses/ml.

25 The consensus sequence of the 5' UTR of HC-J6_{CH} (nts. 17-340) was deduced from 5 clones containing nts. 17-297 and 8 clones containing nts. 86-340. The 5' UTR of the various clones was highly conserved, but the consensus sequence of HC-J6_{CH} differed by 2 nucleotides
30 from that published previously for HC-J6 (Okamoto et al., 1991: C to T at position 36 and T to C at position 222).

35 The consensus sequence of 14 clones of the 3' UTR of HC-J6_{CH} indicated that the 39 nucleotide long variable region was highly conserved in this strain and

- 38 -

° was identical to that previously published for HC-J6 (Okamoto et al., 1991). The polypyrimidine tract varied greatly in length (84-164 nucleotides), and contained some conserved A residues. In the conserved region, the proximal 16 nucleotides were identical to those previously published for isolates of different HCV genotypes (Kolykhalov et al., 1996; Tanaka et al., 1996; Yamada et al., 1996). The remaining 82 nucleotides of the conserved region were determined for other genotype 2a strains (Tanaka et al., 1996) but not for HC-J6 or HC-J6_{CH}.

The ORF of HC-J6_{CH} was amplified in 3 fragments by RT-PCR (Fig. 1). Eight clones of the J6S fragment (nts. 86-2761), 6 clones of the J6B fragment (nts. 2573-5488) and 6 clones of the J6A fragment (nts. 5515-9298) were sequenced. PCR fragments containing nts. 5489-5514 were sequenced directly. A quasispecies was found at 243 nucleotide (2.7%) and 69 amino acid (2.3%) positions, scattered throughout the 9099 nts (3033 aa) of the ORF. However, the majority, 231 nucleotide substitutions, were detected only once and 71.6 % of these represented silent mutations. The 12 remaining nucleotide substitutions were each restricted to 2 clones and only 4 of these resulted in amino acid changes. The nucleotide difference among the J6S clones ranged from 0.1 - 1.3%, among the J6B clones it ranged from 0.1 - 0.3%, and it ranged from 0.2 - 4.0% among the J6A clones (Fig. 2). Three of 8 J6S clones, 4 of 6 J6B clones, and all 6 J6A clones had defective polyproteins due to nucleotide deletions, insertions or substitutions.

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- 39 -

° The sequences of clones of strain HC-J6_{CH} were relatively homogeneous. This was highlighted by the high degree of conservation among clones of the HVR1 (Fig. 3), a region frequently used to study the quasispecies of HCV (Bukh *et al.*, 1995). An exception was the sequence of clone J6A1, which differed by about 4% from the other clones of this region (Fig. 2). Importantly, the consensus sequence of strain HC-J6_{CH} (nts. 17-9629) could be determined with no ambiguity at the nucleotide or deduced amino acid level. The difference between the consensus ORF sequence of HC-J6_{CH} from the experimentally infected chimpanzee and that of HC-J6 of the inoculum (Okamoto *et al.*, 1991) was 4.1 % and 2.2 % at the nucleotide and deduced amino acid levels, respectively (Fig. 2, Table 2). Moreover, we found that 12 (44.4%) of the 27 amino acids constituting HVR1 differed between HC-J6_{CH} and HC-J6 (Fig. 3). Such diversities are greater than the < 2 % generally considered to comprise a quasispecies. In fact, these differences are equivalent to those found between the two prototype strains of HCV genotype 1a [strains HCV-1 (Choo *et al.*, 1991) and H77 (Yanagi *et al.*, 1997)]. These results indicated that HC-J6_{CH}, which represented the major species in the experimentally infected chimpanzee, was a minor species in the original inoculum.

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TABLE 2

Percent difference of nucleotide and predicted amino acid sequences between strain HC-J6 (Okamoto *et al.*, 1991) and strain HC-J6_{CH} from acute phase plasma pool of a chimpanzee inoculated with HC-J6

Genome Region	nt.position ^a	% nt. difference	% a.a. difference
ORF	341-9439	4.1 (373/9099) ^b	2.2 (66/3033) ^b
5' UTR	17-340	0.6 (2/324)	
Core	341-913	0.5 (3/573)	0 (0/191)
E1	914-1489	4.3 (25/576)	2.1 (4/192)
HVR1	1490-1570	24.7 (20/81)	44.4 (12/27)
E2-HVR1	1571-2590	3.9 (40/1020)	3.2 (11/340)
p7	2591-2779	3.7 (7/189)	3.2 (2/63)
NS2	2780-3430	4.0 (26/651)	2.8 (6/217)
NS3	3431-5323	4.0 (76/1893)	0.8 (5/631)
NS4A	5324-5485	4.3 (7/162)	1.9 (1/54)
NS4B	5486-6268	3.7 (29/783)	0.4 (1/261)
NS5A	6269-7666	5.4 (75/1398)	3.4 (16/466)
NS5B	7667-9439	3.7 (65/1773)	1.4 (8/591)
3' UTR	9440-9481	0 (0/42)	

^a The nucleotide positions correspond to those of the infectious full-length genotype 2a clone (pJ6CF).

^b The numbers in parenthesis indicate the nucleotide or amino acid differences for each region.

Example 2

Chimeric molecular clones

As chimeric flaviviruses with substituted structural genes have been useful in defining the biological function of viral sequences or proteins, in analyzing immune responses and in generating attenuated vaccine candidates (Bray and Lai, 1991; Chambers *et al.*, 1999; Pletnev *et al.*, 1992, 1993, 1998). The consensus sequence of the 2a structural genes and surrounding region was substituted for that of the infectious 1a cDNA clone. In the genotype 1a backbone, two silent mutations were introduced for cloning purposes [at positions 2765 (p7) and 9158 (NS5B) of pCV-H77C] (Fig. 4). The complete sequence of each chimera was verified. Infectivity of RNA transcripts from four different

-41-

intertypic chimeric clones (Figs. 4, 5A, 5B) was evaluated by consecutive intrahepatic transfections of a chimpanzee. Clones were considered not to be viable if viral RNA was not detected in the serum within two weeks of the repeat transfection. All chimeric clones contained the C, E1 and E2 genes of genotype 2a. The two chimeric clones tested initially differed from each other in that one had the p7 gene of 2a (pH77CV-J6S) and the other [pH77(p7)CV-J6S] the p7 gene of 1a. They differed from the two other clones in that the 186 nucleotides of the 5' UTR just upstream of the initiation codon were from the 2a genotype. Since neither clone containing the chimeric 5' UTR was infectious, the chimeric 5' UTR was replaced with the consensus genotype 1a 5' UTR to generate the two p7 varieties [pH77-J6S and pH77(p7)-J6S]. After consecutive transfection of the four clones, no HCV RNA, anti-HCV or ALT elevation was detected in the chimpanzee during 28 weeks of follow-up, suggesting that RNA transcripts from these intertypic chimeric clones were not viable *in vivo*.

This finding that the intertypic clones between genotypes 1a and 2a were not viable was surprising since flavivirus chimeras containing the structural region of dengue virus type 1 or 2 or of tick-borne encephalitis virus and the nonstructural region of an infectious dengue type 4 virus were viable (Bray and Lai, 1991; Pletnev et al., 1992, 1993). While considerable sequence variation exists between the infectious genotype 1a and 2a clones of HCV (Table 3), these viruses exhibit a higher degree of genetic heterogeneity than do the major genotypes of HCV. For other flaviviruses, however, it was possible to obtain

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- 42 -

infectious chimeric clones only if the capsid region was derived from the backbone cDNA clone (Chambers et al., 1999; Pletnev and Men, 1998).

TABLE 3

Percent difference of the amino acid sequences between the infectious clone of genotype 1a (pCV-H77C; Yanagi et al., 1997) and the infectious clone of genotype 2a (pJ6CF) of hepatitis C virus

Genome Region ^a	% difference
Polyprotein	27.9 (839/3007) ^b
Core	8.9 (17/191)
E1	37.0 (71/192)
HVR1	59.3 (16/27)
E2 ^{HVR1}	27.1 (91/336)
p7	38.1 (24/63)
NS2	41.9 (91/217)
NS3	19.2 (121/631)
NS4A	33.3 (18/54)
NS4B	26.8 (70/261)
NS5A	38.5 (171/444)
NS5B	25.2 (149/591)

^a Genome regions defined as in Table 1.

^b The numbers in parenthesis indicate the amino acid differences for each region. Positions with deletions or insertions in E2 (4 aa positions) and NS5A (26 aa positions) were not considered.

Trivial explanations may account for the lack of viability of these intertypic chimeras. First, the two silent mutations introduced in the genotype 1a backbone (one in p7 and one in NS5B) for cloning purposes could potentially eliminate infectivity. This is, however, very unlikely since mutations at these positions exist among field isolates of HCV including strain HC-J6_{CH} (Bukh et al., 1998). Also, it is noteworthy that the three previously published infectious clones of strain H77 had numerous silent nucleotide differences (Hong et al., 1999; Kolykhalov et al., 1997; Yanagi et al., 1997). Second, signal peptidases might not cleave the chimeric E2/p7 or p7/NS2

-43-

junction. This seems unlikely, however, since
eukaryotic signal peptidases typically recognize the
amino acid sequences upstream of the cleavage site [the
(-3, -1) rule] (Nielsen *et al.*, 1997) and the amino
acids at these two sites are conserved between genotypes
1a and 2a (Fig. 5B). Finally, the E2/p7 and/or p7/NS2
gene junctions could differ between genotypes 1a and 2a.
The junctions determined for genotypes 1a and 1b were
used (Lin *et al.*, 1994; Mizushima *et al.*, 1994; Selby *et al.*, 1994) because those for genotype 2a have not been
identified. In the latter two cases, further analyses
of genotype 2a should eventually provide sufficient data
to overcome such potential problems and it would most
likely be possible to construct a viable chimera.

More complicated explanations for the lack of
viability of the chimeras might be required if critical
genotype-specific interactions occur as regards the
structural proteins, the nonstructural proteins and the
genomic RNA. For instance, one cannot rule out that the
chimeras were not viable because the IRES function was
compromised. In *in vitro* studies the IRES activity
depended on RNA sequences not only in the 5' UTR but
also extending 3' of the translation initiation site
(Hahm *et al.*, 1998; Lemon and Honda, 1997; Reynolds *et al.*, 1995). Although the 3' border of the HCV IRES is
still controversial it is believed to involve at most
the first 39 nts of the core gene (Lemon and Honda,
1997). The 5' UTR of the intertypic chimeras was either
a chimera of genotype 1a and 2a sequences or the entire
5' UTR was derived from the 1a clone (Figs. 4, 5A).
Importantly, the 5' end of core is conserved among
genotypes 1a and 2a (Fig. 5A). Thus, the predicted

- 44 -

- ° IRES-like secondary structure is maintained in these chimeras, suggesting that the IRES activity most likely was maintained.

Possible interactions between the structural proteins and the nonstructural proteins and/or the genomic RNA, which involve RNA packaging, replication or translation are conceivable. In poliovirus, which is another positive-sense RNA virus, functional coupling of RNA packaging to RNA replication and of RNA replication to translation have been suggested (Novak and Kirkegaard, 1994 ; Nugent et al., 1999). Similar to other viruses of the *Flaviviridae* family, a membrane-associated replicase complex is thought to initiate replication at the 3' end of HCV and to synthesize a complementary negative-strand RNA (Rice, 1996). The putative cis-acting elements at the 5' and 3' termini which are believed to be important for viral genome replication (Rice 1996; Frolov et al., 1998) should be maintained in the intertypic HCV chimeras at least in the two constructs with the authentic 1a 5'UTR. However, it is conceivable that the viral packaging system was interrupted (Frolov et al., 1998). Studies using a Kunjin flavivirus replicon system and providing the structural proteins *in trans* suggested that the essential encapsidation signals did not reside in the structural region of the genome (Khromykh et al., 1997, 1998). The location of the packaging signals of HCV is not known. However, if the structural proteins encapsidate viral RNA via genotype-specific sequences outside of the structural region, the chimeras would be unable to package the RNA and it might be extremely

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- 45 -

- difficult to construct viable chimeras between highly divergent strains.

Example 3

5 A consensus molecular clone of
 genotype 2a is infectious in vivo

 In order to prove that the genotype 2a portion
 used in the 4 intertypic chimeric cDNA clones indeed
 represented the infectious sequence, a consensus full-
10 length cDNA clone of HC-J6_{CH} (pJ6CF) was constructed.
 The core sequence of the T7 promoter, a 5' guanosine
 residue and the full-length sequence of HC-J6_{CH} (9711
 nts) were cloned into pGEM-9Zf vector using NotI/XbaI
15 sites. Within the HCV sequence there were no deduced
 amino acid differences and only 4 nucleotide differences
 (at nucleotide positions 1822, 5494, 9247 and 9289) from
 the consensus sequence of HC-J6_{CH} as determined in the
 present study. The silent mutation at position 1822 was
20 within the structural region and so was also present in
 the four intertypic chimeras. The 5' terminal 16 nts
 and the 3' terminal 82 nts were deduced from previously
 published HCV genotype 2a sequences (Okamoto et al.,
25 1991, Tanaka et al., 1996). The full-length cDNA clone
 of genotype 2a contained a 5' UTR of 340 nts, an ORF of
 9099 nts encoding 3033 amino acids and a 3' UTR
 consisting of a variable region of 39 nts followed by a
 132 nucleotide-long polypyrimidine tract interrupted
30 with 3 A residues and the 3' terminal conserved region
 of 98 nts.

 RNA transcripts from pJ6CF were injected into
 the same chimpanzee used for injection of the 4
35 intertypic chimeras. The chimpanzee became infected at

- 46 -

the first attempt with an HCV titer of 10^2 GE/ml at week 1 post inoculation (p.i.), and 10^3 - 10^4 GE/ml during weeks 2 to 6 p.i. The consensus sequence of PCR products of the complete ORF, amplified from serum obtained during week 5 p.i., was identical to the sequence of pJ6CF and there was no evidence of a quasispecies. Since RNA transcripts of this infectious genotype 2a clone were infectious *in vivo*, and it shared an exact sequence with the non-infectious intertypic chimeric clones, their failure to replicate must have been the result of incompatibilities between the genotype 1a and 2a sequences.

To confirm that the chimpanzee used was susceptible also to infection by genotype 1a, which comprised most of the intertypic chimeras, the chimpanzee was subsequently inoculated with RNA transcripts from the infectious genotype 1a clone (pCV-H77C). Serum samples were tested in an H77-specific RT-PCR assay to identify super-infection with genotype 1a. At week 1 p.i. the total HCV genome titer was 10^4 GE/ml and the H77-specific (1a) genome titer was 10^2 GE/ml. The H77-specific genome titer increased to 10^3 GE/ml at week 2 p.i., and reached 10^4 GE/ml during weeks 3-6 p.i. The consensus sequence of PCR products amplified with H77-specific primers at weeks 1-6 p.i. were found to be identical to that of pCV-H77C. However, the direct sequences of PCR products amplified with the 5' UTR primers at weeks 1-2 after inoculation of pCV-H77C were identical to that of pJ6CF indicating that the 2a genotype was still present and represented the majority species. These experiments confirmed that the inability of the intertypic 1a, 2a

- 47 -

° cDNA clones to infect the chimpanzee was not the result of protective immune responses in the chimpanzee but represented deficiencies intrinsic to the chimeras.

Discussion

5 The published infectious cDNA clones of HCV represent the two most important subtypes of genotype 1 (Hong et al., 1999; Kolykhalov et al., 1997; Yanagi et al., 1997, 1998). However, 5 more major genotypes of
10 HCV are recognized. In the above Examples, the infectivity of a cDNA clone of a second major HCV genotype was demonstrated. As in previous studies, the infectivity of RNA transcripts was demonstrated *in vivo*
15 by intrahepatic transfection of a chimpanzee. This new infectious clone (pJ6CF) encodes the consensus polyprotein of HCV strain HC-J6_{CH}, genotype 2a. Its encoded polyprotein differs from those of the infectious clones of genotypes 1a and 1b by approximately 30%
20 (Table 2). Genotype 2 strains, in particular subtypes 2a and 2b, have a worldwide distribution and important differences between genotypes 1 and 2 with respect to pathogenesis and treatment were indicated in previous
25 studies. The availability of an infectious clone representing a second major genotype of HCV should permit new ways of studying the molecular biology and immunopathology of this important and genetically quite different human pathogen.

30 The 5' and 3' UTRs of HCV are believed to be critical for viral replication, translation and viral packaging (Rice, 1996). The 5' 203 terminal nucleotides and the 3' 101 terminal nucleotides of the published
35 infectious clones of genotypes 1a and 1b were identical.

-48-

However, the sequences of UTRs of the genotype 2a clone differ from those of the genotype 1 clones. Overall, the 5' UTR of the genotype 2a clone has 17 nt differences and a single nucleotide deletion compared with the infectious clones of genotype 1a (Fig. 5A). Five of these differences and the deletion are within the first 30 nucleotides, whereas the remainder are found within the predicted IRES structure. Differences also exist between the 3' UTR of the genotype 2a clone and the clones of genotype 1a (Fig. 5B). The sequences of the variable region are very different. Recent study has shown this region is not critical for infectivity *in vivo* (Yanagi et al., 1999). Within the regions which are critical for infectivity *in vivo* (Yanagi et al., 1999), the 132 nucleotide-long polypyrimidine tract of the genotype 2a clone has 3 unique A residues interspersed and the 3' terminal conserved region of 98 nts has 4 nt differences within the 3' terminal stable stem-loop structure (Fig. 5B) (Kolykhalov et al., 1996; Tanaka et al., 1996). Since the 2a clone was infectious these sequence differences are apparently real and are compatible with infectivity. Further studies are required to determine whether these represent critical genotype-specific sequences.

- 49 -

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- 55 -

WHAT IS CLAIMED IS:

1. A purified and isolated nucleic acid molecule which encodes human hepatitis C virus of genotype 2a, said molecule capable of expressing said virus when transfected into cells.
2. The nucleic acid molecule of claim 1, wherein said molecule encodes the amino acid sequence of SEQ ID NO:2.
3. The nucleic acid molecule of claim 2, wherein said molecule comprises the nucleic acid sequence of SEQ ID NO:1.
4. A DNA construct comprising a nucleic acid molecule according to claim 1.
5. A DNA construct comprising a nucleic acid molecule according to claim 3.
6. An RNA transcript of the DNA construct of claim 4.
7. An RNA transcript of the DNA construct of claim 5.
8. A cell transfected with the DNA construct of claim 4.
9. A cell transfected with the DNA construct of claim 5.
10. A cell transfected with RNA transcript of claim 6.

- 56 -

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11. A cell transfected with RNA transcript of claim 7.

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12. A hepatitis C virus polypeptide produced by the cell of claims 8 or 9.

13. A hepatitis C virus polypeptide produced by the cell of claims 10 or 11.

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14. A hepatitis C virus produced by the cell of claims 8 or 9.

15. A hepatitis C virus produced by the cell of claims 10 or 11.

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16. A hepatitis C virus whose genome comprises a nucleic acid molecule according to claim 1.

17. A hepatitis C virus whose genome comprises a nucleic acid molecule according to claim 3.

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18. A method for producing a hepatitis C virus comprising transfecting a host cell with the RNA transcript of claims 6 or 7.

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19. A polypeptide encoded by a nucleic acid sequence according to claim 1.

20. A polypeptide encoded by a nucleic acid sequence according to claim 3.

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21. The polypeptide of claim 19, wherein said polypeptide is selected from the group consisting of NS3 protease, E1 protein, E2 protein or NS4 protein.

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- 57 -

22. The polypeptide of claim 20, wherein said polypeptide is selected from the group consisting of NS3 protease, E1 protein, E2 protein or NS4 protein.

23. A method for assaying candidate antiviral agents for activity against HCV, comprising:

- a) exposing a cell containing the hepatitis C virus of claims 16 or 17 to the candidate antiviral agent; and
- b) measuring the presence or absence of hepatitis C virus replication in the cell of step (a).

24. The method of claim 23, wherein said replication in step (b) is measured by at least one of the following: negative strand RT-PCR, quantitative RT-PCR, Western blot, immunofluorescence, or infectivity in a susceptible animal.

25. A method for assaying candidate antiviral agents for activity against HCV, comprising:

- a) exposing an HCV protease encoded by a nucleic acid sequence according to claims 1 or 3 or a fragment thereof to the candidate antiviral agent in the presence of a protease substrate; and
- b) measuring the protease activity of said protease.

26. The method of claim 25, wherein said HCV protease is selected from the group consisting of an NS3 domain protease, an NS3-NS4A fusion polypeptide, or an NS2-NS3 protease.

- 58 -

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27. An antiviral agent identified as having antiviral activity for HCV by the method of claim 23.
28. An antiviral agent identified as having antiviral activity for HCV by the method of claim 25.
- 5 29. Antibody to the polypeptide of claim 19.
30. Antibody to the polypeptide of claim 20.
- 10 31. Antibody to the hepatitis C virus of claim 16.
32. Antibody to the hepatitis C virus of claim 17.
- 15 33. A method for determining the susceptibility of cells *in vitro* to support HCV infection, comprising the steps of:
- a) growing animal cells *in vitro*;
- 20 b) transfecting into said cells the nucleic acid of claim 1; and
- c) determining if said cells show indicia of HCV replication.
- 25 34. The method according to claim 33, wherein said cells are human cells.
35. A composition comprising a polypeptide of claim 19 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.
- 30 36. A composition comprising a polypeptide of claim 20 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.
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- 59 -

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37. A composition comprising a nucleic acid molecule of claim 1 suspended in a suitable amount of a pharmaceutically acceptable diluent or excipient.

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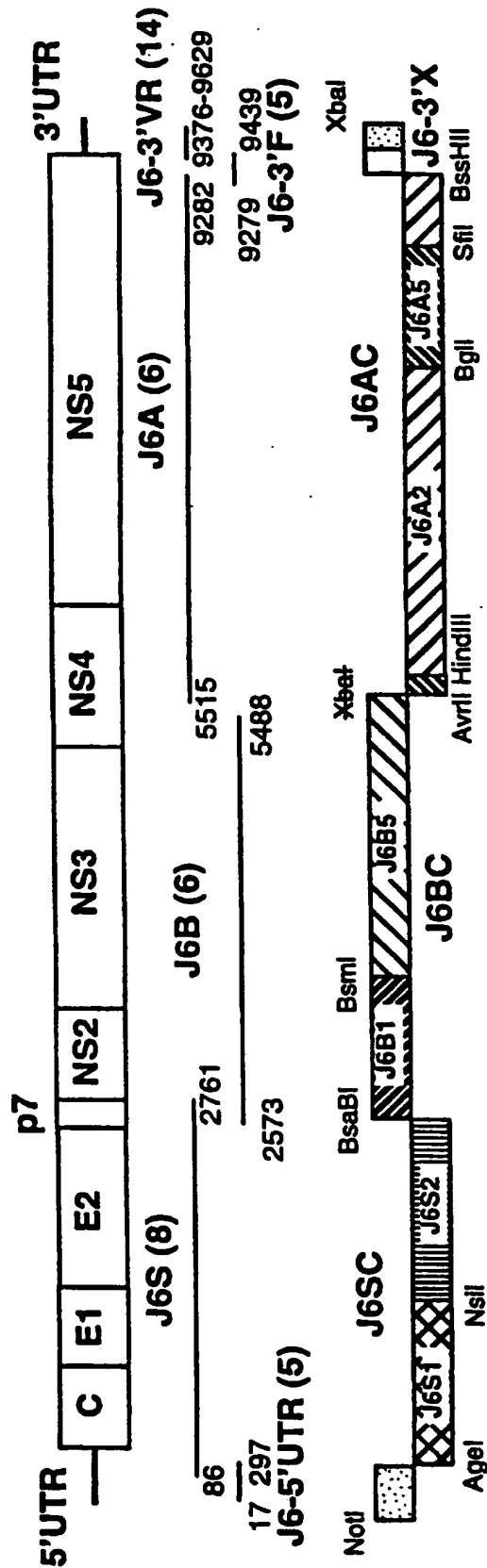


FIG. 1

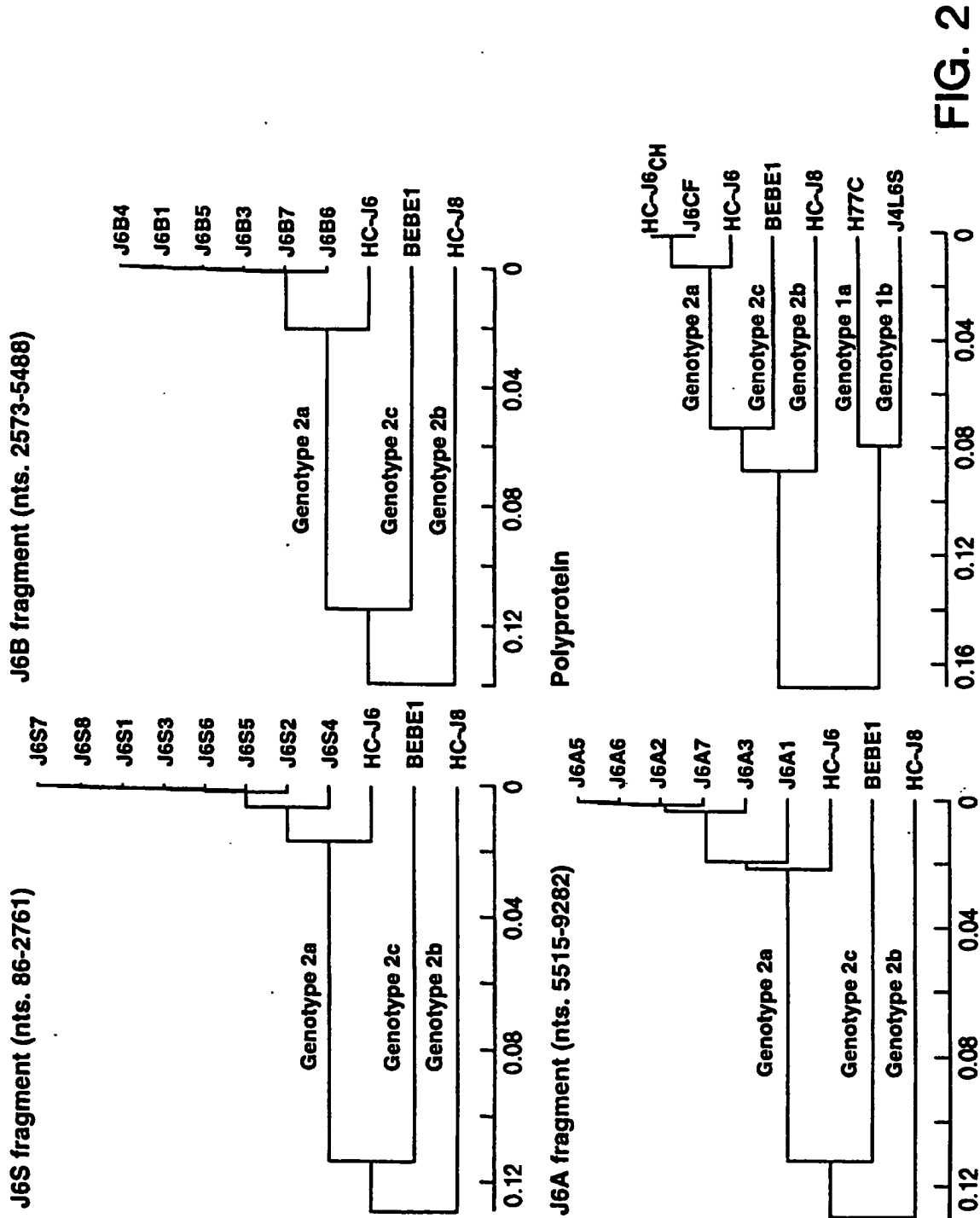


FIG. 2

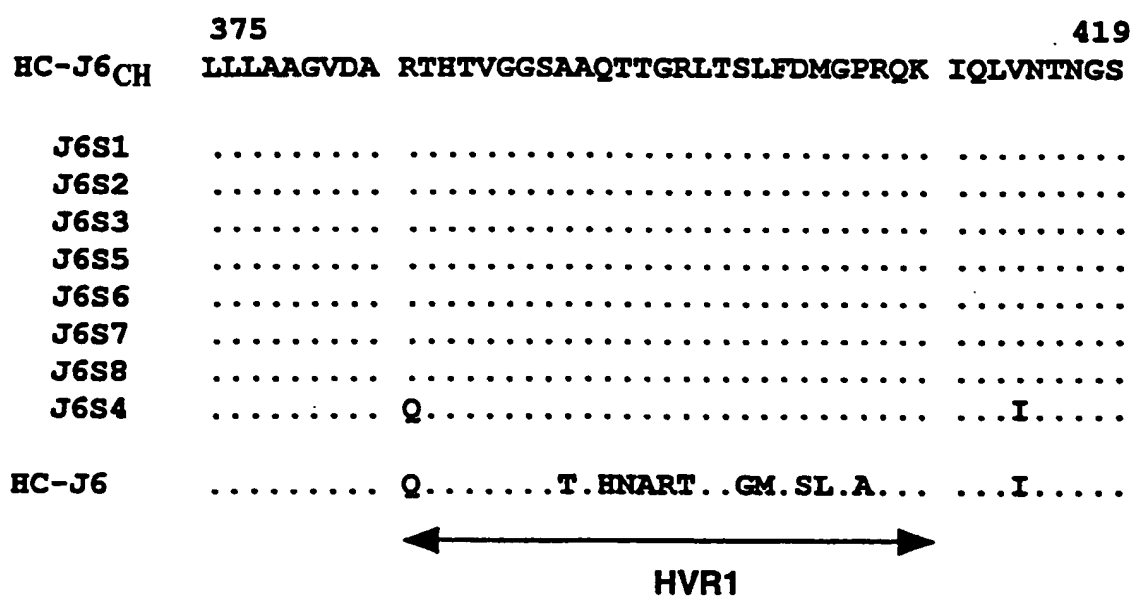


FIG. 3

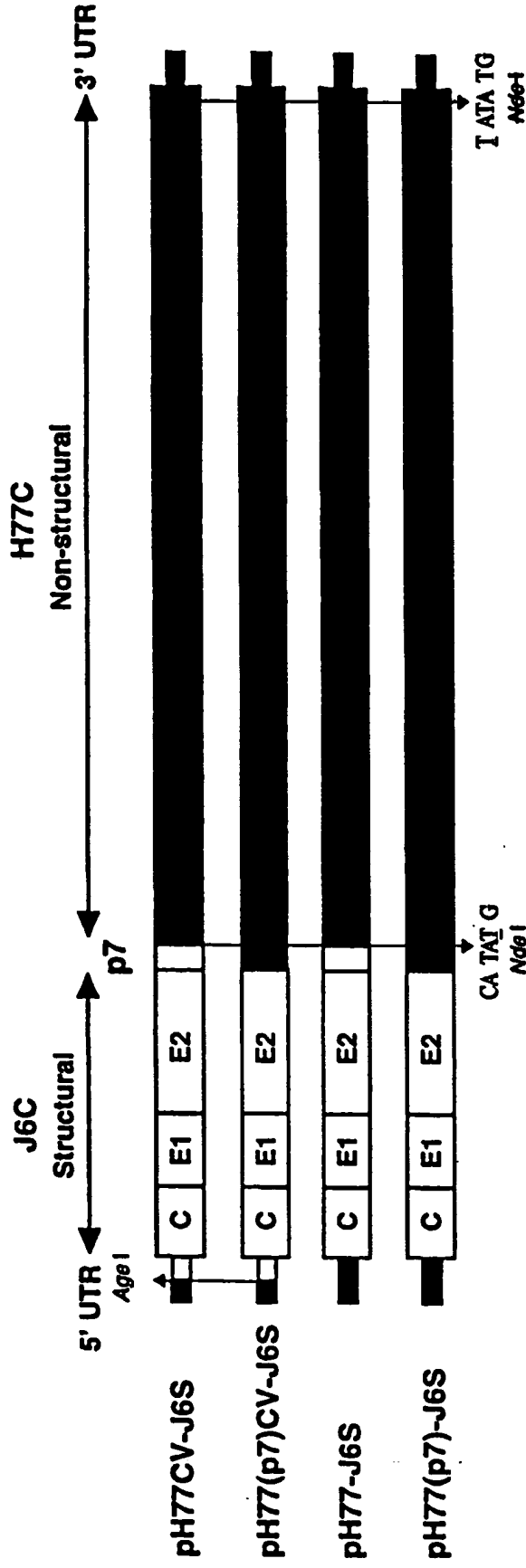


FIG. 4

5' Untranslated Region

FIG. 5A

	1	GCCAGCCCC	TCATGGGGC	GACACTCCAC	CATGAATCAC	TCCCCTGTGA	GGAAGCGTCT	AGCCATGGCG	90
H77C									
H77CV-J6S									
H77(p7)CV-J6S									
H77-J6S									
H77(p7)-J6S									
J6CF		A..C.....	.A..A.....	.G.....					
	91	TTAGTATGAG	TGTGTGTCAG	CCTCCAGGAC	CCCCCTCCC	GGGAGAGCCA	TAGTGTGTCG	CGGAACCGGT	180
H77C									
H77CV-J6S									
H77(p7)CV-J6S									
H77-J6S									
H77(p7)-J6S									
J6CF									
	181	GACGACCGGG	TCCTTCTTGG	GATAAACCGG	CTCAATGCCT	GGAGATTGG	GCGTGCCCCC	GCAAGACTGC	270
H77C									
H77CV-J6S									
H77(p7)CV-J6S									
H77-J6S									
H77(p7)-J6S									
J6CF									
	271	GCGAAAGGCC	TTGTGGTACT	GCCTGATAGG	GTGCTTGCGA	GTGCCCCGGG	AGGTCTCGTA	GACCGTGCAC	360
H77C									
H77CV-J6S									
H77(p7)CV-J6S									
H77-J6S									
H77(p7)-J6S									
J6CF									

Age I

FIG. 5B

3' Untranslated Region

	9375				9518
H77C	TGAAGGTGG	GGTAAACACT	CCGGCCTCTT	AAGCCATTTC	CTG (Polypyrimidine tract)81 AATGGTGGCT CCATCTTAGC
H77CV-J6S
H77(p7)CV-J6S
H77-J6S
H77(p7)-J6S
J6CF	AG..CGCA	CAC.TTAG..	A.ACT.CA.A	GCTAAC.G..	C- (Polypyrimidine tract)132 ---

	9519				9599
H77C	CCTAGTCACG	GCTAGCTGTG	AAAGGTCCGT	GAGCCGCATG	ACTGCAGAGA GTGCTGATAC TGGCCTCTCT GCAGATCATG T
H77CV-J6S
H77(p7)CV-J6S
H77-J6S
H77(p7)-J6S
J6CFC.TA.. ..T

E2/p7/NS2 Region

		E2/p7		p7		p7/NS2
	730					825
H77C	RVCSCLMMLISQAEA	ALENLVILNAASLAGTHGLVSFLVFFCF	AWYLKGRWVPGAVYALYGMWPLLL	LLALLPQRAYA	LDTEVAASCGGVVLVG	
H77CV-J6S	...A...LI.LG...	...K...H...A.SCN.FLY.VI..VA...	I...V...L.T.S.T.L.SFS...	Q...	
H77(p7)CV-J6S	...A...LI.LG...	...K...H...A.SCN.FLY.VI..VA...	I...V...L.T.S.T.L.SFS...	Q...	
H77-J6S	...A...LI.LG...	...K...H...A.SCN.FLY.VI..VA...	I...V...L.T.S.T.L.SFS...	Q...	
H77(p7)-J6S	...A...LI.LG...	...K...H...A.SCN.FLY.VI..VA...	I...V...L.T.S.T.L.SFS...	Q...	
J6CF	...A...LI.LG...	...K...H...A.SCN.FLY.VI..VA...	I...V...L.T.S.T.L.SFS...	Q...	Y.AS.HGQI.AAL..M	

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GCCAGCCCC	TGATGGGGG	GACACTCCAC	CATGAATCAC	TCCCCGTGTA	50
GGAACACTG	TCTTCACGA	GAAAGCGTCT	AGCCATGGCG	TTAGTATGAG	100
TGTGCTGCAG	CTCCAGGAC	CCCCCTCC	GGGAGAGCA	TAGTGGTCTG	150
CGGAACCGGT	GAGTACACG	GAATTGCCAG	GACGACCGGG	TCCCTTCTTG	200
GATAAACCG	CTCAATGCT	GGAGATTG	GCGTCCCC	GCAAGACTGC	250
TAGCCGAGTA	GTGTGGGTC	GCGAAAGGCT	TTGTGGTACT	GCCGTATAGG	300
GTGCTTGGA	GTGCCCCGG	AGGTCTCGTA	GACCGTGCAC	CATGAGCAAG	350
AATCCTAAAC	CTCAAGAAA	AACCAAGCT	AACACCAAC	GTCGCCACA	400
GGAGGTCAAG	TTCCCCGGTG	GCGGTGAGAT	CGTGGGTGGA	GTTTACTTGT	450
TGCCCCGCG	GGGCCCCAGA	TTGGGTGTGC	GCGCGACGAG	GAAGACTTCC	500
GAGCGGTGCG	AACTGAGG	TAGAGGTGAG	CCTATCCCCA	AGGCAGGTG	550
GCCCCAGGG	AGGACCTGG	CTCAGCCCCG	GTACCTTTGG	CCCCCTATG	600
GCAATGAGGG	TTGCGGGTGG	GCGGGATGGC	TCCGTCTCTC	CCGTGGCTCT	650
CGGCTTAGCT	GGGCCCCAC	AGACCCCCG	CGTAGGTGCG	GCAATTTGGG	700
TAAGGTATC	GATACCTTA	CGTGGGCTT	CGCCGACCTC	ATGGGGTACA	750
TACCGCTGCT	CGGCGCCCT	CTTGGAGGCG	CTGCCAGGGC	CCTGGCGCAT	800
GGCGTCCGG	TTCTGGAAGA	CGCGGTGAAC	TATGCAACAG	GGAACTTCC	850
TGGTGTCTCT	TTCTCTATCT	TCCCTCTGGC	CCTGCTCTCT	TGCCGTACTG	900
TGCCCCGCTT	AGCTTACCA	GTGCGCAATT	CCTCGGGGCT	TTACCATGTC	950
ACCAATGATT	GGCTTAACTC	GAGTATTGTG	TACGAGGCGG	CCGATGCCAT	1000
CCTGCACACT	CCGGGGTGTG	TCCCTTGCGT	TGCGAGGGT	AACGCTCGA	1050
GGTGTGGGT	GGGGGTGACC	CCACGGTGG	CCACCAGGGA	CGGCAACTC	1100
CCACAACGC	AGCTTCGAG	TCATATGAT	CTGCTTGTG	GGAGCGCAC	1150
CCTCTGCTCG	GGCTCTACG	TGGGGGACCT	GTGCGGGTCT	GTCCTTCTTG	1200
TTGGTCAACT	GTTTACCTTC	TCTCCAGGC	GCCACTGGAC	GACGCAAGAC	1250
TGCAATTGTT	CTATCTATCC	CGGOCATATA	AGGGGTATC	GCATGGCATG	1300
GGATATGATG	ATGAATGGT	CCCCACGGC	AGGTTGGTG	GTAGCTCAGC	1350
TGCTCCGGAT	CCACAAGCC	ATCATGGACA	TGATGCTGG	TGCTCACTGG	1400
GGAGTCTGG	CGGGCATAGC	GTATTTCTCC	ATGGTGGGA	ACTGGGCGAA	1450
GGTCTGGTA	GTGCTGCTGC	TATTTGCGG	CGTCGAGCG	GAAACCAAG	1500
TCACCGGGGG	AAATGCGGC	CGCACCAAGG	CTGGGCTTGT	TGGTCTCCTT	1550
ACAACAGGG	CCAAGCAGAA	CATCCAAGTG	ATCAACACCA	ACGGCAGTTG	1600
GCACATCAAT	AGCACGGCCT	TGAATTGCAA	TGAAAGCCTT	AACAACGGCT	1650
GGTTAGCAGG	GCTCTTCTAT	CAACACAAAT	TCAACTCTTC	AGGCTGTCTT	1700
GAGAGGTGG	CCAGCTGCGG	ACGCTTAC	GATTTTGCC	AGGGCTGGG	1750
TCCATACAGT	TATGCCAAG	GAAGCGGCT	CGACGAAAGC	CCCTACTGCT	1800
GGCACTACCC	TCAAGACCT	TGTGGCATTG	TGCCCCGAAA	GAGCGTGTGT	1850
GGCCCCGTAT	ATTGCTTCAC	TCCAGCCCC	GTGGTGGTGG	GAACGACCGA	1900

FIG. 6A

SUBSTITUTE SHEET (RULE 26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
CAGGTGCGGC	GCGCCTACCT	ACAGCTGGGG	TGCAAATGAT	ACGGATGTCT	1950
TGTCCTTAA	CAACACCAGG	CCACCGCTGG	GCAATTGGTT	CGGTGTGACC	2000
TGGATGAACT	CAACTGGATT	CACCAAGTIG	TGCGGAGGCG	CCCTTGTGT	2050
CATCGGAGGG	GTGGGCAACA	ACAACCTTGT	CTGCCCCACT	GATTGCTTCC	2100
GCAAACATCC	GGAGGCCACA	TACTCTGGGT	GCGGCTCCGG	TCCCTGGATT	2150
ACACCCAGGT	GCATGGTCCA	CTACCCGTAT	AGGCTTTGGC	ACTATCCTTG	2200
TACCATCAAT	TACACCATAT	TCAAAGTCAG	GATGTACGTG	GGAGGGGTGG	2250
AGCACAGGCT	GGAGGGGCGC	TGCAACTGGA	CGCGGGGGGA	ACGCTGTGAT	2300
CTGGAAGACA	GGGACAGGTC	CGAGCTCAGC	CGGTGCTGTC	TGTCCACCAC	2350
ACAGTGGCAG	GTCTTCCGT	GTCTTTTCAC	GACCCCTGCA	GCCTTGTCCA	2400
CCGGCCTCAT	CCACCTCCAC	CAGAACATTG	TGGACGTGCA	GTACTTGTAC	2450
GGGGTAGGGT	CAAGCATCGC	GTCTGGGGCC	ATTAAGTGGG	AGTACGTGCT	2500
TCTCCGTGTC	CTTCTGCTTG	CAGACGGGGG	CGTCTGCTCC	TGCTTGTGGA	2550
TGATGTACT	CATATCCCAA	GCGGAGGCGG	CTTTGGAGAA	CCTCGTAATA	2600
CTCAATGCAG	CATCCCTGGC	CGGGACGCAC	GGTCTTGTGT	CCTTCCCTGT	2650
GTCTTCTGTC	TTTGGGIGGT	ATCTGAAGGG	TAGGTGGGTG	CCCGGAGCGG	2700
TCTACGCCCT	CTACGGGATG	TGGCCTCTCC	TCCCTGCTCT	GCTGGCGTTG	2750
CCTCAGCGGG	CATACGCACT	GGACACGGAG	GTGCGCGCGT	CGTGTGGCGG	2800
CGTTGTCTTT	GTCGGGTAA	TGGCGCTGAC	TCTGTGCGCA	TATTACAAGC	2850
GCTATATCAG	CTGGTGCATG	TGGTGGCTTC	AGTATTTTCT	GACCAGAGTA	2900
GAAGCGCAAC	TGCAAGTGTG	GGTTCCCCCC	CTCAACGTCC	GGGGGGGGGG	2950
CGATGCCGTC	ATCTTACTCA	TGTGTGTAGT	ACACCCGACC	CTGGTATTTG	3000
ACATCACCAA	ACTACTCCTG	GCCATCTTGG	GACCCCTTTG	GATTCTTCAA	3050
GCCAGTTTGC	TTAAAGTCCC	CTACTTGGTG	CGCGTTCAAG	GCCTTCTCCG	3100
GATCTGCGCG	CTAGCGCGGA	AGATAGCGGG	AGGTCAATTAC	GTGCAAATGG	3150
CCATCATCAA	GTTAGGGGGG	CCTACTGGCA	CCTATGTGTA	TAAOCATCTC	3200
ACCCCTCTTC	GAGACTGGGC	GCACAACGGC	CTGCGAGATC	TGGCGGTGGC	3250
TGTGGAACCA	GTCGTCTTCT	CCCGAATGGA	GACCAAGCTC	ATCACGTGGG	3300
GGGCAGATAC	CGCCCGGTGC	GGTGACATCA	TCAAACGGCTT	GCCCGTCTCT	3350
GCCCGTAGGG	GCCAGGAGAT	ACTGCTTGGG	CCAGCCGACG	GAATGGTCTC	3400
CAAGGGGTGG	AGGTGTGCTG	CGCCCATCAC	GGCGTACGCC	CAGCAGACGA	3450
GAGGCCCTCT	AGGGTGTATA	ATCACCGAGC	TGACTGGGCG	GGACAAAAC	3500
CAAGTGGAGG	GTGAGGTCCA	GATCGTGTCA	ACTGCTACCC	AAACCTTCTT	3550
GGCAACGTGC	ATCAATGGGG	TATGCTGGAC	TGTCTACAC	GGGGCCGGAA	3600
CGAGGACCAT	CGCATCACCC	AAGGGTCTCT	TCATCCAGAT	GTATACCAAT	3650
GTGGACCAAG	ACCTTGTGGG	CTGGCCCGCT	CCTCAAGGTT	CCCGCTCATT	3700
GACACCTGT	ACCTGCGGCT	CCTCGGACCT	TTACCTGGTC	ACGAGGCACG	3750
CCGATGTCAT	TCCCGTGGCG	CGGCGAGGTG	ATAGCAGGGG	TAGCCTGCTT	3800

FIG. 6B

SUBSTITUTE SHEET (RULE 26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TGCCCCCGGC	CCATTTCCTA	CTTGAAAGGC	TCCTGGGGGG	GTCCGCTGTT	3850
GTGCCCCGGG	GGACACGGCG	TGGGCTATT	CAGGGGCGCG	GTGTGCACCC	3900
GTGGAGTGGC	TAAAGCGGTG	GACTTTATCC	CTGTGGAGAA	OCTAGGGACA	3950
ACCATGAGAT	CCCGGTGTT	CACGGACAAC	TCCTCTCCAC	CAGCAGTGCC	4000
CCAGAGCTTC	CAGGTGGGCC	ACCTGCATGC	TCCCACCGGC	AGCGGTAAAG	4050
GCACCAAGGT	CCCGCTGGG	TACGCAGGCC	AGGGCTACAA	GGTGTGGTGG	4100
CTCAACCCCT	CTGTTGCTGC	AACGCTGGGC	TTTGGTGCCT	ACATGTCCAA	4150
GGCCCATGGG	GTGTATCCCTA	ATATCAGGAC	CGGGGTGAGA	ACAATTACCA	4200
CTGGCAGGCC	CATCAGCTAC	TCCACCTAAG	GCAAGTTTCT	TGCGGACGGC	4250
GGGTGCTCAG	GAGGTGCTTA	TGACATAATA	ATTTGTGACG	AGTGCCACTC	4300
CACGGATGCC	ACATCCATCT	TGGGCATCGG	CACTGTCTTT	GACCAAGCAG	4350
AGACTGGGGG	GGCGAGACTG	GTGTGCTTCG	CCACTGCTAC	CCCTCCGGGC	4400
TCCGTCACTG	TGTOCCATCC	TAACATCGAG	GAGGTTCCTC	TGTCCACCAC	4450
CGGAGAGATC	CCCTTTTACG	GCAAGGCTAT	CCCOCTCGAG	GTGATCAAGG	4500
GGGGAAGACA	TCTCATCTTC	TGCCACTCAA	AGAAGAAGTG	CGACGAGCTC	4550
GCCGCGAAGC	TGGTCGCAAT	GGGCATCAAT	GCCGTGGCCT	ACTACCGCGG	4600
TCTTGACGTG	TCTGTTCATCC	CGACCAGCGG	CGATGTTGTC	GTCGTGTGGA	4650
CCGATGCTCT	CATGACTGGC	TTTACCGGGG	ACTTCGACTC	TGTGATAGAC	4700
TGCAACACGT	GTTGCTACTCA	GACAGTCGAT	TTCAGCCTTG	ACCCTACCTT	4750
TACCATTGAG	ACAACCACGC	TCCCCCAGGA	TGCTGTCTCC	AGGACTCAAC	4800
GCCGGGGCAG	GACTGGCAGG	GGGAAGCCAG	GCATCTATAG	ATTTGTGGCA	4850
CCGGGGGAGC	GCCCCCTCCG	CATGTTCCGAC	TGTCGGTCC	TCTGTGAGTG	4900
CTATGACGGG	GGCTGTGCTT	GGTATGAGCT	CACGCCCCGC	GAGACTACAG	4950
TTAGGCTACG	AGCGTACATG	AACACCCCGG	GGCTTCCCGT	GTGCCAGGAC	5000
CATCTTGAAT	TTTGGGAGGG	CGTCTTTACG	GGCTCACTC	ATATAGATGC	5050
CCACTTTTTTA	TCCCAGACAA	AGCAGAGTGG	GGAGAACTTT	OCTTAOCTGG	5100
TAGCGTACCA	AGCCACCGTG	TGCGCTAGGG	CTCAAGCCCC	TCCCCCATCG	5150
TGGGACCAGA	TGTGGAAGTG	TTTGATCCGC	CTTAAACCCA	CCCTCCATGG	5200
GCCAACACCC	CTGCTATACA	GACTGGGGCG	TGTTTCAGAAT	GAAGTCACCC	5250
TGACGCACCC	AATCACCAAA	TACATCATGA	CATGCATGTC	GGCGACCTTG	5300
GAGGTGCTCA	CGAGCAOCTG	GGTCTCGTT	GGCGGGGTCC	TGGCTGCTCT	5350
GGCCGGGTAT	TGCCGTGTCAA	CAGGCTGGGT	GGTCATAGTG	GGCAGGATCG	5400
TCTTGTCCGG	GAAGCCGGCA	ATTATACCTG	ACAGGGAGGT	TCTCTACCAG	5450
GAGTTCGATG	AGATGGAAGA	GTGCTCTCAG	CACTTACCGT	ACATCGAGCA	5500
AGGGATGATG	CTCGCTGAGC	AGTTCAAGCA	GAAGGCCCTC	GGCTOCTGTC	5550
AGACCGGGTC	CCGCCATGCA	GAGGTATATCA	CCCCTGCTGT	CCAGACCAAC	5600
TGCCAGAAAC	TGCAGGTCTT	TTGGGCGAAG	CACATGTGGA	ATTTCAITCAG	5650
TGGGATACAA	TACTTGGCGG	GCCTGTCAAC	GCTGCCGTGGT	AACCCCGCCA	5700

FIG. 6C

SUBSTITUTE SHEET (RULE 26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TTGCTTCATT	GATGGCTTTT	ACAGCTGCGG	TCACCAGGCG	ACTAACCCT	5750
GGCCAAAGCC	TCCTCTTCAA	CATATTGGGG	GGGTGGGTGG	CTGCCCAGCT	5800
CGCGGCCCCC	GGTGCCGCTA	CTGCCTTTGT	GGGTGCTGGC	CTAGCTGGCG	5850
CGCCCATCGG	CAGCGTTGGA	CTGGGGAAGG	TCCTCGTGGA	CATTCTTGCA	5900
GGGTATGGCG	CGGGGTGGC	GGGAGCTCTT	GTAGCATTC	AGATCATGAG	5950
CGGTGAGGTC	CCCTCCAGG	AGGACCTGGT	CAATCTGCTG	CCGGCCATCC	6000
TCTCGCTGG	AGCCCTTGTA	GTCGGTGTGG	TCTGGGCAGC	AATACTGGCG	6050
CGGCAGGTG	GCCCCGGCGA	GGGGGCAGTG	CAATGGATGA	ACGGGCTAAT	6100
AGCCTTCGCC	TCCCGGGGGA	ACCATGTTTC	CCCCAGGCAC	TAGTGGCGG	6150
AGAGCGATGC	AGCGGCCCCG	GTCAGTGGCA	TACTCAGCAG	CCTCAGTGTA	6200
ACCCAGCTCC	TGAGGCGACT	GCATCAGTGG	ATAAGCTCGG	AGTGTACCC	6250
TCCATGCTCC	GGTTCCTGGC	TAAGGGACAT	CTGGGACTGG	ATATGGGAGG	6300
TGCTGAGCGA	CTTTAAGACC	TGGCTGAAAG	CCAAGCTCAT	GCCACAACCTG	6350
CCTGGGATTC	CCTTTGTGTC	CTGCCAGCGC	GGGTATAGGG	GGGTCTGGCG	6400
AGGAGACGGC	ATTATGCACA	CTCGCTGCCA	CTGTGGAGCT	GAGATCAGTG	6900
GACATGTCAA	AAACGGGACG	ATGAGGATCG	TGGTCCCTAG	GACCTGCAGG	6950
AACATGTGGA	GTGGGACGTT	CCCCATTAAC	GCCTACACCA	CGGGCCCCCTG	6550
TACTCCCCCTT	CCTGCGCCGA	ACTATAAGTT	CGCGCTGTGG	AGGGTGTCTG	6600
CAGAGGAATA	CGTGGAGATA	AGGCGGGTGG	GGGACTTCCA	CTACGTATCG	6650
GGTATGACTA	CTGACAATCT	TAAATGCCCG	TGCCAGATCC	CATCGCCCGA	6700
ATTTTTCACA	GAATTGGACG	GGGTGCCGCT	ACACAGGTTT	GCGCCCCCTT	6750
GCAAGCCCTT	GCTGCGGGAG	GAGGTATCAT	TCAGAGTAGG	ACTCCACGAG	6800
TACCCGGTGG	GGTCCCAATT	ACCTTGGCAG	CCCGAACCGG	ACGTAGCCGT	6850
GTTGACGTCC	ATGCTCAGTG	ATCCCTCCCA	TATAACAGCA	GAGGCGGCGG	6900
GGAGAAGGTT	GGCGAGAGGG	TCACCCCTTT	CTATGGCCAG	CTCCTGGCT	6950
AGCCAGCTGT	CGCTCCATC	TCTCAAGGCA	ACTTGCACCG	CCAACCATGA	7000
CTCCCTGAC	GCCGAGCTCA	TAGAGGCTAA	CCTCCTGTGG	AGGCAGGAGA	7050
TGGGCGGCAA	CATCACCAGG	GTGTAGTCAG	AGAACAAGT	GGTGATTCTG	7100
GACTCCTTCG	ATCCGCTTGT	GGCAGAGGAG	GATGAGCGGG	AGGCTCCGT	7150
ACCTGCAGAA	ATTCTGCGGA	AGTCTCGGAG	ATTGCGCCGG	GCCCTGCGCG	7200
TCTGGGCGCG	GCCGACTTAC	AACCCCCCGC	TAGTAGAGAC	GTGGAAAAAG	7250
CCTGACTACG	AACCACTGT	GGTCCATGGC	TGCCCCCTAC	CACCTCCAGG	7300
GTCCCCCTCT	GTGCTCCGC	CTCGGAAAAA	GCGTACGGTG	GTCTCACC	7350
AATCAACCT	ATCTACTGOC	TGGGCGAGC	TGGCAACAA	AAGTTTTGGC	7400
AGCTCCTCAA	CTTCGGGCAT	TACGGGCGAC	AATACGACAA	CATCCTCTGA	7450
GCCCCGCCCT	TCTGGCTGCC	CCCCGACTC	CGACGTTGAG	TCCTATTCTT	7500
CCATGCCCCC	CCTGGAGGGG	GAGCCTGGGG	ATCCGGATCT	CAGCGACGGG	7550
TCATGGTCGA	CGGTACGTAG	TGGGGCGGAC	ACGGAAGATG	TGTGTGCTG	7600

FIG. 6D

SUBSTITUTE SHEET (RULE 26)

11/22

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
CTCAATGICT	TATTCTCGGA	CAGGCGCACT	CGTCACCCCG	TGCGCTGCGG	7650
AAGAACAAAA	ACTGCCCCATC	AACGCACTGA	GCAACTCGTT	GCTACGCCAT	7700
CACAATCTGG	TGTATTCCAC	CACTTCACGC	AGTGCTTGCC	AAAGGCAGAA	7750
GAAAGTCACA	TTTGACAGAC	TGCAAGTTCT	GGACAGCCAT	TACCAGGACG	7800
TGCTCAAGGA	GGTCAAAGCA	GCGGCGTCAA	AAGTGAAGGC	TAACTTGCTA	7850
TOOGTAGAGG	AAGCTTGCG	CCTGAAGCCC	CCACATTGAG	CCAAATCCAA	7900
GTTTGGCTAT	GGGGCAAAAG	ACGTCCGTTG	CCATCCCGA	AAGGCGGTAG	7950
CCCACATCAA	CTCGTGTGG	AAAGACTTTC	TGGAAGACAG	TGTAACACCA	8000
ATAGACACTA	CCATCATGGC	CAAGAACGAG	GTTTTCTGCG	TTCAGCCTGA	8050
GAAGGGGGGT	CGTAAGCCAG	CTCGTCTCAT	CGTGTTCGCC	GACCTGGGCG	8100
TGCGCGTGTG	CGAGAAGATG	GCCCTGTACG	ACGTGGTTAG	CAAGCTCCCC	8150
CTGGCCGTGA	TGGGAAGCTC	CTACGGATTC	CAATACTCAC	CAGGACAGCG	8200
GGTTGAATTC	CTCGTGCAG	CGTGGAAATC	CAAGAAGACC	CCGATGGGGT	8250
TCTCGTATGA	TACCCGCTGT	TTTGACTCCA	CAGTCACTGA	GAGCGACATC	8300
CGTACGGAGG	AGGCAATTTA	CCAATGTGTG	GACCTGGACC	CCCAAGCCCG	8350
CGTGGCCATC	AAGTCCCTCA	CTGAGAGGCT	TTATGTTGGG	GGCCCTCTTA	8400
CCAATTCAAG	GGGGGAAAAC	TGCGGCTACC	GCAGGTGCGG	CGCGAGCGGC	8450
GTA CTGACAA	CTAGCTGTGG	TAACACCCCT	ACTTGCTACA	TCAAGGCCCG	8500
GGCAGCCTGT	CGAGCCGCAG	GGCTCCAGGA	CTGCACCATG	CTCGTGTGTG	8550
GCGACGACTT	AGTCGTTATC	TGTGAAAGTG	CGGGGGTCCA	GGAGGACGCG	8600
GCGAGCCTGA	GAGCCTTCAC	GGAGGCTATG	ACCAGGTA CT	CCCCCCCCC	8650
CGGGGACCCC	CCACAACCAG	AATACGACTT	GGAGCTTATA	ACATCATGCT	8700
CCTCCAAAGT	GTCAGTCGCC	CACGACGGCG	CTGGAAAGAG	GGTCTACTAC	8750
CTTACCCGTG	ACCCCTACAA	CCCCCTCGCG	AGAGCCCGGT	GGGAGACAGC	8800
AAGACACACT	CCAGTCAATT	CCTGGCTAGG	CAACATAATC	ATGTTTGCCC	8850
CCACACTGTG	GGCGAGGATG	ATACTGATGA	CCCATTTCTT	TAGCGTCTCT	8900
ATAGCCAGGG	ATCAGCTTGA	ACAGGCTCTT	AAC TGTGAGA	TCTACGGAGC	8950
CTGCTACTCC	ATAGAAOCAC	TGGATCTACC	TCCAATCATT	CAAAGACTCC	9000
ATGGCCTCAG	CGCATTTTCA	CTCCACAGTT	ACTCTCCAGG	TGAAATCAAT	9050
AGGGTGGCGG	CATGCCTCAG	AAAAC TTGGG	GTCCCGCCCT	TGCGAGCTTG	9100
GAGACACCGG	GCCCCGAGCG	TCCGCGCTAG	GCTTCTGTCC	AGAGGAGGCA	9150
GGGCTGCCAT	ATGTGGCAAG	TACCTCTTCA	ACTGGGCAGT	AAGAACAAAG	9200
CTCAA ACTCA	CTCCAATAGC	GGCCGCTGGC	CGGCTGGACT	TGTCCGGTTG	9250
GTTCAAGGCT	GGCTACAGCG	GGGGAGACAT	TTATCACAGC	GTGTCTCATG	9300
CCCGGCCCCG	CTGGTTCTGG	TTTTTGCTAC	TCCTGCTCGC	TGCAGGGGTA	9350
GGCATCTACC	TCCTCCCCAA	CCGATGAAGG	TGGGGGTAAA	CACTCCGGCC	9400
TCTTAAGOCA	TTTCTGTGTT	TTTTTTTTTT	TTTTTTTTTT	TTTTTCTTTT	9450
TTTTTTTCTT	TCCTTTCTTT	CTTTTTTTTC	TTTCTTTTTC	CCTTCTTTAA	9500

FIG. 6E

SUBSTITUTE SHEET (RULE 26)

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TGGTGGCTCC	ATCTTAGCCC	TAGTCACGGC	TAGCTGTGAA	AGGTCCGIGA	9550
GCCGCATGAC	TGCAGAGAGT	GCTGATACTG	GCCTCTCTGC	AGATCATGT	9599

FIG. 6F

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
MSINPKPQRK	TKRNINRRPQ	DVKFTGGGQI	VGGVYLLPRR	GPRLGVRATR	50
KTISERSQPRG	RRQPIPKARR	PEGRIWAQPG	YFWFLYGNEG	CGWAGWLLSP	100
RGSRPSWGPT	DPRRRSRNLG	KVIDTLTQGF	ADLMGYIPLV	GAPLGGAARA	150
LAHGVRVLED	GVNYATGNLP	GCSFSIFLLA	LLSCLTVPAS	AYQVRNSSGL	200
YHVINDCENS	SIVYEADAI	LHTPGCVPCV	REGNASRCW	AVTPTVATRD	250
GKLPITQLRR	HIDLLVGSAT	LCSALYVGDL	CGSVFLVGQL	FTFSPPRRHW	300
TQDCNCSTYP	GHTTGHMAW	DMMNWSPTA	ALWVAQLLRI	PQAIMDMTAG	350
AHAGVLAGIA	YFSMGNWAK	VLWVLLLFAG	VDAEIHVITG	NAGRITAGLV	400
GLITPGAKQN	IQLININGSW	HINSTALNCN	ESLNTGWLAG	LFYQHKFNSS	450
GCPERLASCR	RLIDFAQGWG	PISYANGSGL	DERPYCWHYP	PRPCGIVPAK	500
SVCGPVYCFT	PSPVWGTID	RSGAPTYSWG	ANDIDVFVLN	NIRPPLGNWF	550
GCTWMNSTGF	TKVCGAPPCV	IGGVGNLILL	CPTDCFRKHP	EATYSRCGSG	600
PWITPRCWD	YPYRLAHYPC	TINYTIFKVR	MYVGGVEHRL	EAACNWIRGE	650
RCDLEDRDRS	ELSPLLLSTT	QWQVLPCSFT	TLPALSTGLI	HLHQNIVDMQ	700
YLYGVGSSIA	SWAIKWEYV	LLFLLADAR	VCSCIAMMLL	ISQAEAALEN	750
LVILNAASLA	GTHGLVSFLV	FFCFAWYKLG	RWVPGAVYAL	YGMWPLLLLL	800
LALPQRAYAL	DTEVAASCGG	VVLVGLMALT	LSPYYKRYIS	WCMWMLQYFL	850
TRVEAQLHW	VPFLNVRGGR	DAVILLMCV	HPTLVFDITK	LLAIFGFLW	900
ILQASLLKVP	YFVRVQGLLR	ICALARKIAG	GHYVQMAIK	LGALTGTIVY	950
NHILPLRDWA	HNGLRDLAVA	VEPVVFSRME	TKLITWGADT	AACGDIINGL	1000
PVSARRQGEI	LLGPADGMVS	KGWRLAPIT	AYAQQIRGLL	GCIITSLTGR	1050
DKNQVEGEVQ	IVSTATQTFL	ATCINGVCWT	VYHGAGIRTI	ASPKGPVIQM	1100
YTNVDQDLVG	WPAQGSRSRL	TFCTCGSSDL	YLVIRHADVI	FVRRRGDSRG	1150
SLLSPRPISY	LKGSSGGPLL	CPAGHAVGLF	RAAVCTRGVA	KAVDFIPVEN	1200
LGTIMRSPVF	TINSSPPAVP	QSFQVAHLHA	PTGSGKSTKV	PAAYAAQGYK	1250
VLVLNPSVAA	TLGFGAYMSK	AHGVDENIRT	GVRTITITGSP	ITYSTYKFL	1300
ADGGCSCGAY	DIICDECHS	TDATSILGIG	TVLDQAETAG	ARLVLATAT	1350
PPGSVTVSHP	NIEEVALSTT	GEIPFYGKAI	PLEVIKGRH	LIFCHSKKKC	1400
DELAACLVAL	GINAVAYYRG	LDVSVIPTSG	DVVVSTIDAL	MIGFTGDFDS	1450
VIDCNCVTIQ	TVDFSLDPTF	TIETITLPOD	AVSRITQRRGR	TGRGKPGIYR	1500
FVAPGERPSG	MFDSSVLCEC	YDAGCAWYEL	TPAETTVRLR	AYMNTPLPV	1550
QCDHLEFWEG	VFTGLTHIDA	HFLSQTKQSG	ENFPYLWAYQ	ATVCARAQAP	1600
PPSWDQWAKC	LIRLKPTLHG	PTFLLYRLGA	VQNEVILTHP	ITKYIMTOMS	1650
ADLEVVTSTW	VLVGGVLAAL	AAAYCLSTGCV	VIVGRIVLSG	KPALIPREV	1700
LYQEFDEMEE	CSQHLPTYEQ	GMLAEQFKQ	KALGLLQTAS	RHAEVITPAV	1750
QINWQKLEVF	WAKHMANFTS	GIQYLAGLST	LPGNPATIASL	MAFTAAVTSP	1800
LTTGQITLLEN	ILGGWAAQL	AAPGAATAFV	GAGLAGAAIG	SVGLGKVLVD	1850
ILAGYGAGVA	GALVAFKIMS	GEVPSTEDLV	NLLPAILSPG	ALVVGWVCAA	1900

FIG. 6G

H77C

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
ILRRHVGPE	GAVQAMNRLI	AFASRGNHVS	PIHYVPESDA	AARVTAILSS	1950
LIVTQLLRL	HQWISSECTT	PCSGSWLRDI	WDWICEVLSD	FKTWLKAKLM	2000
PQLPGIPFVS	QQRGYRGVWR	GDGIMHIRCH	CGAETTGHVK	NGIMRIVGPR	2050
TCRNWMSGIF	PINAYTTIGPC	TPLPAPNYKF	ALWRVSAEEY	VEIRRVGDFH	2100
YVSGMITDNL	KCPQIIPSPE	FFTLDGVRL	HRFAPPCKPL	LREEVSFRVG	2150
LHEYFVGSQ	PCEPEPDVAV	LTSMLTDPESH	TTAEAAGRRL	ARGSPPSMAS	2200
SSASQLSAPS	LKATCTANHD	SPDAELTEAN	LLWRQEMGGN	ITRVESENKV	2250
VILDSFDPLV	AEDEREVS	PAETLRKSRR	FARALPWAR	PDYNPPLVET	2300
WKKPDYEPV	VHGCPLPPPR	SPPVPPPRKK	RTVVLTESTL	STALAEATK	2350
SFGSSSTSGI	TGDNITTSSE	PAPSGCPPDS	DVESYSSMPP	LEGEFGDPL	2400
SDGSWSTVSS	GADTEDVCC	SMSYSWIGAL	VTPCAEEQK	LPINALNSL	2450
LRHNLVYST	TSRSACQK	KVTFDLQVL	DSHYQDLKE	VKAAASKVKA	2500
NLLSVEEACS	LTPPHSAKSK	FGYGAKDVRC	HARKAVAHIN	SWKDLLED	2550
VTPIDITIMA	KNEVFCVQPE	KGGRKPARLI	VFPDLGVRVC	EKMALYDWS	2600
KLPLAVMGSS	YGFQYSPQOR	VEFLVQAWKS	KKTPMGFSYD	TRCFDSTVTE	2650
SDIRTEEATY	QCCDLDPQAR	VAIKSLTERL	YVGGLINSR	GENGYRROR	2700
ASGVLTTSCG	NILTCYIKAR	AACRAAGLQD	CTMLVOGDDL	WICESAGVQ	2750
EDAASLRAFT	EAMIRYSAPP	GDPPQPEYDL	ELITSCSSNV	SVAHDGAGKR	2800
VYYLTRDPTT	FLARAAWETA	RHTFVNSWL	NIIMFAPTLW	ARMILMTHFF	2850
SVLIARDQLE	QALNCETYGA	CYSIEPLDLP	PIIQLRHGLS	AFSLHSYSFG	2900
EINRVAACLR	KLGVPLRAW	RHRARSVRAR	LLSRGGRAAI	CGKYLEFWAV	2950
RIKRLKLTPIA	AAGRDLGSGW	FTAGYSGGDI	YHSVSHARPR	WFWFCLLLA	3000
AGVGIYLLPN	R				3011

FIG. 6H

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
GCCAGCCCCC	TGATGGGGGC	GACACTCCAC	CATGAATCAC	TCCCCGIGA	50
GGAAGTACTG	TCTTACGCA	GAAAGCGTCT	AGCCATGGCG	TTAGTATGAG	100
TGTGCTGCAG	CCTCCAGGAC	CCCCCTCCC	GGGAGAGCCA	TAGTGGTCTG	150
CGGAACCGGT	GAGTACACCG	GAATTGCCAG	GACGACCGGG	TOCTTTCTTG	200
GATCAACCCG	CTCAATGCCT	GGAGATTGCG	GCGTGGCCCC	GCGAGACTGC	250
TAGCCGAGTA	GTGTTGGGTC	GCGAAAGGCC	TTGTGGTACT	GCCTGATAGG	300
GTGCTTGCGA	GTGCCCCGGG	AGGTCTCGTA	GACCGTGCAC	CATGAGCAAG	350
AATCCTAAAC	CTCAAAGAAA	AACCAACGCT	AACACCAACC	GCCGCCACAA	400
GGACGTCAAG	TTCCCCGGCG	GTGGTCAGAT	CGTTGGTGGG	GTTTACCTGT	450
TGCCGCGCAG	GGGCCCCAGG	TTGGGTGTGC	GCGGACTTAG	GAAGGCTTCC	500
GAGCGGTGCG	AACCTCGTGG	AAGGCGACAA	CCTATCCCAA	AGGCTCGCCG	550
ACCCGAGGGC	AGGGCCTGGG	CTCAGCCCCG	GTAACCTTGG	CCCCCTCATG	600
GCAATGAGGG	CCTGGGGTGG	GCAGGATGGC	TOCTGTACCC	CCGCGGCTCC	650
CGGCTAGTTC	GGGGCCCCAC	GGACCCCCCG	CGTAGGTCCG	GTAACCTGGG	700
TAAAGTCATC	GATACCCCTA	CATGCGGCTT	CGCCGATCTC	ATGGGGTACA	750
TTCCGCTCGT	CGGCGCCCCC	CTAGGGGGCG	CTGCCAGGGC	CTTGGCACAC	800
GGTGTCCGGG	TTCTGGAGGA	CGGCGTGAAC	TATGCAACAG	GGAACCTGCC	850
CGGTGTCTCT	TTCTCTATCT	TOCTCTTGGC	TCTGCTGTCC	TGTTTGACCA	900
TOCCAGCTTC	CGCTTATGAA	GTGCGCAACG	TGTCCGGGAT	ATACCATGTC	950
ACGAACGACT	GCTCCAACTC	AAGCATGTGT	TATGAGGCAG	CGGAAGTGAT	1000
CATGCATACT	CCCGGGTGGG	TGCCCCGTGT	TCAGGAGGGT	AACAGCTCCC	1050
GTTGCTGGGT	AGCGCTCACT	CCCAGGCTCG	CGGCCAGGAA	TGCCAGGGTC	1100
CCCACTACGA	CAATACGACG	CCACGTGGAC	TTGCTCGTTG	GGACGGCTGC	1150
TTTCTGCTCC	GCTATGTACG	TGGGGGATCT	CTGGGGATCT	ATTTTCTCTG	1200
TCTCCAGCT	GTTCAACCTC	TCGCCCTGCC	GGCATGAGAC	AGTGCAGGAC	1250
TGCAACTGCT	CAATCTATCC	CGGCCATGTA	TCAGGTACCC	GCATGGCTTG	1300
GGATATGATG	ATGAACTGGT	CACCTACAAC	AGCCCTAGTG	GTGTGGCAGT	1350
TGCTCCGGAT	CCCACAAGCT	GTCGTGGACA	TGGTGGCGGG	GGCCCACTGG	1400
GGAGTCTTGG	CGGGCCTTGC	CTACTATTCC	ATGGTAGGGA	ACTGGGCTAA	1450
GGTTCGTATT	GTGGCGCTAC	TCTTTGCCCG	CGTTGACGGG	GAGACCCACA	1500
CGACGGGGAG	GGTGGCCGGC	CACAACCCTT	CCGGGTTCAC	GTCCTTTTTC	1550
TCATCTGGGG	CGTCTCAGAA	AATCCAGCTT	GTGAATACCA	ACGGCAGCTG	1600
GCACATCAAC	AGGACTGCCC	TAAATTGCAA	TGACTCCCTC	CAAACTGGGT	1650
TCTTTGCCCG	GCTGTFTTAC	GCACACAAGT	TCAACTCGTC	CGGGTGGCCG	1700
GAGCGCATGG	CCAGCTGCCG	CCCCATTGAC	TGGTTGGCCC	AGGGGTGGGG	1750
CCCCATCACC	TATACTAAGC	CTAACAGCTC	GGATCAGAGG	CCTTATGTCT	1800
GGCATTAACG	GCCTCGACCG	TGTGGGTGTC	TACCCGGGTC	GCAGGTGTGT	1850
GGTCCAGTGT	ATTGTFTTAC	CCCAAGCCCT	GTTGTGGTGG	GGACCACCGA	1900

FIG. 7A

SUBSTITUTE SHEET (RULE 26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TCGTTCCGGT	GTCCCTACGT	ATAGCTGGGG	GGAGAATGAG	ACAGAOGTGA	1950
TGCTOCTCAA	CAACACGGGT	COGCCACAAG	GCAACTGGTT	CGCTGTIACA	2000
TGGATGAATA	GTACTGGGTT	CACTAAGACG	TGCGGAGGTC	CCCCGTGTAA	2050
CATCGGGGGG	GTCGGTAACC	GCACCTTGAT	CTGCCCCACG	GACTGCTTCC	2100
GGAAGCACCC	CGAGGCTACT	TACACAAAAT	GTCGGCTGGG	GOCTGGTTG	2150
ACAOC TAGGT	GOCTAGTAGA	CTACCCATAC	AGGCTTTGGC	ACTACCCCTG	2200
CACTCTCAAT	TTTTCCATCT	TTAAGGTTAG	GATGTATGTG	GGGGGGTGG	2250
AGCACAGGCT	CAATGCCGCA	TGCAATTGGA	CTCGAGGAGA	GCGCTGTAA	2300
TTGGAGGACA	GGGATAGGTC	AGAACTCAGC	COGCTGCTGC	TGTCIACAAC	2350
AGAGTGGCAG	ATACTGCOCT	GTCCTTTCAC	CACCCCTACG	GCTTTATCCA	2400
CTGGTTTGAT	CCATCTCCAT	CAGAACATCG	TGGACGTGCA	ATACCTGTAC	2450
GGTGTAGGGT	CAGCGTTTGT	CTCCTTTGCA	ATCAAATGGG	AGTACATCCT	2500
GTGCTTTTTC	CTTCTCCTGG	CAGACGGCGG	CGTGTGTGCC	TGCTGTGTGA	2550
TGATGCTGCT	GATAGCCACG	GCTGAGGCCG	CCTTAGAGAA	CTTGGTGGTC	2600
CTCAATGCCG	CGTCCGTGGC	CGGAGCGCAT	GGTATTCTCT	CCTTTCTTGT	2650
GTCTTTCTGC	GCCGCTGGT	ACATTAAAGG	CAGGCTGGCT	CTGGGGCGG	2700
CGTATGCTTT	TTATGGCGTA	TGGCCGCTGC	TCTGTCTCCT	ACTGGCGTTA	2750
CCACCACGAG	CTTACGCCTT	GGACCGGGAG	ATGGCTGCAT	CGTGGGGGG	2800
TGCGGTTCTT	GTAGGTCTGG	TATTCTTGAC	CTGTGCACCA	TACTACAAAG	2850
TGTTTCTCAC	TAGGCTCATA	TGGTGGTTAC	AATACTTTAT	CACCAGAGCC	2900
GAGGCGCACA	TGCAAGTGTG	GGTCCCCCCC	CTCAACGTTT	GGGAGGGCGG	2950
CGATGCCATC	ATCCTCCTCA	CGTGTGCGGT	TCATCCAGAG	TTAATTTTTG	3000
ACATCACCAA	ACTCCTGCTC	GCCATACTCG	GCCCGCTCAT	GGTGTCCAG	3050
GCTGGCATAA	CGAGAGTGCC	GTACTTGGTG	CGCGCTCAAG	GGCTCATTCG	3100
TGCATGCATG	TTAGTGCGAA	AAGTCCCGCG	GGGTCATTAT	GTCCAAATGG	3150
TCTTCATGAA	GCTGGGCGCG	CTGACAGGTA	CGTACGTTTA	TAACCATCTT	3200
ACCCCACTGC	GGGACTGGGC	CCACGCGGGC	CTACGAGACC	TTGGGGTGGC	3250
GGTAGAGCCC	GTCGTCTTCT	CCGCCATGGA	GACCAAGGTC	ATCAOCTGGG	3300
GAGCAGACAC	CGCTGCGTGT	GGGGACATCA	TCTTGGGTCT	ACCGTCTCC	3350
GCCCGAAGGG	GGAAGGAGAT	ATTTTTTGGG	CCGCTGATA	GTCTOGAAGG	3400
GCAAGGGTGG	CGACTCCTTG	CGCCCATCAC	GGCCTACTCC	CAACAAACGC	3450
GGGGGTACT	TGGTTGCATC	ATCACTAGCC	TCACAGGCCG	GGACAAGAAC	3500
CAGGTGGAAG	GGGAGGTTCA	AGTGGTTTCT	ACCGCAACAC	AATCTTTCTT	3550
GGCGAOC TGC	ATCAACGGCG	TGTGCTGGAC	TGTCIACCAT	GGCGCTGGCT	3600
CGAAGACCC	AGCCGGTCCA	AAAGGTCCAA	TCACCCAAAT	GTACACCAAT	3650
GTAGACCTGG	ACCTCGTCCG	CTGGCAGGCG	CCCCCGGGG	CGCGCTCCAT	3700
GACACCATGC	AGCTGTGGCA	GCTCGGACCT	TTACTTGGTC	ACGAGACATG	3750
CTGATGTCAT	TCCGGTGCGC	CGGCGAGGCG	ACAGCAGGGG	AAGTCTACTC	3800

FIG. 7B

SUBSTITUTE SHEET (RULE 26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TCCCCAGGC	CCGTCTCCTA	CCTGAAAGGC	TCTGGGGTGG	GTCCATTGCT	3850
TTGCCCCCTGG	GGGCAAGTGG	TGGGGGCTCT	CCGGGCTGCT	GTGTGCAACC	3900
GGGGGGTGG	GAAGGGGGTGG	GACTTCATAC	CCGTTGAGTC	TATGGAAACT	3950
ACCATGGGGT	CTCCGGTCTT	CACAGACAAC	TCAACCCCCC	CGGCTGTACC	4000
GCAGACATTC	CAAGTGGCAC	ATCTGCAAGC	TCTTACTGGC	AGGGGCAAGA	4050
GCACCAAGT	GCCGGCTGG	TATGCAGGCC	AAGGGTACAA	GGTGCTGGTC	4100
CTGAACCCGT	CCGTTGCGC	CACTTAGGG	TTTGGGGGGT	ATTATGTCCAA	4150
GGCACACGGT	ATCGACCTA	ACATCAGAAC	TGGGGTAAAG	AOCATTACCA	4200
CGGGCGGCTC	CATTAGGTAC	TCCACCTATG	GCAAGTTCTT	TGCGGACGGT	4250
GGCTGTCTTG	GGGGCGCTA	TGACATCATA	ATATGTGATG	AGTGCCACTC	4300
AACTGACTGG	ACTACCATCT	TGGGCATCGG	CACAGTCTTG	GACCAAGGGG	4350
AGACGGCTGG	AGCGGGGCTC	GTCGTGCTGG	CCACCGCTAC	AOCTCCGGGA	4400
TGGGTACCG	TGCCACACC	CAATATCGAG	GAAATAGGCC	TGTCCAACAA	4450
TGGAGAGATC	CCCTTCTATG	GCAAAGOCAT	CCCATTTGAG	GCCATCAAGG	4500
GGGGGAGGCA	TCTCATTTTC	TGCCATTCCA	AGAAGAAATG	TGACGAGCTC	4550
GCCGCAAGC	TGACAGGCT	CGGACTGAAC	GCTGTAGCAT	ATTACCGGGG	4600
CCTTGATGTG	TCCGTATAC	CGCCTATCGG	AGACGTGGTT	GTCGTGGCAA	4650
CAGACGCTCT	AATGACGGT	TTCACCGGGG	ATTTTGACTC	AGTGATCGAC	4700
TGCAATACAT	GTGTCAACCA	GACAGTCGAC	TTCAGCTTGG	ATCCCACTT	4750
CACCATTTGAG	ACGACGACCG	TGCCCCAAGA	CGCGGTGGTG	CGCTCGCAAC	4800
GGCGAGGTAG	AACTGGCAGG	GGTAGGAGTG	GCATCTACAG	GTTTGTGACT	4850
CCAGGAGAAC	GGCCCTCGGG	CATGTTGAT	TCTTCGGTCC	TGTGTGAGTG	4900
CTATGACCGG	GGCTGTGCTT	GGTATGAGCT	CACGCCCGCT	GAGACCTCGG	4950
TTAGGTTGCG	GGCTTACCTA	AATACACAG	GGTTGCCCGT	CTGCCAGGAC	5000
CATCTGGAGT	TCTGGGAGAG	CGTCTTCACA	GGCTCAACC	ACATAGATGC	5050
CCACTTCTCG	TCCCAGACTA	AACAGGCAGG	AGACAACCTT	CCTTACCTGG	5100
TGGCATATCA	AGCTACAGTG	TGCGCCAGGG	CTCAAGCTCC	ACCTCCATCG	5150
TGGGACCAAA	TGTGGAGTG	TCTCATACGG	CTGAAACCTA	CACTGCAAGG	5200
GCCAACACC	CTGCTGTATA	GGCTAGGAGC	CGTCCAAAT	GAGGTATCC	5250
TCACACACC	CATAACTAAA	TACATCATGG	CATGCATGTC	GGCTGACCTG	5300
GAGGTGCTCA	CTAGCACTCG	GGTGCTGGTA	GGGGAGTCC	TTCAGCTTT	5350
GGCGCATAC	TGCTGACGA	CAGGCAGTGT	GGICATTGTG	GGCAGGATCA	5400
TCTTGTCGGG	GAAGCCAGCT	GTCGTTCCCG	ACAGGGAAGT	CCTCTACCAG	5450
GAGTTGATG	AGATGGAAGA	GTGTGCTTCA	CAACTTCTTT	ACATCGAGCA	5500
GGGAATGCAG	CTCGCCGAGC	AATTCAAGCA	AAAGGGGCTC	GGGTGTGTGC	5550
AAACGGCCAC	CAAGCAAGCG	GAGGCTGCTG	CTCCCGTGGT	GGAGTCCAAG	5600
TGGCGAGCCC	TTGAGACCTT	CTGGGCGAAG	CACATGTGGA	ATTTCAATCAG	5650
CGGAATACAG	TACCTAGCAG	GCTTATCCAC	TCTGCTGGA	AAOCCCGCGA	5700

FIG. 7C

SUBSTITUTE SHEET (RULE 26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
TAGCATCATT	GATGGCATT	ACAGCTTCTA	TCACIAGCCC	GCTCACCACC	5750
CAAAACACCC	TCCTGTTTAA	CATCTTGGGG	GGATGGGTGG	CTGCCCCACT	5800
CGCTCCTCCC	AGCGCTGGGT	CAGCTTTGGT	GGGGGCGGGC	ATGCGCGGAG	5850
CGGCTGTGGG	CAGCATAGGC	CTTGGGAAGG	TGCTGGTGGG	CATCTTGGCG	5900
GGCTATGGGG	CAGGGGTAGC	CGGCGCACTC	GTTGGCTTTA	AGGTACATGAG	5950
CGGCGAGGTG	CCCTCCACCG	AGGACCTGGT	CAACTTACTC	CCTGCCATCC	6000
TCTCTCCTGG	TGCCCCGGTC	GTCGGGGTGG	TGTGGGCAGC	AATACTGGGT	6050
CGGCAAGTGG	GCCCCGGAGA	GGGGGCTGTG	CAGTGGATGA	ACCGGCTGAT	6100
AGGGTTGGCT	TGGGGGGGTA	ACCAAGTCTC	CCCTAAGCAC	TATGTGCCCTG	6150
AGAGCGAAGC	TGCAGCAAGT	GTCATCAGA	TCTCTCTAG	CCTTACCATC	6200
ACTCAACTGC	TGAAGCGGCT	CCACCAGTGG	ATTATGAGG	ACTGCTCTAC	6250
GCCATGCTCC	GGCTGGTGGC	TAAGGGATGT	TGGGATTGG	ATATGCAAGG	6300
TGTTGACTGA	CTTCAAGACC	TGGCTCCAGT	CCAAACTCCT	GCCGCGGTTA	6350
CCGGGAGTCC	CTTTCCTGTC	ATGCCAAGCC	GGGTACAAGG	GAGTCTGGCG	6400
GGGGGACGGC	ATCATGCAAA	CCACCTGCCC	ATGCGGAGCA	CAGATCGCCG	6450
GACATGTCAA	AAACGGTTCC	ATGAGGATCG	TAGGGCCTAG	AACCTGCAGC	6500
AACACGTGGC	ACGGAACGTT	CCCCATCAAC	GCATACAACA	CGGGAOCTTG	6550
CACACCCCTC	CCGGCGCCCA	ACTATTCAG	GGGCTATGG	CGGGTGGCTG	6600
CTGAGGAGTA	CGTGGAGGTT	ACGGGTGTGG	GGGATTTCCA	CTACGTGACG	6650
GGCATGACCA	CTGACAACGT	AAAGTGCCCA	TGCCAGGTTT	CGCCCCCGGA	6700
ATTCTTCAAG	GAGGTGGATG	GAGTGCGGTT	GCACAGGTAC	GCTCCGGGGT	6750
GCAAACCTCT	TCTACGGGAG	GACGTACAGT	TCCAGGTGGG	GCTCAACCAA	6800
TACTTGGTGG	GGTCGCAGCT	CCCATGCGAG	CCCGAACCGG	ACGTACACAGT	6850
GCTTACTTCC	ATGCTACCCG	ATCCCTCCCA	CATTACAGCA	GAGACGGCTA	6900
AGCGTAGGCT	GGCTAGAGGG	TCTCCCCCCT	CTTTAGCCAG	CTCATCAGCT	6950
AGCCAGTTGT	CTGCGCCTTC	TTTGAAGGCG	ACATGCACTA	CCACCATGA	7000
CTCCCCGGAC	GCTGACCTCA	TGAGGGCCAA	CCTCTTGTGG	CGGCAGGAGA	7050
TGGGCGGAAA	CATCACTGGC	GTGGAGTCAG	AGAATAAGGT	AGTAATTCTG	7100
GACTCTTTGG	AACCGCTTCA	CGCGGAGGGG	GATGAGAGGG	AGATATCCGT	7150
CGGGGCGGAG	ATCCTGCGAA	AATCCAGGAA	GTTCOCTCA	GCGTTGCCCC	7200
TATGGGCACG	CCCGGACTAC	AATCCTCCAC	TGCTAGAGTC	CTGGAAGGAC	7250
CCGGACTACG	TCCCTCCGGT	GGTACACGGA	TGCCATTGTC	CACCTACCAA	7300
GGCTCCTCCA	ATACCACCTC	CACGGAGAAA	GAGGAAGGTT	GTCCTGACAG	7350
AATCCAATGT	GTCTTCTGCC	TTGGCGGAGC	TGCGCACTAA	GACCTTCGGT	7400
AGCTCCGGAT	CGTGGCGCGT	TGATAGCGGC	ACGGCGAOCG	CCCTTCCTGA	7450
CCTGGCCTCC	GACGACGGTG	ACAAAGGATC	CGACGTTGAG	TGTTACTCCT	7500
CCATGCCCCC	CCTTGAAGGG	GAGCCGGGGG	ACCCCGATCT	CAGCGACGGG	7550
TCTTGGICTA	CCGTGAGTGA	GGAGGCTAGT	GAGGATGTGG	TCTGCTGCTC	7600

FIG. 7D

SUBSTITUTE SHEET (RULE 26)

HC-J4

10	20	30	40	50	
1234567890	1234567890	1234567890	1234567890	1234567890	
AATGTCTAT	ACGTGGACAG	GCGCCCTGAT	CACGCCATGC	GCTGCGGAGG	7650
AAAGTAACT	GCCCATCAAC	CCGTGTAGCA	ACTCTTTGCT	GCGTCAACCAC	7700
AACATGGTCT	ACGCCACAAC	ATCCCGCAGC	GCAAGCCTCC	GCCAGAAGAA	7750
GGTCACTTT	GACAGATTGC	AAGTCTGGA	TGATCATTAC	CGGCACTTAC	7800
TCAAGGAGAT	GAAGGCGAAG	GCGTCCACAG	TTAAGGCTAA	GCTTCTATCT	7850
ATAGAGGAGG	CCTGCAAGCT	GACGCCCCCA	CATTGGGCA	AATCCAAATT	7900
TGGCTATGGG	GCAAAGGACG	TCCGGAACCT	ATCCAGCAGG	GCGTTAACC	7950
ACATCCGCTC	CGTGTGGGAG	GACTTGCTGG	AAGACACTGA	AACAACAATT	8000
GACACCAACA	TCATGGCAAA	AAGTGAGGTT	TTCTGGGTCC	AACCAGAGAA	8050
GGGAGGCGC	AAGCCAGCTC	GCTTATCGT	ATTCCAGAC	CTGGGAGTTC	8100
GTGTATGCGA	GAAGATGGCC	CTTTACGACG	TGGTCTCCAC	CCTTCTCCAG	8150
GCCGTGATGG	GCTCTCATA	CGGATTTCAA	TACTCCCCCA	AGCAGCGGGT	8200
CGAGTTCCTG	GTGAATACCT	GGAAATCAAA	GAAATGCGCT	ATGGGCTTCT	8250
CATATGACAC	CCGCTGTTTT	GACTCAACGG	TCACTGAGAG	TGACATTGCT	8300
GTTGAGGAGT	CAATTTACCA	ATGTTGTGAC	TTGGCCCCCG	AGGCCAGACA	8350
GGCCATAAGG	TGGCTCACAG	AGCGGCTTTA	CATCGGGGGT	CCCCTGACTA	8400
ACTCAAAAGG	GCAGAACTGC	GGTTATCGCC	GGTGGCGGCG	AAGTGGCGTG	8450
CTGACGACTA	GCTGCGGTAA	TACCCTCACA	TGTTACTTGA	AGGCCACTGC	8500
AGCCTGTCCA	GCTGCAAAGC	TCCAGGACTG	CACGATGCTC	GTGAACGGAG	8550
ACGACCTTGT	CGTTATCTGT	GAAAGCGCGG	GAACCCAGGA	GGATGCGGCG	8600
GCCCTACGAG	CCTTCACGGA	GGCTATGACT	AGGTATTCGG	CCCCCCCCGG	8650
GGATCCGCCC	CAACCAGAAT	ACGACCTGGA	GCTGATAACA	TCATGTTCTT	8700
CCAATGTGTC	AGTCGCGCAC	GATGCATCTG	GCAAAAGGGT	ATACTACCTC	8750
ACCCGTGACC	CCACCACCCC	CCTTGCACGG	GCTGCGTGGG	AGACAGCTAG	8800
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CCCTATGGGC	AAGGATGATT	CTGATGACTC	ACTTTTCTTC	CATCTTCTTA	8900
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CTACTCCATT	GAGCCACTTG	ACCTACCTCA	GATCATTGAA	CGACTCCATG	9000
GTCTTAGCGC	ATTTACACTC	CACAGTTACT	CTCCAGGTGA	GATCAATAGG	9050
GTGGCTTCAT	GCTTCAGGAA	ACTTGGGGTA	CCACCTTTCG	GAACCTGGAG	9100
ACATCGGGCC	AGAAGTGTCC	GCGCTAAGCT	ACTGTCCAG	GGGGGGAGGG	9150
CCGCCACTTG	TGGCAGATAC	CTCTTTAACT	GGGCAGTAAG	GACCAAGCTT	9200
AAACTCACTC	CAATCCCGGC	CGGTCCAG	CTGGACTTGT	CTGGCTGGTT	9250
CGTGGCTGGT	TACAGCGGGG	GAGACATATA	TCACAGCCTG	TCTCGTGCCC	9300
GACCCGCTG	GTTTCCGTTG	TGCCCTACTC	TACTTTCTGT	AGGGGTAGGC	9350
ATTTACCTGC	TCCCCAACCG	ATGAACGGGG	AGCTAACAC	TCCAGGCTTT	9400
AAGCCATTTC	CTGTTTTTTT	TTTTTTTTTT	TTTTTTTTTT	TCTTTTTTTT	9450
TTCTTTTCT	TTCTTCTTT	TTTCTCTTC	TTTTTCCCTT	CTTTAATGGT	9500

FIG. 7E

SUBSTITUTE SHEET (RULE 26)

10	20	30	40	50	
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GGCTCCATCT	TAGCOCTAGT	CACGGCTAGC	TGIGAAAGGT	COGIGAGCOG	9550
CATGACTGCA	GAGAGTGCTG	ATACTGGCOCT	CCTGTCAGAT	CATGT	9595

FIG. 7F

10	20	30	40	50	
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KASERSQPRG	RRQPIPKARR	PEGRAWAQPG	YFWPLYGNEG	LGWAGALLSP	100
RGSRPSWGPT	DPRRRSRNLG	KVIDILITCGF	ADLMGYIPLV	GAPLGGAARA	150
LAHGVRVLED	GVMYATGNLP	GCSFSIFLLA	LLSCLTIPAS	AYEVRNWSGI	200
YHVINDCSNS	SIVYEADVI	MHTPGCVPCV	QEGNSSROW	ALITPLAARN	250
ASVPTTTIRR	HVDLLVGTA	FCSAMYVGL	CGSIFLVSQ	FTFSPPRHET	300
VQDQNCSTYP	GHSVGHMAW	DMTNWSPTT	ALVVSQLLRI	PQAVVDMVAG	350
AHWGVLAGLA	YYSVMGNWAK	VLIVALLFAG	VDGEIHITGR	VAGHTTSGFT	400
SLFSSGASQK	IQLVNINGSW	HINRTALNCH	DSLQITGFFA	LFYAHKFNSS	450
GCPERMASCR	PIDWFAQGW	PITYTKFNSS	DQPFYQWHA	PRPGVWPAS	500
QVCGPVYCFT	PSFVVGITD	RSGVPTYSWG	ENEIDVMLIN	NIRPPQGNWF	550
GCTWMNSTGF	TKTCGGPPCN	IGGVGNRTLI	CPIDCFRKHP	EATYIKOGSG	600
FWLTPRCLVD	YPYRLWHYPC	TLNFSIFKVR	MYVGGVEHRL	NAACNWIRGE	650
RQNLDRDRS	ELSPILLSTT	EWQILPCAFT	TLPALSTGLI	HLHQNVVDVQ	700
YLYGVGSAFV	SFAIKWEYIL	LLFILLADAR	VCACIWMMLL	IAQAEAALEN	750
LVLNAASVA	GAHGILSFLV	FFCAAWYIKG	RLAPGAAYAF	YGWPFLLLLL	800
LALPPRAYAL	DREMAASCGG	AVLVGLVFLT	LSPYKVFILT	RLIWWLQYFI	850
TRAEAHMQW	VPPLNVRGGR	DAIILLTCAV	HPELIFDITK	LLAILILPLM	900
VLQAGITRVP	YFVRAQGLIR	ACMLVRKVAG	GHYVQVFMK	LGALITGYVY	950
NHLTPLRDWA	HAGLRDLAVA	VEPVVFSAME	TKVTIWGADT	AACGDIILGL	1000
PVSARRGKEI	FLGPADSLEG	QGWRLAPITT	AYSQITRGVL	GCITTSITGR	1050
DKNQVEGEVQ	WSTATQSF	ATCINGVOWT	VYHGAGSKTL	AGPKGPITQM	1100
YTNVDLILVG	WQAPFGARSM	TPCSGSSDL	YLVIRHADVI	PVRRRGDSRG	1150
SLLSRPVSY	LKGSSGGPLL	CPSGHVGVF	RAAVCIRGVA	KAVDFIPVES	1200
METIMRSPVF	TINSTPPAVP	QTFQVAHLHA	PTGSGKSTKV	PAAYAAQGYK	1250
VLVLNPSVAA	TLGFGAYMSK	AHGIDPNIRT	GVRTITITGGS	ITYSTYKFL	1300
ADGGCSGGAY	DIICDECHS	TDSTITLIGI	TVLDQAETAG	ARLVVLATAT	1350
PPGSVTVPHP	NIEEIGLSNN	GETPFYKAI	PTEAIKGRH	LIFCHSKKKC	1400
DELAACKITGL	GLNAVAYYRG	LIVSVIPPIG	DVVVAITDAL	MIGFTIGDFS	1450
VIDQNTCVTQ	TVDFSLDPTF	TIETITVPQD	AVRSQRGR	TGRGRSGIYR	1500
FVTGERPSG	MFDSSVLCEC	YDAGCAWYEL	TPAETSVRLR	AYLNTFGLPV	1550
QCHLEFWES	VFTGLTHIDA	HFLSQIKQAG	DNFPYLVAYQ	ATVCARAQAP	1600
PPSWDQMKC	LIRLKPILHG	PTPLLYRLGA	VQNEVILTHP	ITKYIMACMS	1650
ADLEVVTSTW	VLVGVLAAL	AAYCLTTGSV	VIVGRIILSG	KPAVVPDREV	1700
LYQEFDEMEE	CASQLPYIEQ	GMQLAEQFKQ	KALGLLQIAT	KQAEAAAPVV	1750
ESKWRALETF	WAKHMANFIS	GIQYLAGLST	LEGNPATIASL	MAFTASTTSP	1800
LTTQNTLLFN	ILGGWAAQL	APPSAASAFV	GAGLAGAAGV	SIGLGKVLVD	1850
ILAGYGAGVA	GALVAFKMS	GEVPSTEDLV	NLLPAILSPG	ALVGVVCAA	1900

FIG. 7G

10	20	30	40	50	
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ILRRHVGPGE	GAVQWMNRLI	AFASRGNHVS	PIHYVPESDA	AARVTQILSS	1950
LITTQLLKRL	HQWINECST	PCSGSWLRDV	WDWICTVLTD	FKTWLQSKLL	2000
PRLPGVPFLS	CQRGYKGWAR	GDGIMQTTCP	CGAQIAGHVK	NGSMRIVGPR	2050
TCSNIWHGTF	PINAYTTGFC	TPSPAIFYSR	ALWRVAAEEY	VEVIRVGDFH	2100
YVTGMTIDNV	KCPCQVPAPF	FFTEVDGVR	HRYPACKPL	LREDVTFQVG	2150
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SSASQLSAPS	LKATCTTHHD	SPDADLJEAN	LLWRQEMGGN	TIRVESENKV	2250
VILDSFEPLH	AEGDERELSV	AAETLRKSRK	FPSALPTIWAR	PDYNPPLLES	2300
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RHNMVYATT	SRSASLRQKK	VTFDRLQVLD	DHYRDVLKEM	KAKASTVKAK	2500
LLSTEEACKL	TPPHSAKSKF	GYGAKDVRNL	SSRAVNHIRS	WEDLLEDTE	2550
TPIDTTIMAK	SEVFCVQPEK	GGRKPARLIV	FPDLGVRVCE	KMALYDVST	2600
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ILLAQEQLK	ALDCQIYGAC	YSIEPLDLFQ	IIERLHGLSA	FTLHSYSPGE	2900
INRVASCLRK	LGVPLRTWR	HRARSVRACL	LSQGGRAATC	GRYLFNWAVR	2950
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FIG. 7H

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 Emerson, Suzanne
 Bukh, Jens
 Purcell, Robert

<120> Cloned Genome of Infectious Hepatitis C Viruses of
 Genotype 2a and Uses Thereof

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<140> TBA

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<150> 60/137,693

<151> 1999-06-04

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<213> Hepatitis C virus

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Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro
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Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly
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Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp
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Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Asn Asp Pro
 100 105 110

Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys
 115 120 125

Gly Phe Ala Asp Leu Met Gly Tyr Ile Pro Val Val Gly Ala Pro Leu
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Gly Gly Val Ala Arg Ala Leu Ala His Gly Val Arg Val Leu Glu Asp
 145 150 155 160

Gly Val Asn Phe Ala Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile
 165 170 175

Phe Leu Leu Ala Leu Leu Ser Cys Ile Thr Thr Pro Val Ser Ala Ala
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Glu Val Lys Asn Ile Ser Thr Gly Tyr Met Val Thr Asn Asp Cys Thr
 195 200 205

Asn Asp Ser Ile Thr Trp Gln Leu Gln Ala Ala Val Leu His Val Pro
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Gly Cys Val Pro Cys Glu Lys Val Gly Asn Ala Ser Gln Cys Trp Ile
 225 230 235 240
 Pro Val Ser Pro Asn Val Ala Val Gln Arg Pro Gly Ala Leu Thr Gln
 245 250 255
 Gly Leu Arg Thr His Ile Asp Met Val Val Met Ser Ala Thr Leu Cys
 260 265 270
 Ser Ala Leu Tyr Val Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ala
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 Gln Met Phe Ile Val Ser Pro Gln His His Trp Phe Val Gln Asp Cys
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 Asn Cys Ser Ile Tyr Pro Gly Thr Ile Thr Gly His Arg Met Ala Trp
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 Asp Met Met Met Asn Trp Ser Pro Thr Ala Thr Met Ile Leu Ala Tyr
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 Ala Met Arg Val Pro Glu Val Ile Ile Asp Ile Ile Ser Gly Ala His
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 Trp Gly Val Met Phe Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp
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 Ala Lys Val Val Val Ile Leu Leu Leu Ala Ala Gly Val Asp Ala Arg
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 Thr His Thr Val Gly Gly Ser Ala Ala Gln Thr Thr Gly Arg Leu Thr
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 Ser Leu Phe Asp Met Gly Pro Arg Gln Lys Ile Gln Leu Val Asn Thr
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 Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser
 420 425 430
 Leu His Thr Gly Phe Ile Ala Ser Leu Phe Tyr Thr His Ser Phe Asn
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 Ser Ser Gly Cys Pro Glu Arg Met Ser Ala Cys Arg Ser Ile Glu Ala
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 Phe Arg Val Gly Trp Gly Ala Leu Gln Tyr Glu Asp Asn Val Thr Asn
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Pro Glu Asp Met Arg Pro Tyr Cys Trp His Tyr Pro Pro Arg Gln Cys
 485 490 495

Gly Val Val Ser Ala Lys Thr Val Cys Gly Pro Val Tyr Cys Phe Thr
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Pro Ser Pro Val Val Val Gly Thr Thr Asp Arg Leu Gly Ala Pro Thr
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Tyr Thr Trp Gly Glu Asn Glu Thr Asp Val Phe Leu Leu Asn Ser Thr
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Arg Pro Pro Leu Gly Ser Trp Phe Gly Cys Thr Trp Met Asn Ser Ser
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Gly Tyr Thr Lys Thr Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp
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Phe Asn Ala Ser Thr Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys
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His Pro Asp Thr Thr Tyr Leu Lys Cys Gly Ser Gly Pro Trp Leu Thr
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Pro Arg Cys Leu Ile Asp Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys
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Glu His Arg Leu Thr Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys
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Asn Leu Glu Asp Arg Asp Arg Ser Gln Leu Ser Pro Leu Leu His Ser
 660 665 670

Thr Thr Glu Trp Ala Ile Leu Pro Cys Ser Tyr Ser Asp Leu Pro Ala
 675 680 685

Leu Ser Thr Gly Leu Leu His Leu His Gln Asn Ile Val Asp Val Gln
 690 695 700

Phe Met Tyr Gly Leu Ser Pro Ala Leu Thr Lys Tyr Ile Val Arg Trp
 705 710 715 720

Glu Trp Val Ile Leu Leu Phe Leu Leu Leu Ala Asp Ala Arg Val Cys
 725 730 735

Ala Cys Leu Trp Met Leu Ile Leu Leu Gly Gln Ala Glu Ala Ala Leu			
740	745	750	
Glu Lys Leu Val Ile Leu His Ala Ala Ser Ala Ala Ser Cys Asn Gly			
755	760	765	
Phe Leu Tyr Phe Val Ile Phe Phe Val Ala Ala Trp Tyr Ile Lys Gly			
770	775	780	
Arg Val Val Pro Leu Ala Thr Tyr Ser Leu Thr Gly Leu Trp Ser Phe			
785	790	795	800
Ser Leu Leu Leu Leu Ala Leu Pro Gln Gln Ala Tyr Ala Tyr Asp Ala			
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Ser Val His Gly Gln Ile Gly Ala Ala Leu Leu Val Met Ile Thr Leu			
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Phe Thr Leu Thr Pro Gly Tyr Lys Thr Leu Leu Ser Arg Phe Leu Trp			
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Trp Leu Cys Tyr Leu Leu Thr Leu Gly Glu Ala Met Val Gln Glu Trp			
	850	855	860
Ala Pro Pro Met Gln Val Arg Gly Gly Arg Asp Gly Ile Ile Trp Ala			
865	870	875	880
Val Ala Ile Phe Tyr Pro Gly Val Val Phe Asp Ile Thr Lys Trp Leu			
	885	890	895
Leu Ala Val Leu Gly Pro Ala Tyr Leu Leu Lys Gly Ala Leu Thr Arg			
	900	905	910
Val Pro Tyr Phe Val Arg Ala His Ala Leu Leu Arg Met Cys Thr Met			
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Ala Arg His Leu Ala Gly Gly Arg Tyr Val Gln Met Ala Leu Leu Ala			
	930	935	940
Leu Gly Arg Trp Thr Gly Thr Tyr Ile Tyr Asp His Leu Thr Pro Met			
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Ser Asp Trp Ala Ala Ser Gly Leu Arg Asp Leu Ala Val Ala Val Glu			
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Pro Ile Ile Phe Ser Pro Met Glu Lys Lys Val Ile Val Trp Gly Ala			
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Glu Thr Ala Ala Cys Gly Asp Ile Leu His Gly Leu Pro Val Ser Ala
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Arg Leu Gly Arg Glu Val Leu Leu Gly Pro Ala Asp Gly Tyr Thr Ser
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Lys Gly Trp Ser Leu Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr
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Arg Gly Leu Leu Gly Thr Ile Val Val Ser Met Thr Gly Arg Asp Lys
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Thr Glu Gln Ala Gly Glu Ile Gln Val Leu Ser Thr Val Thr Gln Ser
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Phe Leu Gly Thr Ser Ile Ser Gly Val Leu Trp Thr Val Tyr His Gly
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Ala Gly Asn Lys Thr Leu Ala Gly Ser Arg Gly Pro Val Thr Gln Met
 1090 1095 1100

Tyr Ser Ser Ala Glu Gly Asp Leu Val Gly Trp Pro Ser Pro Pro Gly
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Pro Val Glu Thr Leu Asp Ile Val Thr Arg Ser Pro Thr Phe Ser Asp
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Asn Ser Thr Pro Pro Ala Val Pro Gln Thr Tyr Gln Val Gly Tyr Leu
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His Ala Pro Thr Gly Ser Gly Lys Ser Thr Lys Val Pro Val Ala Tyr
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Thr Leu Gly Phe Gly Ala Tyr Leu Ser Lys Ala His Gly Ile Asn Pro
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Asn Ile Arg Thr Gly Val Arg Thr Val Thr Thr Gly Ala Pro Ile Thr
 1285 1290 1295

Tyr Ser Thr Tyr Gly Lys Phe Leu Ala Asp Gly Gly Cys Ala Gly Gly
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Ala Tyr Asp Ile Ile Ile Cys Asp Glu Cys His Ala Val Asp Ser Thr
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Thr Ile Leu Gly Ile Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly
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Val Arg Leu Thr Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr
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Thr Pro His Pro Asn Ile Glu Glu Val Ala Leu Gly Gln Glu Gly Glu
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Ile Pro Phe Tyr Gly Arg Ala Ile Pro Leu Ser Tyr Ile Lys Gly Gly
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Arg His Leu Ile Phe Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala
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Ala Ala Leu Arg Gly Met Gly Leu Asn Ser Val Ala Tyr Tyr Arg Gly
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Leu Asp Val Ser Val Ile Pro Thr Gln Gly Asp Val Val Val Val Ala
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Thr Asp Ala Leu Met Thr Gly Tyr Thr Gly Asp Phe Asp Ser Val Ile
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Asp Cys Asn Val Ala Val Thr Gln Val Val Asp Phe Ser Leu Asp Pro
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Thr Phe Thr Ile Thr Thr Gln Ile Val Pro Gln Asp Ala Val Ser Arg
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Ser Gln Arg Arg Gly Arg Thr Gly Arg Gly Arg Leu Gly Ile Tyr Arg
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Tyr Val Ser Thr Gly Glu Arg Ala Ser Gly Met Phe Asp Ser Val Val
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Leu Cys Glu Cys Tyr Asp Ala Gly Ala Ala Trp Tyr Glu Leu Thr Pro
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Ser Glu Thr Thr Val Arg Leu Arg Ala Tyr Phe Asn Thr Pro Gly Leu
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Pro Val Cys Gln Asp His Leu Glu Phe Trp Glu Ala Val Phe Thr Gly
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Leu Thr His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly
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Glu Asn Phe Ala Tyr Leu Thr Ala Tyr Gln Ala Thr Val Cys Ala Arg
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Ala Lys Ala Pro Pro Pro Ser Trp Asp Val Met Trp Lys Cys Leu Thr
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Arg Leu Lys Pro Thr Leu Val Gly Pro Thr Pro Leu Leu Tyr Arg Leu
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Gly Ser Val Thr Asn Glu Val Thr Leu Thr His Pro Val Thr Lys Tyr
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Val Leu Ala Gly Gly Val Leu Ala Ala Val Ala Ala Tyr Cys Leu Ala
 1665 1670 1675 1680

Thr Gly Cys Val Cys Ile Ile Gly Arg Leu His Ile Asn Gln Arg Ala
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Val Val Ala Pro Asp Lys Glu Val Leu Tyr Glu Ala Phe Asp Glu Met
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Ala Glu Met Leu Lys Ser Lys Ile Gln Gly Leu Leu Gln Gln Ala Ser
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Lys Gln Ala Gln Asp Ile Gln Pro Thr Val Gln Ala Ser Trp Pro Lys
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Val Glu Gln Phe Trp Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile			
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Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Val Ala			
1780	1785	1790	
Ser Met Met Ala Phe Ser Ala Ala Leu Thr Ser Pro Leu Ser Thr Ser			
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Thr Thr Ile Leu Leu Asn Ile Leu Gly Gly Trp Leu Ala Ser Gln Ile			
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Ala Gly Tyr Gly Ala Gly Ile Ser Gly Ala Leu Val Ala Phe Lys Ile			
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Met Ser Gly Glu Lys Pro Ser Met Glu Asp Val Val Asn Leu Leu Pro			
1875	1880	1885	
Gly Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Ile Cys Ala Ala			
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Asn Arg Leu Ile Ala Phe Ala Ser Arg Gly Asn His Val Ala Pro Thr			
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His Tyr Val Thr Glu Ser Asp Ala Ser Gln Arg Val Thr Gln Leu Leu			
1940	1945	1950	
Gly Ser Leu Thr Ile Thr Ser Leu Leu Arg Arg Leu His Asn Trp Ile			
1955	1960	1965	
Thr Glu Asp Cys Pro Ile Pro Cys Gly Gly Ser Trp Leu Arg Asp Val			
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Trp Asp Trp Val Cys Thr Ile Leu Thr Asp Phe Lys Asn Trp Leu Thr			
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Ser Lys Leu Phe Pro Lys Met Pro Gly Leu Pro Phe Val Ser Cys Gln			
2005	2010	2015	

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Glu Asp Ser Ile Gly Asp Ala Leu Gln Gln Leu Ala Ile Lys Ser Phe
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Gly Gln Pro Pro Pro Ser Gly Asp Ser Gly Leu Ser Thr Gly Ala Gly
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Met Gly Phe Ser Tyr Asp Thr Arg Cys Phe Asp Ser Thr Val Thr Glu
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Glu Glu Ala His Thr Ala Ile His Ser Leu Thr Glu Arg Leu Tyr Val
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Cys Tyr Val Lys Ala Leu Ala Ala Cys Lys Ala Ala Gly Ile Ile Ala
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Pro Thr Met Leu Val Cys Gly Asp Asp Leu Val Val Ile Ser Glu Ser
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Ser Trp Leu Gly Asn Ile Ile Gln Tyr Ala Pro Thr Ile Trp Ala Arg
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Met Val Leu Met Thr His Phe Phe Ser Ile Leu Met Ala Gln Asp Thr
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Ser Ala Leu Arg Lys Leu Gly Ala Pro Pro Leu Arg Ala Trp Lys Ser
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Ala Val Cys Gly Arg Tyr Leu Phe Asn Trp Ala Val Lys Thr Lys Leu
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Lys Leu Thr Pro Leu Pro Glu Ala Arg Leu Leu Asp Leu Ser Ser Trp
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Phe Thr Val Gly Ala Gly Gly Gly Asp Ile Tyr His Ser Val Ser Arg
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<211> 3015

<212> PRT

<213> Hepatitis C virus

<400> 4

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 35 40 45

Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro
 50 55 60

Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly
 65 70 75 80

Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp

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Thr Thr Glu Trp Ala Ile Leu Pro Cys Ser Tyr Ser Asp Leu Pro Ala		
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<212> PRT

<213> Hepatitis C virus

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 35 40 45

Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro
 50 55 60

Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly
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Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp
 85 90 95

Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Asn Asp Pro
 100 105 110

Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys
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Gly Phe Ala Asp Leu Met Gly Tyr Ile Pro Val Val Gly Ala Pro Leu
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Gly Gly Val Ala Arg Ala Leu Ala His Gly Val Arg Val Leu Glu Asp
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Gly Val Asn Phe Ala Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile
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Phe Leu Leu Ala Leu Leu Ser Cys Ile Thr Thr Pro Val Ser Ala Ala
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Glu Val Lys Asn Ile Ser Thr Gly Tyr Met Val Thr Asn Asp Cys Thr
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Asn Asp Ser Ile Thr Trp Gln Leu Gln Ala Ala Val Leu His Val Pro
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Gly Cys Val Pro Cys Glu Lys Val Gly Asn Ala Ser Gln Cys Trp Ile
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Pro Val Ser Pro Asn Val Ala Val Gln Arg Pro Gly Ala Leu Thr Gln
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Gly Leu Arg Thr His Ile Asp Met Val Val Met Ser Ala Thr Leu Cys
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Ser Ala Leu Tyr Val Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ala
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Gln Met Phe Ile Val Ser Pro Gln His His Trp Phe Val Gln Asp Cys
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Asn Cys Ser Ile Tyr Pro Gly Thr Ile Thr Gly His Arg Met Ala Trp
 305 310 315 320

Asp Met Met Met Asn Trp Ser Pro Thr Ala Thr Met Ile Leu Ala Tyr
 325 330 335

Ala Met Arg Val Pro Glu Val Ile Ile Asp Ile Ile Ser Gly Ala His
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Trp Gly Val Met Phe Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp
 355 360 365

Ala Lys Val Val Val Ile Leu Leu Leu Ala Ala Gly Val Asp Ala Arg
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Thr His Thr Val Gly Gly Ser Ala Ala Gln Thr Thr Gly Arg Leu Thr
 385 390 395 400

Ser Leu Phe Asp Met Gly Pro Arg Gln Lys Ile Gln Leu Val Asn Thr
 405 410 415

Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser
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Leu His Thr Gly Phe Ile Ala Ser Leu Phe Tyr Thr His Ser Phe Asn
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Ser Ser Gly Cys Pro Glu Arg Met Ser Ala Cys Arg Ser Ile Glu Ala
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Phe Arg Val Gly Trp Gly Ala Leu Gln Tyr Glu Asp Asn Val Thr Asn
 465 470 475 480

Pro Glu Asp Met Arg Pro Tyr Cys Trp His Tyr Pro Pro Arg Gln Cys
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Gly Val Val Ser Ala Lys Thr Val Cys Gly Pro Val Tyr Cys Phe Thr
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Pro Ser Pro Val Val Val Gly Thr Thr Asp Arg Leu Gly Ala Pro Thr
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Tyr Thr Trp Gly Glu Asn Glu Thr Asp Val Phe Leu Leu Asn Ser Thr
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Arg Pro Pro Leu Gly Ser Trp Phe Gly Cys Thr Trp Met Asn Ser Ser
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Gly Tyr Thr Lys Thr Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp
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Phe Asn Ala Ser Thr Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys
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His Pro Asp Thr Thr Tyr Leu Lys Cys Gly Ser Gly Pro Trp Leu Thr
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Pro Arg Cys Leu Ile Asp Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys
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Thr Val Asn Tyr Thr Ile Phe Lys Ile Arg Met Tyr Val Gly Gly Val
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Glu His Arg Leu Thr Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys
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Asn Leu Glu Asp Arg Asp Arg Ser Gln Leu Ser Pro Leu Leu His Ser
 660 665 670

Thr Thr Glu Trp Ala Ile Leu Pro Cys Ser Tyr Ser Asp Leu Pro Ala
 675 680 685

Leu Ser Thr Gly Leu Leu His Leu His Gln Asn Ile Val Asp Val Gln
 690 695 700

Phe Met Tyr Gly Leu Ser Pro Ala Leu Thr Lys Tyr Ile Val Arg Trp
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Glu Trp Val Ile Leu Leu Phe Leu Leu Leu Ala Asp Ala Arg Val Cys
 725 730 735

Ala Cys Leu Trp Met Leu Ile Leu Leu Gly Gln Ala Glu Ala Ala Leu	740	745	750
Glu Asn Leu Val Ile Leu Asn Ala Ala Ser Leu Ala Gly Thr His Gly	755	760	765
Leu Val Ser Phe Leu Val Phe Phe Cys Phe Ala Trp Tyr Leu Lys Gly	770	775	780
Arg Trp Val Pro Gly Ala Val Tyr Ala Leu Tyr Gly Met Trp Pro Leu	785	790	795
Leu Leu Leu Leu Leu Ala Leu Pro Gln Arg Ala Tyr Ala Leu Asp Thr	805	810	815
Glu Val Ala Ala Ser Cys Gly Gly Val Val Leu Val Gly Leu Met Ala	820	825	830
Leu Thr Leu Ser Pro Tyr Tyr Lys Arg Tyr Ile Ser Trp Cys Met Trp	835	840	845
Trp Leu Gln Tyr Phe Leu Thr Arg Val Glu Ala Gln Leu His Val Trp	850	855	860
Val Pro Pro Leu Asn Val Arg Gly Gly Arg Asp Ala Val Ile Leu Leu	865	870	875
Met Cys Val Val His Pro Thr Leu Val Phe Asp Ile Thr Lys Leu Leu	885	890	895
Leu Ala Ile Phe Gly Pro Leu Trp Ile Leu Gln Ala Ser Leu Leu Lys	900	905	910
Val Pro Tyr Phe Val Arg Val Gln Gly Leu Leu Arg Ile Cys Ala Leu	915	920	925
Ala Arg Lys Ile Ala Gly Gly His Tyr Val Gln Met Ala Ile Ile Lys	930	935	940
Leu Gly Ala Leu Thr Gly Thr Tyr Val Tyr Asn His Leu Thr Pro Leu	945	950	955
Arg Asp Trp Ala His Asn Gly Leu Arg Asp Leu Ala Val Ala Val Glu	965	970	975
Pro Val Val Phe Ser Arg Met Glu Thr Lys Leu Ile Thr Trp Gly Ala	980	985	990

Asp Thr Ala Ala Cys Gly Asp Ile Ile Asn Gly Leu Pro Val Ser Ala
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Lys Gly Trp Arg Leu Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr
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Phe Leu Ala Thr Cys Ile Asn Gly Val Cys Trp Thr Val Tyr His Gly
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Ala Gly Thr Arg Thr Ile Ala Ser Pro Lys Gly Pro Val Ile Gln Met
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Tyr Thr Asn Val Asp Gln Asp Leu Val Gly Trp Pro Ala Pro Gln Gly
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His Ala Pro Thr Gly Ser Gly Lys Ser Thr Lys Val Pro Ala Ala Tyr
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Ala Ala Gln Gly Tyr Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala
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Thr Leu Gly Phe Gly Ala Tyr Met Ser Lys Ala His Gly Val Asp Pro
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Asn Ile Arg Thr Gly Val Arg Thr Ile Thr Thr Gly Ser Pro Ile Thr
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Tyr Ser Thr Tyr Gly Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly
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Ala Tyr Asp Ile Ile Ile Cys Asp Glu Cys His Ser Thr Asp Ala Thr
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Ser Ile Leu Gly Ile Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly
 1330 1335 1340

Ala Arg Leu Val Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr
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Val Ser His Pro Asn Ile Glu Glu Val Ala Leu Ser Thr Thr Gly Glu
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Ile Pro Phe Tyr Gly Lys Ala Ile Pro Leu Glu Val Ile Lys Gly Gly
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Arg His Leu Ile Phe Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala
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Ala Lys Leu Val Ala Leu Gly Ile Asn Ala Val Ala Tyr Tyr Arg Gly
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Leu Asp Val Ser Val Ile Pro Thr Ser Gly Asp Val Val Val Val Ser
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Thr Asp Ala Leu Met Thr Gly Phe Thr Gly Asp Phe Asp Ser Val Ile
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Asp Cys Asn Thr Cys Val Thr Gln Thr Val Asp Phe Ser Leu Asp Pro
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Thr Phe Thr Ile Glu Thr Thr Thr Leu Pro Gln Asp Ala Val Ser Arg
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Thr Gln Arg Arg Gly Arg Thr Gly Arg Gly Lys Pro Gly Ile Tyr Arg
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Ala Glu Thr Thr Val Arg Leu Arg Ala Tyr Met Asn Thr Pro Gly Leu
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Pro Val Cys Gln Asp His Leu Glu Phe Trp Glu Gly Val Phe Thr Gly
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Leu Thr His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly
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Glu Asn Phe Pro Tyr Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg
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Ala Gln Ala Pro Pro Pro Ser Trp Asp Gln Met Trp Lys Cys Leu Ile
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Arg Leu Lys Pro Thr Leu His Gly Pro Thr Pro Leu Leu Tyr Arg Leu
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Gly Ala Val Gln Asn Glu Val Thr Leu Thr His Pro Ile Thr Lys Tyr
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Ile Met Thr Cys Met Ser Ala Asp Leu Glu Val Val Thr Ser Thr Trp
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Val Leu Val Gly Gly Val Leu Ala Ala Leu Ala Ala Tyr Cys Leu Ser
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Thr Gly Cys Val Val Ile Val Gly Arg Ile Val Leu Ser Gly Lys Pro
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Glu Glu Cys Ser Gln His Leu Pro Tyr Ile Glu Gln Gly Met Met Leu
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Ala Glu Gln Phe Lys Gln Lys Ala Leu Gly Leu Leu Gln Thr Ala Ser
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Arg His Ala Glu Val Ile Thr Pro Ala Val Gln Thr Asn Trp Gln Lys
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Leu Glu Val Phe Trp Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile
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Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Ile Ala
 1780 1785 1790

Ser Leu Met Ala Phe Thr Ala Ala Val Thr Ser Pro Leu Thr Thr Gly
 1795 1800 1805

Gln Thr Leu Leu Phe Asn Ile Leu Gly Gly Trp Val Ala Ala Gln Leu
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Ala Ala Pro Gly Ala Ala Thr Ala Phe Val Gly Ala Gly Leu Ala Gly
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Ala Ala Ile Gly Ser Val Gly Leu Gly Lys Val Leu Val Asp Ile Leu
 1845 1850 1855

Ala Gly Tyr Gly Ala Gly Val Ala Gly Ala Leu Val Ala Phe Lys Ile
 1860 1865 1870

Met Ser Gly Glu Val Pro Ser Thr Glu Asp Leu Val Asn Leu Leu Pro
 1875 1880 1885

Ala Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Val Cys Ala Ala
 1890 1895 1900

Ile Leu Arg Arg His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met
 1905 1910 1915 1920

Asn Arg Leu Ile Ala Phe Ala Ser Arg Gly Asn His Val Ser Pro Thr
 1925 1930 1935

His Tyr Val Pro Glu Ser Asp Ala Ala Ala Arg Val Thr Ala Ile Leu
 1940 1945 1950

Ser Ser Leu Thr Val Thr Gln Leu Leu Arg Arg Leu His Gln Trp Ile
 1955 1960 1965

Ser Ser Glu Cys Thr Thr Pro Cys Ser Gly Ser Trp Leu Arg Asp Ile
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Trp Asp Trp Ile Cys Glu Val Leu Ser Asp Phe Lys Thr Trp Leu Lys
 1985 1990 1995 2000

Ala Lys Leu Met Pro Gln Leu Pro Gly Ile Pro Phe Val Ser Cys Gln
 2005 2010 2015

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Ile Arg Arg Val Gly Asp Phe His Tyr Val Ser Gly Met Thr Thr Asp			
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Asn Leu Lys Cys Pro Cys Gln Ile Pro Ser Pro Glu Phe Phe Thr Glu			
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Leu Asp Gly Val Arg Leu His Arg Phe Ala Pro Pro Cys Lys Pro Leu			
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Leu Arg Glu Glu Val Ser Phe Arg Val Gly Leu His Glu Tyr Pro Val			
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Gly Ser Gln Leu Pro Cys Glu Pro Glu Pro Asp Val Ala Val Leu Thr			
2165	2170	2175	
Ser Met Leu Thr Asp Pro Ser His Ile Thr Ala Glu Ala Ala Gly Arg			
2180	2185	2190	
Arg Leu Ala Arg Gly Ser Pro Pro Ser Met Ala Ser Ser Ser Ala Ser			
2195	2200	2205	
Gln Leu Ser Ala Pro Ser Leu Lys Ala Thr Cys Thr Ala Asn His Asp			
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Ser Pro Asp Ala Glu Leu Ile Glu Ala Asn Leu Leu Trp Arg Gln Glu			
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Met Gly Gly Asn Ile Thr Arg Val Glu Ser Glu Asn Lys Val Val Ile			
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Leu Pro Val Trp Ala Arg Pro Asp Tyr Asn Pro Pro Leu Val Glu Thr
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Trp Lys Lys Pro Asp Tyr Glu Pro Pro Val Val His Gly Cys Pro Leu
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Pro Pro Pro Arg Ser Pro Pro Val Pro Pro Pro Arg Lys Lys Arg Thr
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Val Val Leu Thr Glu Ser Thr Leu Ser Thr Ala Leu Ala Glu Leu Ala
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Thr Lys Ser Phe Gly Ser Ser Ser Thr Ser Gly Ile Thr Gly Asp Asn
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Asp Thr Glu Asp Val Val Cys Cys Ser Met Ser Tyr Ser Trp Thr Gly
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Leu Gln Val Leu Asp Ser His Tyr Gln Asp Val Leu Lys Glu Val Lys
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Cys Ser Leu Thr Pro Pro His Ser Ala Lys Ser Lys Phe Gly Tyr Gly
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Ala Lys Asp Val Arg Cys His Ala Arg Lys Ala Val Ala His Ile Asn
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Ser Val Trp Lys Asp Leu Leu Glu Asp Ser Val Thr Pro Ile Asp Thr
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Gly Arg Lys Pro Ala Arg Leu Ile Val Phe Pro Asp Leu Gly Val Arg
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Val Cys Glu Lys Met Ala Leu Tyr Asp Val Val Ser Lys Leu Pro Leu
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Val Glu Phe Leu Val Gln Ala Trp Lys Ser Lys Lys Thr Pro Met Gly
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Ala Arg Val Ala Ile Lys Ser Leu Thr Glu Arg Leu Tyr Val Gly Gly
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Pro Leu Thr Asn Ser Arg Gly Glu Asn Cys Gly Tyr Arg Arg Cys Arg
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Ala Ser Gly Val Leu Thr Thr Ser Cys Gly Asn Thr Leu Thr Cys Tyr
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Ile Lys Ala Arg Ala Ala Cys Arg Ala Ala Gly Leu Gln Asp Cys Thr
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Met Leu Val Cys Gly Asp Asp Leu Val Val Ile Cys Glu Ser Ala Gly
2740 2745 2750

Val Gln Glu Asp Ala Ala Ser Leu Arg Ala Phe Thr Glu Ala Met Thr
2755 2760 2765

Arg Tyr Ser Ala Pro Pro Gly Asp Pro Pro Gln Pro Glu Tyr Asp Leu
2770 2775 2780

Glu Leu Ile Thr Ser Cys Ser Ser Asn Val Ser Val Ala His Asp Gly
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Ala Gly Lys Arg Val Tyr Tyr Leu Thr Arg Asp Pro Thr Thr Pro Leu
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Leu Gly Asn Ile Ile Met Phe Ala Pro Thr Leu Trp Ala Arg Met Ile
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Leu Met Thr His Phe Phe Ser Val Leu Ile Ala Arg Asp Gln Leu Glu
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Gln Ala Leu Asn Cys Glu Ile Tyr Gly Ala Cys Tyr Ser Ile Glu Pro
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Leu Asp Leu Pro Pro Ile Ile Gln Arg Leu His Gly Leu Ser Ala Phe
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Ser Leu His Ser Tyr Ser Pro Gly Glu Ile Asn Arg Val Ala Ala Cys
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Arg Ser Val Arg Ala Arg Leu Leu Ser Arg Gly Gly Arg Ala Ala Ile
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Cys Gly Lys Tyr Leu Phe Asn Trp Ala Val Arg Thr Lys Leu Lys Leu
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Thr Pro Ile Ala Ala Ala Gly Arg Leu Asp Leu Ser Gly Trp Phe Thr
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Ala Gly Tyr Ser Gly Gly Asp Ile Tyr His Ser Val Ser His Ala Arg
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<213> Hepatitis C virus

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Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly
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Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp
85 90 95

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Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys
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900 905 910

Val Pro Tyr Phe Val Arg Val Gln Gly Leu Leu Arg Ile Cys Ala Leu
915 920 925

Ala Arg Lys Ile Ala Gly Gly His Tyr Val Gln Met Ala Ile Ile Lys
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ttcttttttt cttttctttt tcccttcttt aatggtggct ccatcttagc cctagtcacg 9540
gctagctgtg aaaggtccgt gagccgcatg actgcagaga gtgctgatac tggcctctct 9600
gcagatcatg t 9611

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<211> 3015

<212> PRT

<213> Hepatitis C virus

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Met Ser Thr Asn Pro Lys Pro Gln Arg Lys Thr Lys Arg Asn Thr Asn

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Arg Arg Pro Gln Asp Val Lys Phe Pro Gly Gly Gly Gln Ile Val Gly

20 25 30

Gly Val Tyr Leu Leu Pro Arg Arg Gly Pro Arg Leu Gly Val Arg Ala

35 40 45

Thr Arg Lys Thr Ser Glu Arg Ser Gln Pro Arg Gly Arg Arg Gln Pro

50 55 60

Ile Pro Lys Asp Arg Arg Ser Thr Gly Lys Ser Trp Gly Lys Pro Gly

65 70 75 80

Tyr Pro Trp Pro Leu Tyr Gly Asn Glu Gly Leu Gly Trp Ala Gly Trp

85 90 95

Leu Leu Ser Pro Arg Gly Ser Arg Pro Ser Trp Gly Pro Asn Asp Pro

100 105 110

Arg His Arg Ser Arg Asn Val Gly Lys Val Ile Asp Thr Leu Thr Cys

115 120 125

Gly Phe Ala Asp Leu Met Gly Tyr Ile Pro Val Val Gly Ala Pro Leu

130 135 140

Gly Gly Val Ala Arg Ala Leu Ala His Gly Val Arg Val Leu Glu Asp

145 150 155 160

Gly Val Asn Phe Ala Thr Gly Asn Leu Pro Gly Cys Ser Phe Ser Ile

165 170 175

Phe Leu Leu Ala Leu Leu Ser Cys Ile Thr Thr Pro Val Ser Ala Ala

180 185 190

Glu Val Lys Asn Ile Ser Thr Gly Tyr Met Val Thr Asn Asp Cys Thr

195 200 205

Asn Asp Ser Ile Thr Trp Gln Leu Gln Ala Ala Val Leu His Val Pro

210 215 220

Gly Cys Val Pro Cys Glu Lys Val Gly Asn Ala Ser Gln Cys Trp Ile

225	230	235	240
Pro Val Ser Pro Asn Val Ala Val Gln Arg Pro Gly Ala Leu Thr Gln			
	245	250	255
Gly Leu Arg Thr His Ile Asp Met Val Val Met Ser Ala Thr Leu Cys			
	260	265	270
Ser Ala Leu Tyr Val Gly Asp Leu Cys Gly Gly Val Met Leu Ala Ala			
	275	280	285
Gln Met Phe Ile Val Ser Pro Gln His His Trp Phe Val Gln Asp Cys			
	290	295	300
Asn Cys Ser Ile Tyr Pro Gly Thr Ile Thr Gly His Arg Met Ala Trp			
305	310	315	320
Asp Met Met Met Asn Trp Ser Pro Thr Ala Thr Met Ile Leu Ala Tyr			
	325	330	335
Ala Met Arg Val Pro Glu Val Ile Ile Asp Ile Ile Ser Gly Ala His			
	340	345	350
Trp Gly Val Met Phe Gly Leu Ala Tyr Phe Ser Met Gln Gly Ala Trp			
	355	360	365
Ala Lys Val Val Val Ile Leu Leu Leu Ala Ala Gly Val Asp Ala Arg			
	370	375	380
Thr His Thr Val Gly Gly Ser Ala Ala Gln Thr Thr Gly Arg Leu Thr			
385	390	395	400
Ser Leu Phe Asp Met Gly Pro Arg Gln Lys Ile Gln Leu Val Asn Thr			
	405	410	415
Asn Gly Ser Trp His Ile Asn Arg Thr Ala Leu Asn Cys Asn Asp Ser			
	420	425	430
Leu His Thr Gly Phe Ile Ala Ser Leu Phe Tyr Thr His Ser Phe Asn			
	435	440	445
Ser Ser Gly Cys Pro Glu Arg Met Ser Ala Cys Arg Ser Ile Glu Ala			
	450	455	460
Phe Arg Val Gly Trp Gly Ala Leu Gln Tyr Glu Asp Asn Val Thr Asn			
465	470	475	480
Pro Glu Asp Met Arg Pro Tyr Cys Trp His Tyr Pro Pro Arg Gln Cys			

485	490	495
Gly Val Val Ser Ala Lys Thr Val Cys Gly Pro Val Tyr Cys Phe Thr		
500	505	510
Pro Ser Pro Val Val Val Gly Thr Thr Asp Arg Leu Gly Ala Pro Thr		
515	520	525
Tyr Thr Trp Gly Glu Asn Glu Thr Asp Val Phe Leu Leu Asn Ser Thr		
530	535	540
Arg Pro Pro Leu Gly Ser Trp Phe Gly Cys Thr Trp Met Asn Ser Ser		
545	550	555
Gly Tyr Thr Lys Thr Cys Gly Ala Pro Pro Cys Arg Thr Arg Ala Asp		
565	570	575
Phe Asn Ala Ser Thr Asp Leu Leu Cys Pro Thr Asp Cys Phe Arg Lys		
580	585	590
His Pro Asp Thr Thr Tyr Leu Lys Cys Gly Ser Gly Pro Trp Leu Thr		
595	600	605
Pro Arg Cys Leu Ile Asp Tyr Pro Tyr Arg Leu Trp His Tyr Pro Cys		
610	615	620
Thr Val Asn Tyr Thr Ile Phe Lys Ile Arg Met Tyr Val Gly Gly Val		
625	630	635
Glu His Arg Leu Thr Ala Ala Cys Asn Phe Thr Arg Gly Asp Arg Cys		
645	650	655
Asn Leu Glu Asp Arg Asp Arg Ser Gln Leu Ser Pro Leu Leu His Ser		
660	665	670
Thr Thr Glu Trp Ala Ile Leu Pro Cys Ser Tyr Ser Asp Leu Pro Ala		
675	680	685
Leu Ser Thr Gly Leu Leu His Leu His Gln Asn Ile Val Asp Val Gln		
690	695	700
Phe Met Tyr Gly Leu Ser Pro Ala Leu Thr Lys Tyr Ile Val Arg Trp		
705	710	715
Glu Trp Val Ile Leu Leu Phe Leu Leu Leu Ala Asp Ala Arg Val Cys		
725	730	735
Ala Cys Leu Trp Met Leu Ile Leu Leu Gly Gln Ala Glu Ala Ala Leu		

740	745	750
Glu Asn Leu Val Ile Leu Asn Ala Ala Ser Leu Ala Gly Thr His Gly		
755	760	765
Leu Val Ser Phe Leu Val Phe Phe Cys Phe Ala Trp Tyr Leu Lys Gly		
770	775	780
Arg Trp Val Pro Gly Ala Val Tyr Ala Leu Tyr Gly Met Trp Pro Leu		
785	790	795 800
Leu Leu Leu Leu Leu Ala Leu Pro Gln Arg Ala Tyr Ala Leu Asp Thr		
805	810	815
Glu Val Ala Ala Ser Cys Gly Gly Val Val Leu Val Gly Leu Met Ala		
820	825	830
Leu Thr Leu Ser Pro Tyr Tyr Lys Arg Tyr Ile Ser Trp Cys Met Trp		
835	840	845
Trp Leu Gln Tyr Phe Leu Thr Arg Val Glu Ala Gln Leu His Val Trp		
850	855	860
Val Pro Pro Leu Asn Val Arg Gly Gly Arg Asp Ala Val Ile Leu Leu		
865	870	875 880
Met Cys Val Val His Pro Thr Leu Val Phe Asp Ile Thr Lys Leu Leu		
885	890	895
Leu Ala Ile Phe Gly Pro Leu Trp Ile Leu Gln Ala Ser Leu Leu Lys		
900	905	910
Val Pro Tyr Phe Val Arg Val Gln Gly Leu Leu Arg Ile Cys Ala Leu		
915	920	925
Ala Arg Lys Ile Ala Gly Gly His Tyr Val Gln Met Ala Ile Ile Lys		
930	935	940
Leu Gly Ala Leu Thr Gly Thr Tyr Val Tyr Asn His Leu Thr Pro Leu		
945	950	955 960
Arg Asp Trp Ala His Asn Gly Leu Arg Asp Leu Ala Val Ala Val Glu		
965	970	975
Pro Val Val Phe Ser Arg Met Glu Thr Lys Leu Ile Thr Trp Gly Ala		
980	985	990
Asp Thr Ala Ala Cys Gly Asp Ile Ile Asn Gly Leu Pro Val Ser Ala		

995	1000	1005
Arg Arg Gly Gln Glu Ile Leu Leu Gly Pro Ala Asp Gly Met Val Ser		
1010	1015	1020
Lys Gly Trp Arg Leu Leu Ala Pro Ile Thr Ala Tyr Ala Gln Gln Thr		
1025	1030	1035 1040
Arg Gly Leu Leu Gly Cys Ile Ile Thr Ser Leu Thr Gly Arg Asp Lys		
1045	1050	1055
Asn Gln Val Glu Gly Glu Val Gln Ile Val Ser Thr Ala Thr Gln Thr		
1060	1065	1070
Phe Leu Ala Thr Cys Ile Asn Gly Val Cys Trp Thr Val Tyr His Gly		
1075	1080	1085
Ala Gly Thr Arg Thr Ile Ala Ser Pro Lys Gly Pro Val Ile Gln Met		
1090	1095	1100
Tyr Thr Asn Val Asp Gln Asp Leu Val Gly Trp Pro Ala Pro Gln Gly		
1105	1110	1115 1120
Ser Arg Ser Leu Thr Pro Cys Thr Cys Gly Ser Ser Asp Leu Tyr Leu		
1125	1130	1135
Val Thr Arg His Ala Asp Val Ile Pro Val Arg Arg Arg Gly Asp Ser		
1140	1145	1150
Arg Gly Ser Leu Leu Ser Pro Arg Pro Ile Ser Tyr Leu Lys Gly Ser		
1155	1160	1165
Ser Gly Gly Pro Leu Leu Cys Pro Ala Gly His Ala Val Gly Leu Phe		
1170	1175	1180
Arg Ala Ala Val Cys Thr Arg Gly Val Ala Lys Ala Val Asp Phe Ile		
1185	1190	1195 1200
Pro Val Glu Asn Leu Gly Thr Thr Met Arg Ser Pro Val Phe Thr Asp		
1205	1210	1215
Asn Ser Ser Pro Pro Ala Val Pro Gln Ser Phe Gln Val Ala His Leu		
1220	1225	1230
His Ala Pro Thr Gly Ser Gly Lys Ser Thr Lys Val Pro Ala Ala Tyr		
1235	1240	1245
Ala Ala Gln Gly Tyr Lys Val Leu Val Leu Asn Pro Ser Val Ala Ala		

1250	1255	1260	
Thr Leu Gly Phe Gly Ala Tyr Met Ser Lys Ala His Gly Val Asp Pro			
1265	1270	1275	1280
Asn Ile Arg Thr Gly Val Arg Thr Ile Thr Thr Gly Ser Pro Ile Thr			
1285	1290	1295	
Tyr Ser Thr Tyr Gly Lys Phe Leu Ala Asp Gly Gly Cys Ser Gly Gly			
1300	1305	1310	
Ala Tyr Asp Ile Ile Ile Cys Asp Glu Cys His Ser Thr Asp Ala Thr			
1315	1320	1325	
Ser Ile Leu Gly Ile Gly Thr Val Leu Asp Gln Ala Glu Thr Ala Gly			
1330	1335	1340	
Ala Arg Leu Val Val Leu Ala Thr Ala Thr Pro Pro Gly Ser Val Thr			
1345	1350	1355	1360
Val Ser His Pro Asn Ile Glu Glu Val Ala Leu Ser Thr Thr Gly Glu			
1365	1370	1375	
Ile Pro Phe Tyr Gly Lys Ala Ile Pro Leu Glu Val Ile Lys Gly Gly			
1380	1385	1390	
Arg His Leu Ile Phe Cys His Ser Lys Lys Lys Cys Asp Glu Leu Ala			
1395	1400	1405	
Ala Lys Leu Val Ala Leu Gly Ile Asn Ala Val Ala Tyr Tyr Arg Gly			
1410	1415	1420	
Leu Asp Val Ser Val Ile Pro Thr Ser Gly Asp Val Val Val Val Ser			
1425	1430	1435	1440
Thr Asp Ala Leu Met Thr Gly Phe Thr Gly Asp Phe Asp Ser Val Ile			
1445	1450	1455	
Asp Cys Asn Thr Cys Val Thr Gln Thr Val Asp Phe Ser Leu Asp Pro			
1460	1465	1470	
Thr Phe Thr Ile Glu Thr Thr Thr Leu Pro Gln Asp Ala Val Ser Arg			
1475	1480	1485	
Thr Gln Arg Arg Gly Arg Thr Gly Arg Gly Lys Pro Gly Ile Tyr Arg			
1490	1495	1500	
Phe Val Ala Pro Gly Glu Arg Pro Ser Gly Met Phe Asp Ser Ser Val			

1505	1510	1515	1520
Leu Cys Glu Cys Tyr Asp Ala Gly Cys Ala Trp Tyr Glu Leu Thr Pro			
1525	1530	1535	
Ala Glu Thr Thr Val Arg Leu Arg Ala Tyr Met Asn Thr Pro Gly Leu			
1540	1545	1550	
Pro Val Cys Gln Asp His Leu Glu Phe Trp Glu Gly Val Phe Thr Gly			
1555	1560	1565	
Leu Thr His Ile Asp Ala His Phe Leu Ser Gln Thr Lys Gln Ser Gly			
1570	1575	1580	
Glu Asn Phe Pro Tyr Leu Val Ala Tyr Gln Ala Thr Val Cys Ala Arg			
1585	1590	1595	1600
Ala Gln Ala Pro Pro Pro Ser Trp Asp Gln Met Trp Lys Cys Leu Ile			
1605	1610	1615	
Arg Leu Lys Pro Thr Leu His Gly Pro Thr Pro Leu Leu Tyr Arg Leu			
1620	1625	1630	
Gly Ala Val Gln Asn Glu Val Thr Leu Thr His Pro Ile Thr Lys Tyr			
1635	1640	1645	
Ile Met Thr Cys Met Ser Ala Asp Leu Glu Val Val Thr Ser Thr Trp			
1650	1655	1660	
Val Leu Val Gly Gly Val Leu Ala Ala Leu Ala Ala Tyr Cys Leu Ser			
1665	1670	1675	1680
Thr Gly Cys Val Val Ile Val Gly Arg Ile Val Leu Ser Gly Lys Pro			
1685	1690	1695	
Ala Ile Ile Pro Asp Arg Glu Val Leu Tyr Gln Glu Phe Asp Glu Met			
1700	1705	1710	
Glu Glu Cys Ser Gln His Leu Pro Tyr Ile Glu Gln Gly Met Met Leu			
1715	1720	1725	
Ala Glu Gln Phe Lys Gln Lys Ala Leu Gly Leu Leu Gln Thr Ala Ser			
1730	1735	1740	
Arg His Ala Glu Val Ile Thr Pro Ala Val Gln Thr Asn Trp Gln Lys			
1745	1750	1755	1760
Leu Glu Val Phe Trp Ala Lys His Met Trp Asn Phe Ile Ser Gly Ile			

1765	1770	1775
Gln Tyr Leu Ala Gly Leu Ser Thr Leu Pro Gly Asn Pro Ala Ile Ala		
1780	1785	1790
Ser Leu Met Ala Phe Thr Ala Ala Val Thr Ser Pro Leu Thr Thr Gly		
1795	1800	1805
Gln Thr Leu Leu Phe Asn Ile Leu Gly Gly Trp Val Ala Ala Gln Leu		
1810	1815	1820
Ala Ala Pro Gly Ala Ala Thr Ala Phe Val Gly Ala Gly Leu Ala Gly		
1825	1830	1835 1840
Ala Ala Ile Gly Ser Val Gly Leu Gly Lys Val Leu Val Asp Ile Leu		
1845	1850	1855
Ala Gly Tyr Gly Ala Gly Val Ala Gly Ala Leu Val Ala Phe Lys Ile		
1860	1865	1870
Met Ser Gly Glu Val Pro Ser Thr Glu Asp Leu Val Asn Leu Leu Pro		
1875	1880	1885
Ala Ile Leu Ser Pro Gly Ala Leu Val Val Gly Val Val Cys Ala Ala		
1890	1895	1900
Ile Leu Arg Arg His Val Gly Pro Gly Glu Gly Ala Val Gln Trp Met		
1905	1910	1915 1920
Asn Arg Leu Ile Ala Phe Ala Ser Arg Gly Asn His Val Ser Pro Thr		
1925	1930	1935
His Tyr Val Pro Glu Ser Asp Ala Ala Ala Arg Val Thr Ala Ile Leu		
1940	1945	1950
Ser Ser Leu Thr Val Thr Gln Leu Leu Arg Arg Leu His Gln Trp Ile		
1955	1960	1965
Ser Ser Glu Cys Thr Thr Pro Cys Ser Gly Ser Trp Leu Arg Asp Ile		
1970	1975	1980
Trp Asp Trp Ile Cys Glu Val Leu Ser Asp Phe Lys Thr Trp Leu Lys		
1985	1990	1995 2000
Ala Lys Leu Met Pro Gln Leu Pro Gly Ile Pro Phe Val Ser Cys Gln		
2005	2010	2015
Arg Gly Tyr Arg Gly Val Trp Arg Gly Asp Gly Ile Met His Thr Arg		

2020	2025	2030
Cys His Cys Gly Ala Glu Ile Thr Gly His Val Lys Asn Gly Thr Met 2035	2040	2045
Arg Ile Val Gly Pro Arg Thr Cys Arg Asn Met Trp Ser Gly Thr Phe 2050	2055	2060
Pro Ile Asn Ala Tyr Thr Thr Gly Pro Cys Thr Pro Leu Pro Ala Pro 2065	2070	2075 2080
Asn Tyr Lys Phe Ala Leu Trp Arg Val Ser Ala Glu Glu Tyr Val Glu 2085	2090	2095
Ile Arg Arg Val Gly Asp Phe His Tyr Val Ser Gly Met Thr Thr Asp 2100	2105	2110
Asn Leu Lys Cys Pro Cys Gln Ile Pro Ser Pro Glu Phe Phe Thr Glu 2115	2120	2125
Leu Asp Gly Val Arg Leu His Arg Phe Ala Pro Pro Cys Lys Pro Leu 2130	2135	2140
Leu Arg Glu Glu Val Ser Phe Arg Val Gly Leu His Glu Tyr Pro Val 2145	2150	2155 2160
Gly Ser Gln Leu Pro Cys Glu Pro Glu Pro Asp Val Ala Val Leu Thr 2165	2170	2175
Ser Met Leu Thr Asp Pro Ser His Ile Thr Ala Glu Ala Ala Gly Arg 2180	2185	2190
Arg Leu Ala Arg Gly Ser Pro Pro Ser Met Ala Ser Ser Ser Ala Ser 2195	2200	2205
Gln Leu Ser Ala Pro Ser Leu Lys Ala Thr Cys Thr Ala Asn His Asp 2210	2215	2220
Ser Pro Asp Ala Glu Leu Ile Glu Ala Asn Leu Leu Trp Arg Gln Glu 2225	2230	2235 2240
Met Gly Gly Asn Ile Thr Arg Val Glu Ser Glu Asn Lys Val Val Ile 2245	2250	2255
Leu Asp Ser Phe Asp Pro Leu Val Ala Glu Glu Asp Glu Arg Glu Val 2260	2265	2270
Ser Val Pro Ala Glu Ile Leu Arg Lys Ser Arg Arg Phe Ala Arg Ala		

2275	2280	2285
Leu Pro Val Trp Ala Arg Pro Asp Tyr Asn Pro Pro Leu Val Glu Thr		
2290	2295	2300
Trp Lys Lys Pro Asp Tyr Glu Pro Pro Val Val His Gly Cys Pro Leu		
2305	2310	2315 2320
Pro Pro Pro Arg Ser Pro Pro Val Pro Pro Pro Arg Lys Lys Arg Thr		
2325	2330	2335
Val Val Leu Thr Glu Ser Thr Leu Ser Thr Ala Leu Ala Glu Leu Ala		
2340	2345	2350
Thr Lys Ser Phe Gly Ser Ser Ser Thr Ser Gly Ile Thr Gly Asp Asn		
2355	2360	2365
Thr Thr Thr Ser Ser Glu Pro Ala Pro Ser Gly Cys Pro Pro Asp Ser		
2370	2375	2380
Asp Val Glu Ser Tyr Ser Ser Met Pro Pro Leu Glu Gly Glu Pro Gly		
2385	2390	2395 2400
Asp Pro Asp Leu Ser Asp Gly Ser Trp Ser Thr Val Ser Ser Gly Ala		
2405	2410	2415
Asp Thr Glu Asp Val Val Cys Cys Ser Met Ser Tyr Ser Trp Thr Gly		
2420	2425	2430
Ala Leu Val Thr Pro Cys Ala Ala Glu Glu Gln Lys Leu Pro Ile Asn		
2435	2440	2445
Ala Leu Ser Asn Ser Leu Leu Arg His His Asn Leu Val Tyr Ser Thr		
2450	2455	2460
Thr Ser Arg Ser Ala Cys Gln Arg Gln Lys Lys Val Thr Phe Asp Arg		
2465	2470	2475 2480
Leu Gln Val Leu Asp Ser His Tyr Gln Asp Val Leu Lys Glu Val Lys		
2485	2490	2495
Ala Ala Ala Ser Lys Val Lys Ala Asn Leu Leu Ser Val Glu Glu Ala		
2500	2505	2510
Cys Ser Leu Thr Pro Pro His Ser Ala Lys Ser Lys Phe Gly Tyr Gly		
2515	2520	2525
Ala Lys Asp Val Arg Cys His Ala Arg Lys Ala Val Ala His Ile Asn		

2530	2535	2540	
Ser Val Trp Lys Asp Leu Leu Glu Asp Ser Val Thr Pro Ile Asp Thr			
2545	2550	2555	2560
Thr Ile Met Ala Lys Asn Glu Val Phe Cys Val Gln Pro Glu Lys Gly			
2565	2570	2575	
Gly Arg Lys Pro Ala Arg Leu Ile Val Phe Pro Asp Leu Gly Val Arg			
2580	2585	2590	
Val Cys Glu Lys Met Ala Leu Tyr Asp Val Val Ser Lys Leu Pro Leu			
2595	2600	2605	
Ala Val Met Gly Ser Ser Tyr Gly Phe Gln Tyr Ser Pro Gly Gln Arg			
2610	2615	2620	
Val Glu Phe Leu Val Gln Ala Trp Lys Ser Lys Lys Thr Pro Met Gly			
2625	2630	2635	2640
Phe Ser Tyr Asp Thr Arg Cys Phe Asp Ser Thr Val Thr Glu Ser Asp			
2645	2650	2655	
Ile Arg Thr Glu Glu Ala Ile Tyr Gln Cys Cys Asp Leu Asp Pro Gln			
2660	2665	2670	
Ala Arg Val Ala Ile Lys Ser Leu Thr Glu Arg Leu Tyr Val Gly Gly			
2675	2680	2685	
Pro Leu Thr Asn Ser Arg Gly Glu Asn Cys Gly Tyr Arg Arg Cys Arg			
2690	2695	2700	
Ala Ser Gly Val Leu Thr Thr Ser Cys Gly Asn Thr Leu Thr Cys Tyr			
2705	2710	2715	2720
Ile Lys Ala Arg Ala Ala Cys Arg Ala Ala Gly Leu Gln Asp Cys Thr			
2725	2730	2735	
Met Leu Val Cys Gly Asp Asp Leu Val Val Ile Cys Glu Ser Ala Gly			
2740	2745	2750	
Val Gln Glu Asp Ala Ala Ser Leu Arg Ala Phe Thr Glu Ala Met Thr			
2755	2760	2765	
Arg Tyr Ser Ala Pro Pro Gly Asp Pro Pro Gln Pro Glu Tyr Asp Leu			
2770	2775	2780	
Glu Leu Ile Thr Ser Cys Ser Ser Asn Val Ser Val Ala His Asp Gly			

2785	2790	2795	2800
Ala Gly Lys Arg Val Tyr Tyr Leu Thr Arg Asp Pro Thr Thr Pro Leu			
	2805	2810	2815
Ala Arg Ala Ala Trp Glu Thr Ala Arg His Thr Pro Val Asn Ser Trp			
	2820	2825	2830
Leu Gly Asn Ile Ile Met Phe Ala Pro Thr Leu Trp Ala Arg Met Ile			
	2835	2840	2845
Leu Met Thr His Phe Phe Ser Val Leu Ile Ala Arg Asp Gln Leu Glu			
	2850	2855	2860
Gln Ala Leu Asn Cys Glu Ile Tyr Gly Ala Cys Tyr Ser Ile Glu Pro			
	2865	2870	2875
Leu Asp Leu Pro Pro Ile Ile Gln Arg Leu His Gly Leu Ser Ala Phe			
	2885	2890	2895
Ser Leu His Ser Tyr Ser Pro Gly Glu Ile Asn Arg Val Ala Ala Cys			
	2900	2905	2910
Leu Arg Lys Leu Gly Val Pro Pro Leu Arg Ala Trp Arg His Arg Ala			
	2915	2920	2925
Arg Ser Val Arg Ala Arg Leu Leu Ser Arg Gly Gly Arg Ala Ala Ile			
	2930	2935	2940
Cys Gly Lys Tyr Leu Phe Asn Trp Ala Val Arg Thr Lys Leu Lys Leu			
	2945	2950	2955
Thr Pro Ile Ala Ala Ala Gly Arg Leu Asp Leu Ser Gly Trp Phe Thr			
	2965	2970	2975
Ala Gly Tyr Ser Gly Gly Asp Ile Tyr His Ser Val Ser His Ala Arg			
	2980	2985	2990
Pro Arg Trp Phe Trp Phe Cys Leu Leu Leu Leu Ala Ala Gly Val Gly			
	2995	3000	3005
Ile Tyr Leu Leu Pro Asn Arg			
	3010	3015	

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<213> Hepatitis C virus

<400> 11

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24

<210> 12

<211> 24

<212> DNA

<213> Hepatitis C virus

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24

<210> 13

<211> 24

<212> DNA

<213> Hepatitis C virus

<400> 13

gggtgtacta cacacatgag taag

24

<210> 14

<211> 22

<212> DNA

<213> Hepatitis C virus

<400> 14

aagcgcccct aacttgatga tg

22

<210> 15

<211> 40

<212> DNA

<213> Hepatitis C virus

<400> 15

cgatcatgat acctcagcgg gcatatgcac tggacacgga

40

<210> 16

<211> 24

<212> DNA

<213> Hepatitis C virus

<400> 16

gtccagtga tatgcccgt gagg

24

<210> 17

<211> 32

<212> DNA

<213> Hepatitis C virus

<400> 17

catgcaccag ctgatatagc gcttgtaata tg

32

<210> 18

<211> 30

<212> DNA

<213> Hepatitis C virus

<400> 18

tccgtagagg aagcttgag cctgacgccc

30

<210> 19

<211> 34

<212> DNA

<213> Hepatitis C virus

<400> 19

cagaggaggc agggctgcta tatgtggcaa gtac

34

<210> 20

<211> 34

<212> DNA

<213> Hepatitis C virus

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<210> 21

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43

<210> 22

<211> 65

<212> DNA

<213> Hepatitis C virus

<400> 22

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aatgc 65

<210> 23

<211> 32

<212> DNA

<213> Hepatitis C virus

<400> 23

aggatttgtg ctcatgggtgc acggtctacg ag 32

<210> 24

<211> 50

<212> DNA

<213> Hepatitis C virus

<400> 24

ttttttttgc ggcgcctaatac gactcact atagaccgc ccctaataagg 50

<210> 25

<211> 31

<212> DNA

<213> Hepatitis C virus

<400> 25

ccgtgcacca tgagcacaaa tcctaaacct c 31

<210> 26

<211> 26

<212> DNA

<213> Hepatitis C virus

<400> 26

ggatgtaccc catgaggctg gcaaag 26

<210> 27

<211> 30

<212> DNA

<213> Hepatitis C virus

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gtttgcgcct gcttatggat gctcatcttg

30

<210> 28

<211> 26

<212> DNA

<213> Hepatitis C virus

<400> 28

gcgtcataag catatgcctg ttgggg

26

<210> 29

<211> 23

<212> DNA

<213> Hepatitis C virus

<400> 29

ccctcagcac tggagtacat ctg

23

<210> 30

<211> 39

<212> DNA

<213> Hepatitis C virus

<400> 30

cgtcatgcat acccctaggg cggctctcat tgaagaggg

39

<210> 31

<211> 30

<212> DNA

<213> Hepatitis C virus

<400> 31

cgtccctct tcaatgagag ccgctctaga

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<210> 32

<211> 28

<212> DNA

<213> Hepatitis C virus

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gcggtgaaga ccaagctcaa actcactc

28

<210> 33

<211> 41

<212> DNA

<213> Hepatitis C virus

<400> 33

aatctagaag gcgcgcttcc ggcaatggag tgagtttgag c

41

<210> 34

<211> 38

<212> DNA

<213> Hepatitis C virus

<400> 34

cgtctctaga ggataaatcc aggaggcgcg cttccggc

38

<210> 35

<211> 27

<212> DNA

<213> Hepatitis C virus

<400> 35

tactttttgt aggggtaggc cttttcc

27

<210> 36

<211> 34

<212> DNA

<213> Hepatitis C virus

<400> 36

cgtctctaga gtgtagctaa tgtgtgccgc tcta

34

<210> 37

<211> 24

<212> DNA

<213> Hepatitis C virus

<400> 37

ctatggagtg tagctaattgt gtgc

24

<210> 38

<211> 66

<212> DNA

<213> Hepatitis C virus

<400> 38

cgtctctaga catgatctgc agagagacca gttacggcac tctctgcagt catgcggctc 60
acggac 66

<210> 39

<211> 41

<212> DNA

<213> Hepatitis C virus

<400> 39

ctttcacagc tagccgtgac tagggctaag atggagccac c 41